

PRE-CONGRESS COURSE 2

SIG Embryology “The human IVF lab in 2008 and beyond”

CONTENTS

| | |
|---|-------|
| Program overview | p. 1 |
| Speakers' contributions | |
| • Quality in the IVF lab: how to do? - <i>P. Kastrop (NL)</i> | p. 2 |
| • Errors in the IVF lab: how not to do? - <i>S. Ziebe (DK)</i> | p. 13 |
| • European cell and tissue directives now implemented: What are the practical implications in the IVF lab? - <i>J. Van der Elst (B)</i> | p. 22 |
| • The importance of collection and registration of laboratory data. - <i>K. Erb (DK)</i> | p. 38 |
| • Automation of embryo selection and production - <i>J. Thompson (Aus)</i> | p. 49 |
| • Non-invasive embryo assessment: proteomic and metabolomic biomarkers - <i>P. Nagy (USA)</i> | p. 60 |
| • An update on embryo culture for human ART: media, performance and safety - <i>T. Pool (USA)</i> | p. 77 |
| • Safe and efficient vitrification methods for human oocytes: how to do it? - <i>L. Rienzi (I)</i> | p. 98 |

PRE-CONGRESS COURSE 2 - PROGRAMME

SIG Embryology

The human IVF lab in 2008 and beyond

Course co-ordinators: E. Van den Abbeel (B), K. Lundin (S), C. Magli (I), D. Royere (F)

Course description: This basic course aims at discussing (1) IVF laboratory safety and quality issues and (2) developments in embryology and IVF technology.

Target audience: Clinical embryologists and clinicians

Programme

| | |
|----------------------|---|
| 09.00 – 09.30 | Quality in the IVF lab: how to do? - P. Kastrop (NL) |
| 09.30 – 09.45 | <i>Discussion</i> |
| 09.45 – 10.15 | Errors in the IVF lab: how not to do? - S. Ziebe (DK) |
| 10.15 – 10.30 | <i>Discussion</i> |
| 10.30 – 11.00 | Coffee break |
| 11.00 – 11.30 | European cell and tissue directives now implemented: What are the practical implications in the IVF lab? - J. Van der Elst (B) |
| 11.30 – 11.45 | <i>Discussion</i> |
| 11.45 – 12.15 | The importance of collection and registration of laboratory data. - K. Erb (DK) |
| 12.15 – 12.30 | <i>Discussion</i> |
| 12.30 – 13.30 | Lunch |
| 13.30 – 14.00 | Automation of embryo selection and production - J. Thompson (Aus) |
| 14.00 – 14.15 | <i>Discussion</i> |
| 14.15 – 14.45 | Non-invasive embryo assessment: proteomic and metabolomic biomarkers - P. Nagy (USA) |
| 14.45 – 15.00 | <i>Discussion</i> |
| 15.00 – 15.30 | Coffee break |
| 15.30 – 16.00 | An update on embryo culture for human ART: media, performance and safety. - T. Pool (USA) |
| 16.00 – 16.15 | <i>Discussion</i> |
| 16.15 – 16.45 | Safe and efficient vitrification methods for human oocytes: how to do it? - L. Rienzi (I) |
| 16.45 – 17.00 | <i>Discussion</i> |
| 17.00 – 18.00 | Special Interest Group Embryology Business meeting |

Quality in the IVF lab: how to do?

Peter Kastrop, Ph.D.
Clinical embryologist

University Medical Center Utrecht

ESHRE Barcelona 6 July 2008

Content

- Quality
- Quality management
- Performance indicators
- Standards / Guidelines
- How to do

Quality

Invisible when GOOD, impossible to ignore when BAD

People forget how fast you did a job but they remember how well you did it

Howard W Newton

Quality is never an accident; it is always the result of intelligent efforts

John Ruskin

Quality is never an accident; it is always the result of high intention, sincere effort, intelligent direction and skilful execution; it represents the wise choice of many alternatives

William A. Foster

Quality

ISO definition:

the total sum of properties and characteristics of a product, process, or service which is vital in order to meet the requirements as determined or assumed needs

expectation / trust / satisfaction

Quality in the IVF laboratory

- product / service
- end product / endpoint
- measurable
- manageable
- expectations of the patients

Quality in the IVF laboratory

- highest level of patients care
- highest success rates

Quality management

- **Quality Control (QC):**

the operational techniques and activities which are carried out in order to meet the quality requirements

- **Quality Assurance (QA):**

the total sum of all planned and systematic activities required in order to establish sufficient trust that a product or service meets the quality requirements as determined

ISO definitions

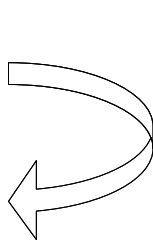
- **Quality Improvement**

Quality Improvement

- Risk management
- Quality assessments
- Problem management

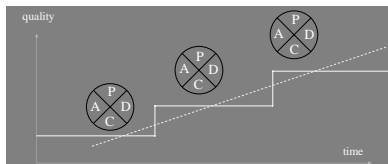
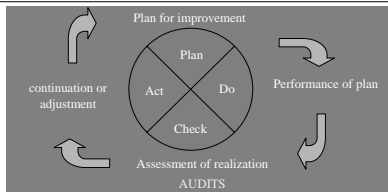
PDCA cycle

Plan – Do – Check- Act



Quality Improvement

PDCA cycle



Risk management

- **Risk analysis**
- What are critical steps?
- **Corrective activities**
- What to do when something went wrong?
- **Preventive activities:**
- What to do to prevent that something goes wrong?

Risk management

Critical steps:

- all actions in which samples are transferred from one dish or tube to another
- all moments where samples of different origin come close to each other
- all transfers of samples to or from physicians or patients

Quality assessments

- Internal audit
- External audit
- Management Review
- Inter professional assessment

Quality assessments

Internal audit

- Quality manager and/or qualified personnel
- Defined and documented standard procedure
- Periodically and systematically
- Non-conformities (deficiencies / deviations)
- Preventive and corrective measures
- Inter / External quality assurance programs

Quality assessments

External audit

- Independent external body
- National / International standard
- Periodically and systematically
- Non-conformities (deficiencies / deviations)
- Preventive and corrective measures
- Internal / External quality assurance programs

> Formal recognition (certification / accreditation)

Quality assessments

Management Review

- Management
- Annually
- Suitability and effectiveness QMS
- Outcome of internal / external assessments
- Non-conformities and complaints
- Preventive and corrective measures
- Performance measures / quality indicators
- Patient satisfaction assessments
- Continuous improvement results

Quality assessments

Inter Professional assessment

- Independent (external) colleagues
- Defined and documented regulation
- Periodically
- Professional competence

Problem management

Problem \iff Solve / Correct / Prevent

- Accident
- Complaint
- Defect
- Deficiency
- Deviation
- Error
- Failure
- Incident
- Infection
- Mistake
- Non-conformity

Problem management

- To minimise the number and severity
- To reduce the adverse impact
- To improve quality

Premise: What can we learn from it?

NOT: Who is to blame?

Performance indicators

- Operational
 - Equipment
 - Personnel
 - Methods (IQC and EQA programmes)
- Outcome
 - Methods
 - Programme
 - “Problems”
- Financial

Performance indicators

IVF / ICSI / PGD / IUI / Cryopreservation (embryos / sperm)

Methods

- Fertilisation rate (2PN, 1PN, 3PN, degen.)
- Damage rate
- Cleavage rate
- Fragmentation rate
- Embryo scoring rate
- Embryo survival rate
- Sperm recovery rate
-

Programme

- Oocyte scoring rate
- Implantation rate
- Biochemical pregnancy rate
- Clinical pregnancy rate
- Multiple rate
- Abortion rate
- Live births
-

Standards / Guidelines

- by professional societies
ESHRE, ACE, AFS
- (Inter)national standards
ISO, ACHS, CCHSA, CPA, CCKL
- Quality management models
TQM, EFQM

Standards / Guidelines

- Guidelines for human embryology and andrology laboratories
 - The American Fertility Society , 1992
- Guidelines for good practice in IVF laboratories
 - ESHRE 2000 / 2008?
 - <http://www.eshre.com>
- Accreditation standards and guidelines for IVF laboratories
 - Association of Clinical Embryologists, 1999
 - http://www.embryologists.org.uk/downloads/ACCREDITATION_STANDARDS_AND_GUIDELINES.doc

Standards / Guidelines

- ISO 9001: (2000)
Quality management systems – Requirements
- ISO 17025: (1999)
General requirements for the competence of testing and calibration laboratories
- ISO 15189: (2007)
Medical laboratories – Particular requirements for quality and competence

Standards / Guidelines

- Total Quality Management (TQM)
- EFQM Excellence Model
(European Foundation for Quality Management)
- Strategic way of quality management (philosophy)
- embedded within whole organization
- total commitment of entire organization
- strive of continuous improvement
- continuous scrutiny of all components of the quality system
- assessments and audits (periodically and systematically)

How to do

Quality Control

- detailed written standard procedures
- procedure, safety and policy manuals
- appropriately educated and trained personnel
- correct operation and calibration of instruments
- consistent and proper execution of appropriate techniques and methods
- documentation and record keeping
- system for patient sample collection and management
- system for the appraisal of performance and correction of deficiencies

How to do

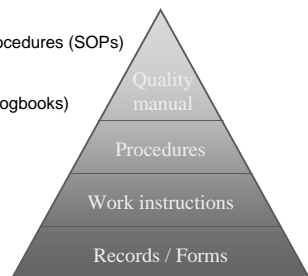
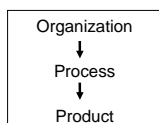
Quality Assurance

- system for unambiguous patient and patient sample identification
- ongoing method of assessing staff competency in terms of their clinical and clerical skills
- monitoring and evaluation of number and type of accidents, mistakes and deviations
- system for addressing and documenting complaints
- system for the implementation of advances and improvements
- application of apparent laboratory performance indicators

How to do

Documentation

- Quality manual
- Detailed written standard procedures (SOPs)
- Working instructions
- Laboratory forms / records (logbooks)



How to do

Additional documents

- Job descriptions and curricula vitae
- Training programmes
- (Annual) reports
- Management review

Document control System

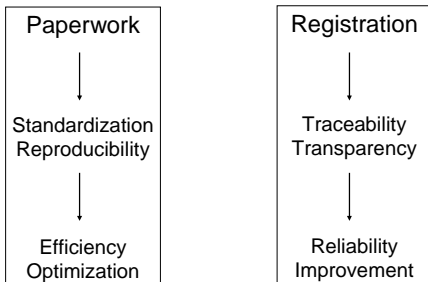
How to do

Requirements

In daily practice:

- separated working places
- recording of every action performed
- verification of critical steps by a second person
- at least 2 persons in the weekend
- minimize unrest
- prevent diminished concentration

How to do



How to do



How to do

Principle:

*Say what you do
do what you say
and
show that you do as you say*

Errors in the IVF lab:
How not to do?
Søren Ziebe



Errors in the IVF lab:
How not to do!
Søren Ziebe



– Examples from real life....

Errors in the IVF lab:
How not to do – anymore!
Søren Ziebe



– Examples from real life....

Errors in the IVF lab:

What not to accept – anymore!

Søren Ziebe



– Examples from real life....

Errors in the IVF lab:

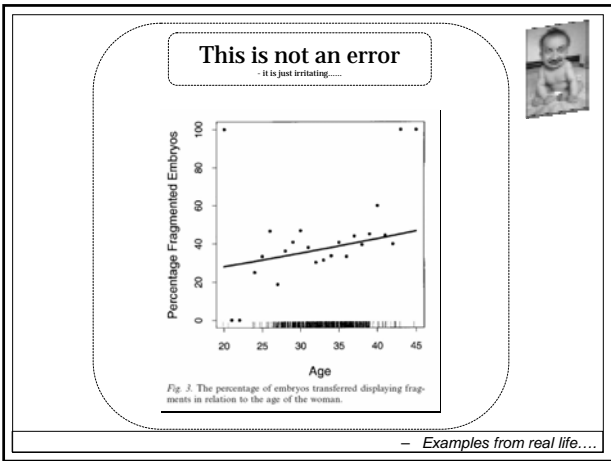
How not to do?

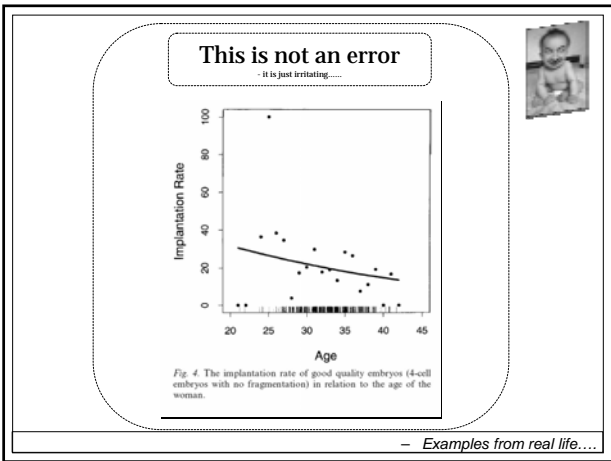
Søren Ziebe

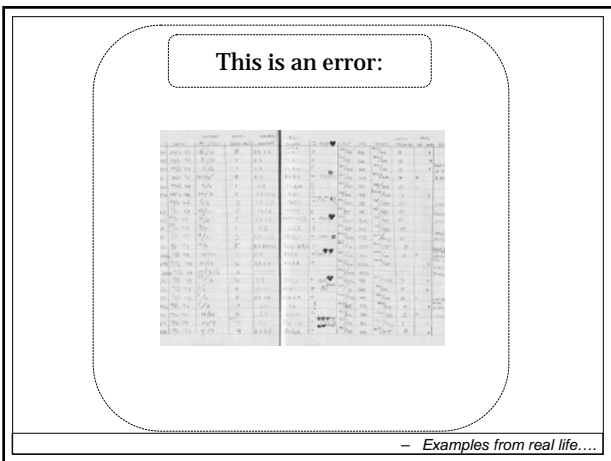


What is an error?

Don't mix up biological variation and errors just because money and emotions is involved!





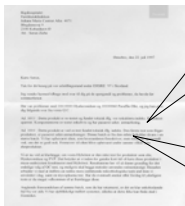


This is an error:



- Examples from real life....

This is an error:



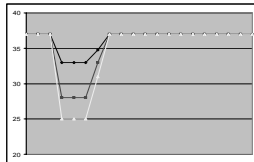
The Hyaluronidase:
"This product has been re-tested and was found **toxic** according to our toxicity index...."

The ICSI Oil:
"This product has been re-tested and was found **toxic** according to our toxicity index...."

- Examples from real life....

The gray zone:

Spindles disassembled after cooling

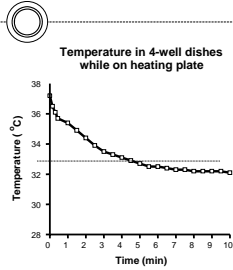
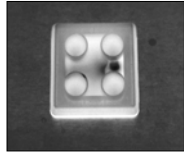


| Temperature | Re-assembly |
|-------------|-------------|
| 33 degrees | 100% |
| 28 degrees | 40% |
| 25 degrees | 0% |

Spindle re-assembly when oocytes were cooled and kept at 33, 28 or 25°C for 10 min

Wang et al. Human Reproduction, Vol. 16, No. 11, 2374-2378, 2001

The gray zone:



Ziebe, Unpublished

Communication

In the old days:
A professional life isolated in the laboratory



Modern ART treatment:
The laboratory are in close contact with the patients



- Examples from real life....

Communication to the patients



"Remember - if the sperm sample is more than **60.29** minutes old you will never have a baby !!!"


- Examples from real life....

Consequence....

This car was stopped by the police due to strange driving.....

The couple in the car were trying to produce a sperm sample while driving to the fertility clinic.....

Fik taget sædprøve under bilkørsel



Politiet troede, der var tale om promillekørsel, da de så en bil slingre lidt fra side til side. Men det var det ikke. Slet ikke!

Da politibilen kom nærmere kunne betjentene konstatere, at der sad en mand bag rattet, og ved siden af en kvinde, der åbenbart havde travlt med gearstangen. Men det var nu ikke gearet, hun skiftede. Det var nærmest i vejen. For da politibetjente lidt udenfor Førde i Norge, fik stoppet bilen gjorde de.....store øjne.

Kvinden i vognen havde såmænd taget en sædprøve på manden, hvilket var årsag til hans slingre kørsel. Begge var adru.

Og forklaringen: Ægteparret, for det var det, var på vej til Førde Centralsygehus, skriver VG Nett, for at aflevere en dugfrisk sædprøve. Og hvad er mere frisk end noget, der netop er leveret ned i glasset.

Polititalkene kunne ved selvsyn konstatere, at ægteparret talte sandt, og parret slap for en bøde, men fik dog at vide, at fremover, hvis det blev aktuelt igen, skulle de stoppe bilen, før de tog prøven.

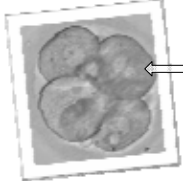
hajer@et.dk

Lagt på www.bt.dk den 30. marts 2006 kl. 17:30

— Examples from real life....

You need to know your biology!

When we are getting desperate....



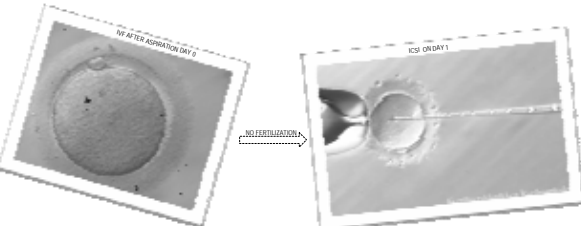
This is **not** a cell who has duplicated its DNA as part of the cleavage process

— Examples from real life....

You need to know your biology!

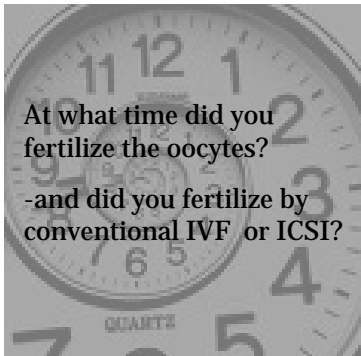
When we are getting desperate....

Rescue ICSI



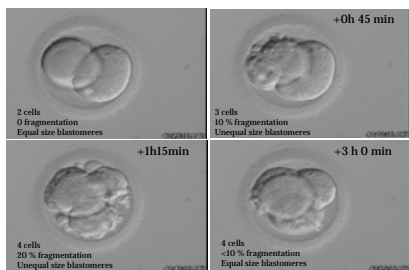
— Examples from real life....

Don't disregard timing



- Examples from real life....

Don't disregard timing



- Examples from real life....

Don't be too focused on details – forgetting the big picture

I wonder if this pen is embryo toxic???



- Examples from real life....

The laboratory is intimately integrated with the rest of the fertility clinic:

A poor laboratory will negatively impact outcome even after an optimal clinical performance

Not even the best laboratory can compensate for a poor clinical performance

An error in the laboratory can therefore be mediated, initiated or closely integrated with the rest of the clinic

Consequence:

An embryologist should choose his/her clinical staff very carefully.....

**European Cell and Tissue directives now implemented:
What are the practical implications in the IVF lab?**

Prof. Josiane Van der Elst, Ph.D.

EACC

Universitair Ziekenhuis Brussel
Vrije Universiteit Brussel

Centrum voor Reproductieve Geneeskunde

ESHRE 2008 PCC Embryology

Disclosure

There are no commercial relationships or other activities that might be perceived as a potential conflict of interest

Universitair Ziekenhuis Brussel
Vrije Universiteit Brussel

Centrum voor Reproductieve Geneeskunde

2

Learning objectives

- To learn about the origin of the European Cell and Tissue Directives (EUTCD)
- To learn and understand the goal of the EUTCD
- To learn and understand the impact of implementation of EUTCD on IVF laboratory operation
- To learn about communication channels on EUTCD

Universitair Ziekenhuis Brussel
Vrije Universiteit Brussel

Centrum voor Reproductieve Geneeskunde


3

Outline

- EU and EU Cell and Tissue Directives
- Implementation
- Impact for IVF laboratories
- Differences within Europe
- Channels of communication

4

European Union



- The European Union was founded in 1957
- Political treaties define cooperation between Member States
- Treaties result in European Directives
- EU Directives are made by European Institutions
 - Council of Ministers
 - European Parliament
 - European Commission

5

European Treaties and public health

- Treaty of Rome (1957) (EEC)
 - economic cooperation, no public health issues
- Treaty of Maastricht (1992) (EU)
 - cooperation on public health: information, education
- Treaty of Amsterdam (1999)
 - cooperation on public health: protection
 - provides legal tools to ensure health protection

6

Directive 2004 / 23 / EC

- The European Union Tissues and Cells Directive (EUTCD) is a legal document originating from the European Union's public health programme
- standards of quality and safety for human tissues and cells intended for human applications, in order to ensure a high level of protection of human health
- particularly in order to prevent the transmission of diseases



7

The mother and technical directives

- 2004/23/EC of European Parliament and of the Council of 31 March 2004
aka Mother Directive
standards of quality and safety for application of human tissues and cells to the human body
Into force in EU on 7 April 2006
- 2006/17/EC Commission Directive of 8 February 2006
aka Technical Directive 1, EUTD1
donor centered
donation, procurement, testing
Into force in EU on 1 November 2006
- 2007/86/EC Commission Directive of 24 October 2006
aka Technical Directive 2, EUTD2
cell- and -tissue centered
coding, processing, preservation, storage and distribution
Into force in EU on 1 September 2007



8

Outline


- EU and EU Cell and Tissue Directives
- Implementation
- Impact for IVF laboratories
- Differences within Europe
- Channels of communication



9

Implementation of EUTCD

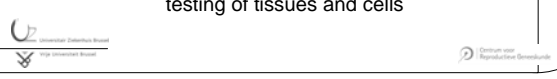
- Obligations for Member States
 - transpose EUTCD in national legislation
 - install a Competent Authority
 - set up licensing system for tissue establishments
 - organize inspections
 - set up a system for notification of serious adverse reactions/events
- report back to European Commission



10

Implementation of EUTCD

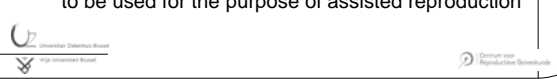
- Obligations for tissue establishments
 - 'tissue establishment'
 - a tissue bank or a unit of a hospital or another body where activities of processing, preservation, storage or distribution of human tissues and cells are undertaken.
 - may also be responsible for procurement or testing of tissues and cells



11

Implementation of EUTCD

- This Directive should apply to tissues and cells including haematopoietic peripheral blood, umbilical-cord (blood) and bone-marrow stem cells, reproductive cells (eggs, sperm), foetal tissues and cells and adult and embryonic stem cells.
- 'reproductive cells' means all tissues and cells intended to be used for the purpose of assisted reproduction



12

Implementation of EUTCD

- IVF centres are considered as tissue establishments
- Have to fulfill safety and quality criteria
 - for cell donation, procurement and testing (TD1)
 - for coding, processing, preservation, storage and distribution of human tissues and cells (TD2)



13



Implementation of EUTCD

- Reproductive cells have, due to the specific nature of their application, specific quality and safety characteristics
- 'partner donation' means the donation of reproductive cells between a man and a woman who declare that they have an intimate physical relationship;
- 'direct use' means any procedure where cells are donated and used without any banking;



14



Requirements of TD1: donor-centered

Conditions for donation, procurement

- voluntary donation
- unpaid donation
- informed consent
- unique donor identification
- safe procurement: use of CE - labelled medical devices, wherever possible



15





Requirements of TD1: donor-centered

Conditions for laboratory testing

- infectious serological testing for partner donation
 - HIV1,2, HBV (sAG and core antibodies), HCV
- infectious serological testing non-partner donation
 - HIV1,2, HBV (sAG and core antibodies), HCV
 - Syphilis
 - Chlamydia

→ In case of partner donation and direct use laboratory testing may not be necessary






16

**Requirements of TD 2:
Cell - and - tissue centered**

- Quality Management System
- Air quality
- Frozen storage
- Critical materials
- Traceability
- Coding

- Notification of adverse reactions and events






17

Outline

- EU and EU Cell and Tissue Directives
- Implementation
- Impact for IVF laboratories
- Differences within Europe


- Channels of communication

18

Quality Management System


- There must be a documented quality management system applied to the activities for which accreditation/designation/authorisation or licensing is sought, in accordance with the standards laid down in this Directive
- ISO 9001:2000, ISO 15189 are examples



19

Quality Management System


- The personnel in tissue establishments
 - sufficient number
 - qualified for the tasks they perform
 - competency must be evaluated at appropriate intervals
- Opportunity: a new ESHRE initiative is the establishment of a certification system for clinical and senior clinical embryologists. The system aims at certifying both practical and theoretical competence of the laboratory staff



20

Air quality

- where tissues or cells are exposed to the environment during processing
- an air quality with particle counts and microbial colony counts equivalent to those of Grade A as defined in the current European Guide to Good Manufacturing Practice (GMP), Annex 1 and Directive 2003/94/EC is required
- with a background environment at least equivalent to GMP Grade D in terms of particles and microbial counts



21

Air quality

A less stringent environment may be acceptable where

- A validated microbiological inactivation or terminal sterilisation process is applied
- where it is demonstrated that exposure in a Grade A environment has a detrimental effect on the required properties of the tissue or cell concerned;

International Standards Board High Intensity System Centrum voor Reproductieve Geneeskunde

22

Air quality

A less stringent environment may be acceptable where

- It is demonstrated that the mode and route of application to the body implies a significantly lower risk of transmitting bacterial or fungal infection to the recipient than with cell and tissue transplantation
- where it is not technically possible to carry out the required process in a Grade A environment (for example, due to requirements for specific equipment in the processing area that is not fully compatible with Grade A)

International Standards Board High Intensity System Centrum voor Reproductieve Geneeskunde

23

Air quality

- but, even in case exceptions are allowed
- an environment must be specified
- it must be demonstrated and documented that the chosen environment achieves the quality and safety required

International Standards Board High Intensity System Centrum voor Reproductieve Geneeskunde

24

Frozen storage

- a system of separate storage must be devised
 - where HIV 1,2, hepatitis B or hepatitis C test results are positive or unavailable,
 - or where the donor is known to be a source of infection risk
- storage facilities must be provided that clearly separate
 - tissues and cells prior to release/in quarantine
 - from those that are released
 - from those that are rejected
 - in order to prevent mix-up and cross-contamination between them



25

Traceability

- Member States shall ensure that all tissues and cells procured, processed, stored or distributed on their territory can be traced from the donor to the recipient and vice versa
- Tissue establishments shall keep the data necessary to ensure traceability at all stages
- Data required for full traceability shall be kept for a minimum of 30 years after clinical use
- Data storage may also be in electronic form



26

Critical materials


- The traceability requirements for tissues and cells, as well as for products and materials coming into contact with these tissues and cells and having an effect on their quality and safety,
- Procurement procedures for tissues and cells
 - Wherever possible, only CE marked medical devices must be used
- Culture media: CE label?
 - Cf. France, AFSSAPS: culture media = produits thérapeutiques



27

Coding


- A single European identifying code shall be allocated to all donated material at the tissue establishment
 - to ensure proper identification of the donor and
 - the traceability of all donated material and
 - to provide information on the main characteristics and properties of tissues and cells
- shall not apply to partner donation of reproductive cells



28

Coding

- EC has the duty to develop a unique European code for cells and tissues
- EC ordered a CEN workshop to propose a code
- CEN = European Committee for Standardisation (Comité Européen de Normalisation)
- Three coding systems were compared
 - Italian coding system
 - Spanish coding system
 - ISBT 128




29

Notification adverse reactions

TD2- Annex V - Annual notification format - part A

- Transmitted bacterial infection
- Transmitted viral infection
- Transmitted parasitical infection
- Transmitted malignant diseases
- Other disease transmissions
- Other serious reactions



30

Notification adverse events

TD2 - Annex V - Annual notification format -part B

- Adverse events which may have affected quality and safety of tissues and cells due to
 - Tissues and cells defect
 - Equipment failure
 - Human error
 - Other

Outline



- EU and EU Cell and Tissue Directives
- Implementation
- Impact for IVF laboratories
- Differences within Europe
- Channels of communication

EU today



Implementation - questions asked



- Has implementation been done?
- Air quality required?
- Serology “at the time of donation”?

34

Ireland (source Tim Dineen)



- Mother directive and technical directives passed into law
- Grade D air quality according to GMP standard, i.e. with the need for pressurized rooms, air-locks etc. Fertility clinics disputing the need for GMP requirements, suggesting that Grade D in terms of microbial monitoring should be sufficient

35

Ireland (source Tim Dineen)

- For IVF/ICSI there will be a baseline serology done
 - when the couple first present to the clinic,
 - and then a serology will be done within 30 days of an egg collection
 - To be done for all egg collections
- For IUI, again all couples will need to have a serology done when they first present to the clinic, and they will subsequently need testing every 6 months

36

Denmark (source Soren Ziebe)

- EUTCD has been implemented
- no specification on air quality except that IVF does not call for the stringent quality
- Serology: at the time of donation: It is stated that the test should be done "prior" to aspiration. In case of oocyte donation it should be done no more than 30 day before.

Further, the test is valid for 24 month



37

Norway (source Arne Sunde)

- EUTCD has been implemented
 - no specific demand concerning air quality in ART
 - Serology: Partner donation – testing
- ART is considered as ONE treatment comprised of a series of interventions. This means that we only need to do serological testing every 12 months and not every time we collect oocytes/semen



38

Norway (source Arne Sunde)

- Direct use is defined as a procedure where there is no "storage". Likely interpretation is that processing of semen prior to IUI is not defined as "storage"
- Major changes in procedures/consumables/equipment must be approved by the authorities prior to implementation in the clinic. As an example, change of culture media is defined as a major change



39

Various countries, to be extended

- No implementation in national law so far in Greece, Belgium...
- Only the mother directive (2004/23/EC) has been transposed in Italy
- In Spain the EUTD has been transposed long time ago and the implementation at the level of tissues and cells other than reproductive cells is being undertaken by the national organisation of transplants, together with the Spanish drug agency

No competent authority has been designed for reproductive cells



40

Outline

- EU and EU Cell and Tissue Directives
- Implementation
- Impact for IVF laboratories
- Differences within Europe
- Channels of communication



41

Channels of communication

- ESHRE's Focus on Reproduction

ESHRE position paper on EUTCD in Focus
on Reproduction, January 2008

- ESHRE's EACC



42

EACC: an ESHRE offer you can't refuse

- European Assisted Conception Consortium
- Joint venture between ESHRE and HFEA
- Member state organisation
- Not - for - profit initiative supported by ESHRE
- Established at ESHRE 2005 Copenhagen
- Two annual meetings

→ one at the Annual ESHRE meeting



43

EACC Objectives

- Bring together regulators and IVF professionals from member states
- Communication between member states
- Communication to European Commission
 - seek how can we work together with the Commission to support implementation of the Directive
 - present joint position of regulators and practitioners
 - give expert advice to EC



44

EACC Format: Members

All EU member states

Per member state / three members

- Two practitioners
 - one clinician
 - one embryologist
- One regulator

Please check ESHRE Website – link to EACC - membership:
check for members of your country
they are your representatives

Non-EU members allowed to join for information



45

EACC Format: Executive Committee

→ Five practitioners

- Anna Veiga (Spain)
- Ioannis Messinis (Greece)
- Josiane Van der Elst (Belgium)
- Cristina Magli (Italy)
- Arne Sunde (Norway)

→ Three regulators

- Angela McNab (chair) (UK)
- Bernard Loty (Fr)
- Basil Tarlatzis (Greece)

46

References

- Link to the directives:
http://europa.eu.int/comm/health/ph_threats/human_substance/tissues_en.htm
- Link to EACC:
<http://www.eshre.com/emc.asp?pagelD=678>

47

THE IMPORTANCE OF COLLECTION AND REGISTRATION OF LABORATORY DATA

KARIN ERB
ODENSE UNIVERSITY HOSPITAL
DENMARK

Eshre Barcelona 2008

- WHY** collect and register laboratory data?
- WHAT** should we collect?
- HOW** to do it?

WHY

- Most large registers and studies focus on:
 - Number of started cycles
and
 - Pregnancy rates

Everything in between is usually not taken into account - such as the lab processes

Why - Purpose

- Control and administer cycles
- Follow-up and monitor own performance
- Generate data for stakeholders
- Compare key monitoring data with other clinics

Success indicators in IVF lab

Fertilisation rates
Development variables ("GQE")
Survival cryopreservation
Implantation
Live birth

WHY collect

- Different results from different laboratories
- Different methods/media/setup used

 **■ Larger data cohorts needed**

Why the differences between studies?

- Patient populations / group sizes
- Embryo / blastocyst morphology and development
- Culture conditions / blastocyst development rates

WHY collect

- Collecting data ensures large-scale information on the influence of:
 - Different aspects of oocyte and embryo morphology
 - Different culture conditions
 - Timing

WHAT to collect

- Oocyte info (stimulation regimes)
- Fertilisation info
(IVF/ICSI, short time incubation - O/N)
- Developmental rates
- Embryo morphology (GQE)
- Implantation rates

WHAT to collect

- Media info (simple, sequential)
- Culture conditions (Oxygen pressure)
- Screening; timing info
- ET timing (day 1,2,3,4,5,6,7?)
- Freezing / thawing (criteria / survival)

Variables for embryo selection

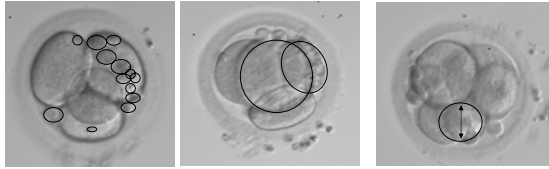
- Oocyte assessments
- Zygote scoring
- Cleavage rates
- Morphology (fragmentation, cell size)
- Number of nuclei
- Metabolic / genetic status?

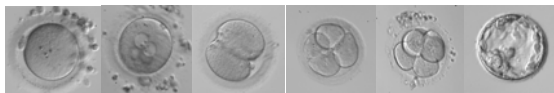
HOW to collect

- We need a common system
- Oocyte morphology
- Embryo / blastocyst morphology
- Sperm morphology (?)

- What is a good quality / top quality embryo?

Are we scoring the same things?





**How do we score
"good quality embryo"?**

Consensus?
Validation?

Common grading / scoring
system?

INTEROBSERVER AND INTRAOBSERVER
VARIATION IN DAY 3 EMBRYO GRADING

BAXTER AB, MAYER JF, SHIPLEY SK AND CATHERINO WH
FERTIL STERIL 2006; 86: 1608-1615

Design, Baxter et al.

- 26 embryologists at ASRM in Philadelphia
- 35 embryo videos recorded
(interobserver variation)
- 7 embryos shown several times
(intraobserver variation)
- Scale with 5 embryo grades (Veeck)
- Kappa values used for statistics

Results, Baxter et al.

- Interobserver variability (median, range)
Kappa 0.24 (0.03-0.49) poor

- Intraobserver variability (median, range)
Kappa 0.69 (0.44-1.00) good

Conclusions, Baxter et al.

We don't always agree

- Only use one embryologist for scoring?
- Use consensus scoring from several embryologists?
- Simplify the scoring system?

INTEROBSERVER AGREEMENT AND INTRAOBSERVER REPRODUCIBILITY OF EMBRYO QUALITY ASSESSMENTS

ARCE JC, ZIEBE, S, LUNDIN K, JANSSENS R, HELMGAARD L
AND SØRENSEN P. HUM REP 2006; 21: 2141-2148

Level of agreement – Top Quality Embryos

| | Kappa |
|-------------------------------|-------|
| Agreement among embryologists | |
| Central 1 vs Central 2 | 0.74 |
| Central 1 vs Central 3 | 0.74 |
| Central 2 vs Central 3 | 0.72 |
| Consolidated Central vs Local | 0.65 |

Reproducibility (intraobserver)

| | 28h | 44h | 68h |
|----------------------------|-----------|-----------|------------------|
| Cleavage stage | 0.97-0.98 | 0.85-0.90 | 0.72-0.83 |
| Blastomere uniformity | NA | 0.69-0.80 | 0.63-0.81 |
| Degree of fragmentation | NA | 0.64-0.77 | 0.68-0.72 |
| Multinucleation | NA | 0.53-0.88 | 0.48-0.66 |
| Cytoplasmic appearance | NA | 0.24-0.79 | 0.65-0.74 |
| Top quality embryos | | | 0.80-0.81 |

Conclusions

- Embryo quality assessments can be associated with high interobserver and intraobserver agreements
- Competence, accuracy and consistency can and should be acquired through continuous training and validation
- For large multicenter studies, a combination of local and central evaluations should provide the most effective and reliable approach to determine embryo quality

"EMBRYO REGISTRATION"

Inge Agerholm¹, Karin Erb², Marie Louise Grøndahl³, Søren Ziebe⁴

Fertilitetsklinikkerne

- ¹Brødstrup Sygehus
- ²Odense Universitetshospital
- ³Hvidovre Universitetshospital
- ⁴Rigshospitalet

| | |
|--|--|
| | |
|--|--|



Standardisation

-We need to use the same system

Definitions

- We must use the same definitions for the various parameters

The Danish (Nordic?) system

Register the number of blastomers

Register the degree of fragmentation

The Danish (Nordic?) system

Localisation of fragments:

- Score A1 Locally fragmented
- Score A2 Dispersed fragment

Blastomere size:

- Score B1 Equally sized blastomeres
- Score B2 Unequal sized blastomeres

Cytoplasm:

- Score C1 Homogenous cytoplasm
- Score C2 Unhomogenous/Granulated/Vacuolated

The Danish (Nordic?) system

Number of nuclei:

Score D1 No multi-nucleate blastomers present

Score D2 Multi-nucleate blastomers present

Early cleavage

Score E1 Early cleavage

Score E2 No Early cleavage

Zona Pellucida variation

Score Z1 Zona variation

Score Z2 No zona variation

And so on ...

The future – the dream

Common database

- Using the same registration system

Purpose

- Collecting data for national and international reports

- Webservice based data collection

The future – the dream

Stakeholders

- Your own laboratory data
- National Fertility Society
- Eshre – EIM

Authorities

- National Medical Agency (EU directive)
- National Board of Health

- Learning center – improving quality

Training and validation

Internal / external

- Scoring together
- Validation with images / videos

External, e.g. QAP online

- Database with pictures (videos coming?)
- Educational and quality control
- Scoring and comparison with all others or only own clinic
- Questions, such as "would you transfer/ freeze/ culture to blastocyst...", or "rank from best to worst"

Today's lesson

We have to collect and register laboratory data

Control and administer cycles

Follow-up and monitor our performance

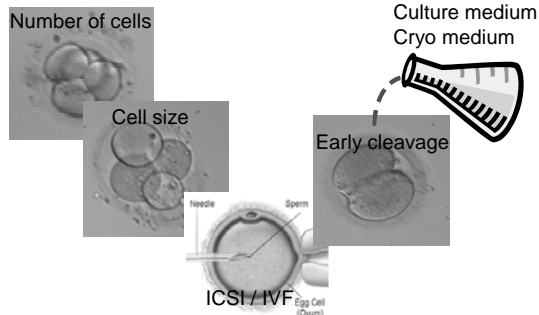
- ***you must know your own system***

Generate data to stakeholders

Compare key monitoring data with other clinics

Goal: To improve outcome

Summary



Automation of embryo selection and production

Assoc Prof Jeremy G Thompson BSc(Hon)
PhD

Research Centre for Reproductive Health
School of Paediatrics and Reproductive Health
The University of Adelaide
Adelaide 5005

jeremy.thompson@adelaide.edu.au

Jeremy G Thompson has obtained and currently holds research funding and has consultancy agreements with William A Cook Australia Pty Ltd

Learning objectives

- To define what “automation” in the IVF lab means
- To examine the impact of automation within an IVF lab
- To examine the development and application of technologies for embryo selection
- To examine the development of new embryo production systems

What is meant by “automation”

- Automating activities in the IVF lab that are currently performed by “hand”
 - Removing the “human factor” in a process
- Natural evolution for a technology that is robust and routinely practised

Benefits of automation

- Reliability
 - Does not become distracted
 - Parameters same every time
- Quality
 - Reduced variability
 - Measurable parameters checked
- Improved performance
 - Must be an improvement in performance for automation to be seen as attractive

Will I still have a job?!

- Probable consequence is reducing the need for staff with a *wide range* of skills
- Will allow for increased specialisation
- Will allow more rapid adoption of new techniques that require new training and development

Automation systems

- Analysis of embryos and embryo quality

- Embryo production systems

Automation of embryo quality assessment

Automated visual analysis of embryos

- Adds to or even replaces the subjective visual analysis
- Able to measure the kinetics of embryo development
 - Time lapse videomicroscopy
 - Digital analysis
- Digital imaging and analysis of cell morphology

Digital analysis of embryos

Embryo-guard system
Embryo-guard (www.gvmt.com)

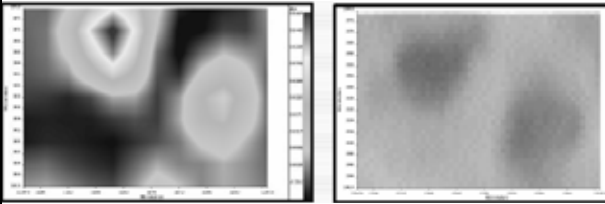


Digital analysis

- Light-penetration
 - Analysis of grey-scale pixelation
 - Analysis of light polarisation
 - Analysis of Infrared spectrum
 - Density & localisation of organelles
- Morphological features
 - Fragmentation
 - Cell number and allocation

New types of microscopy developing

Near Infrared (left) microscopy image of tumour cells (right) – chemical profiling

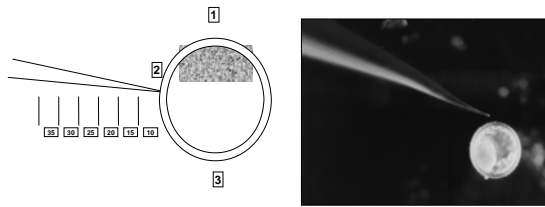


Baianu et al. <http://fs512.fshn.uiuc.edu/Soy2002-Imaging-2.pdf>

Non-invasive assays for candidate substrates

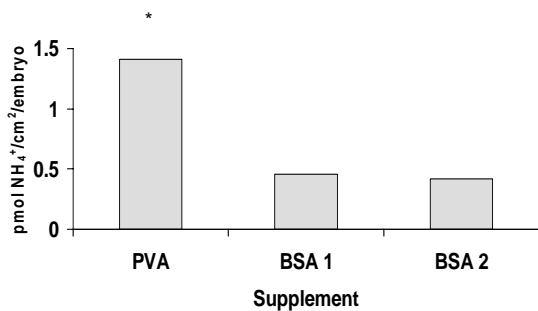
- **NAD(P)H assays of “spent” medium**
 - Based on Lowry assays
 - Fluorescence
 - $\text{Pyruvate} + \text{NAD}^+ \xrightarrow{\text{LDH}} \text{Lactate} + \text{NADH}$
- **HPLC and mass spectrometry of “spent” medium**
 - Amino acid analysis
- **Electrode technology – real time measurement of metabolic activity**
 - Automated scanning electrode – many different ions and compounds
 - Polarographic electrode technology - O_2 consumption

ASET measurement of NH_4^+



Also available for a wide range of ions and compounds, including oxygen & H^+

Ammonium production from bovine blastocysts following culture in different supplements



Biosensors and laminates

- “Smart sensor” technology
- Biofilms sandwiched between impermeable and permeable layers
- Ligand-induced reactions
 - Immunological interactions
 - Receptor-ligand interactions
 - Chemical reactions shifting colour, charge which is measurable

The 'Omics era is upon us!

- Genomics
Transcriptomics
Proteomics
Metabolomics
- Comparative Genomic Hybridisation is an example of Genomics applied to embryo selection
- Transcriptome microarrays of cumulus cells and embryos are widely used in research
 - Already some application with association between levels of some cumulus cell gene expression and oocyte developmental competence
- Next speaker talking on proteomics and metabolomics

Embryo production/manipulation

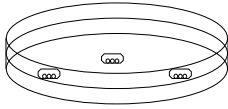
Laser Light trapping



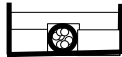
- Particles are "trapped" in the highest energy level of a focussed laser beam
- Possible applications
 - Blastomere removal
 - ICSI
 - Diagnostics on embryos
 - Fertilization

Micro culture systems

Most culture systems are "swimming pools"



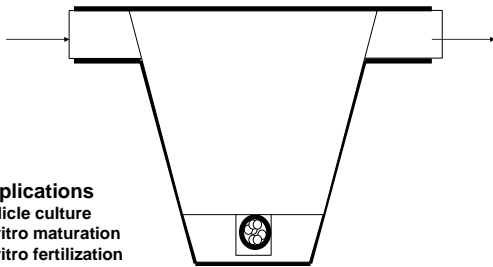
Introduction of microwells, glass oviducts, agar wells



Microperfusion systems

medium in

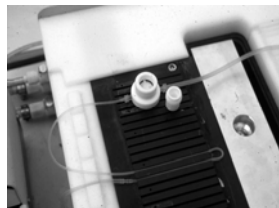
medium out



Applications

- Follicle culture
- In vitro maturation
- In vitro fertilization
- In vitro culture
- Post-hatching development
- Preparation for storage

Micro-perfusion prototype in our laboratory (1995)



Advantages/disadvantages

| Advantages | Disadvantages |
|---|-----------------------|
| Change components Add new components Oil-free Removal of toxins Media stability Measure effluent | Dilute paracrine GF's |

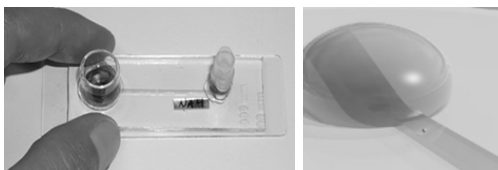
Adapted from Thompson, Theriogenology 2007

Bovine Day 2 - 7 embryo development in perfusion system

| | No. cleaved embryos | Blastocysts (%) | Transferable quality (%) | Cell number |
|-----------|---------------------|-----------------|--------------------------|-----------------------|
| Static | 246 | 44 ± 5 | 31 ± 4 | 175 ± 9 ^a |
| Perfusion | 131 | 44 ± 6 | 19 ± 5 | 131 ± 12 ^b |

Adapted from McGowan & Thompson, Proc. Aust. Soc. Reprod. Biol. 1997

Microfluidic systems involve creating microenvironments
- improved embryo culture success

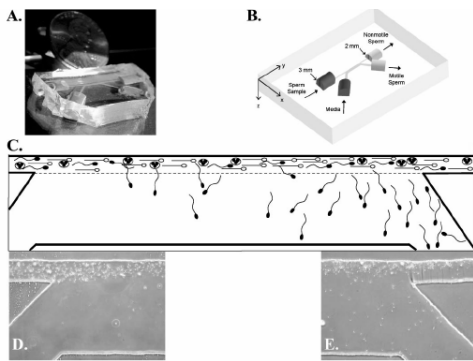


From Vitae LLC www.vitaellc.com

Microfluidics

- Uses the principles of "laminar flow"
- Can move gametes and embryos to different locations in "circuit"
- Can use the principles of "laminar flow" to perform mechanical functions
 - e.g. Cumulus cell and zona pellucida removal already possible
- Represents the next "era" in culture systems
 - Less gamete and embryo handling (including ICSI)
 - Less cellular stress

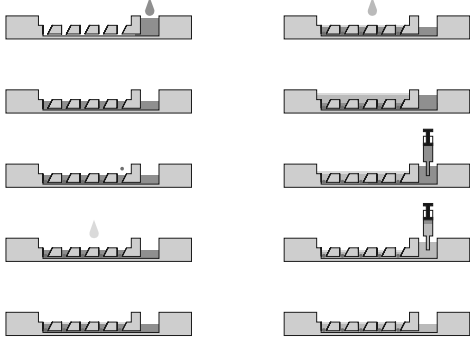
Microfluidics to separate motile/non motile sperm (Suh et al., J. Androl. 2005)

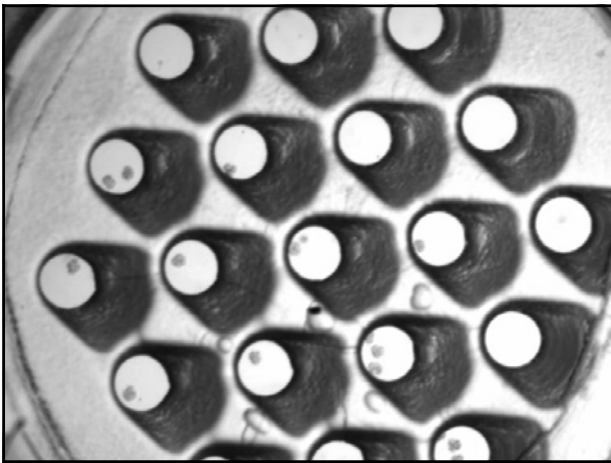


Microfluidic chip – unique addressing



Operation of micro-chip





Why have microfluidic systems yet to make an impact on IVF?

- Material sciences and performance?
 - Properties of materials, especially plastics and resins not adequate
 - Gas permeability – especially using HCO_3^- -buffer
 - Water permeability – many lithographic materials and plastics are H_2O permeable
 - Both problematic for delivery of a pre-warmed & gassed media
- Clear advantage/improvement in process and/or outcomes?
 - Cost vs benefits

Many thanks to the following for their inspiration:

- Lindsay McGowan
Debbie Berg
Jim Dunlop
(AgR New Zealand)
- Michelle Lane
Kara Cashman
(Uni. Adelaide)
- David Gardner
(Uni. Melbourne)
- Matt Wheeler
(Uni. Illinois)
- Kim Giliam
Jason Spittle
Andrew Hirsch
Sean O'Brien
Mike Junger
(Cook Australia)
- Michael Barry
(Repromed)

References

- McGowan LT, Thompson JG (1997) Perfusion culture of bovine in vitro produced embryos. Proc. Aust. Soc. Reprod. Biol. 28: 24.
- Suh R, Takayama S, Smith GD (2005) Microfluidic applications for andrology. J. Androl. 26: 664-670.
- Thompson JG (2007) Culture without the petri-dish. Theriogenology 67: 16-20.






Non-invasive Embryo Assessment: Proteomic and Metabolomic Biomarkers

Zsolt Peter NAGY M.D., Ph.D., HCLD
Scientific and Laboratory Director
Reproductive Biology Associates
1150 Lake Hearn Dr. Suite 600
Atlanta, GA, 30342
Disclosure: Member of Scientific Advisory Board of Molecular Biometrics

Learning Objectives

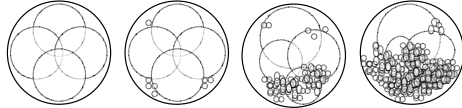
- To review the traditional tools of embryo assessment (advantages and limits)
- To review the need for new/improved techniques
- To review alternative, non-invasive embryo assessment techniques
- Future Perspectives / Conclusions

Historic and Current Approaches

| | | |
|-------|---|---|
| Day 1 |  | Oocyte quality: zona, cytoplasm, PB's 2PN Assessments - Z scoring |
| Day 2 |  | Early cleavage/ Multinucleation |
| Day 3 |  | Genetic screen (PGD); metabolic evaluation, morphologic evaluation; extended culture decision |
| Day 4 |  | ? |
| Day 5 |  | Selection blastocyst for transfer, embryo cyopreservation, PGD |

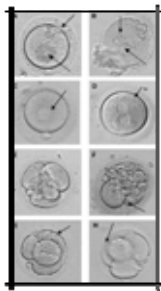


Traditional Embryo Development Grading



| A | B | C | D |
|------------------|---|---|-------------------------|
| Even Blastomeres | → | | Uneven Blastomeres |
| No Fragmentation | → | | Increased Fragmentation |

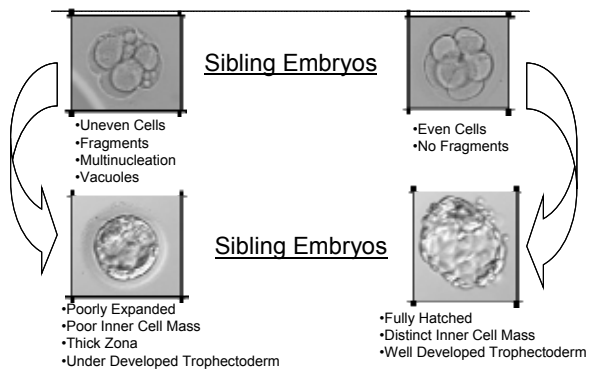
Embryo Score and Blastocyst Development



Examples of normal and abnormal morphological characteristics of oocytes, zygotes, and embryos. (A) Oocyte with a large perivitelline space (solid arrow) and a dark patch (dashed arrow); (B) oocyte with a vacuole (dashed arrow) and a necrotic patch (solid arrow); (C) oocyte accumulation of smooth endoplasmic reticulum; (D) normal zygote with halo (arrow), central pronuclei and polarized and coalesced nucleoli; (E) embryo with much fragments (out of focus, fragmentation score 1) uneven-sized and irregular-shaped (oval) blastomeres; (F) highly fragmented embryo (fragmentation score 0) and abnormal cytoplasm in one of the blastomeres (clear zone at arrow); (G) four-cell embryo with slightly irregular-shaped (oval) blastomeres, one of which jagged membrane; (H) four-cell embryo with slightly irregular-shaped and uneven sized blastomeres, and a gap between the blastomeres and the zona pellucida, thus classified as "Does not fill the space under the zona."

Sjöblom. Embryo score and blastocyst development. Fertil Steril 2006.

Day 3 to Day 5 Development



Early Stage Morphological Characteristics

Evaluation of the pronuclear-stage oocyte

1. Number of pronuclei

- 0 Not fertilized
- 1 Parthenogenetic activation (or asynchronous development of pronuclei – check later)
- 2 Normal fertilization (if there is the presence of 2 PBs at the same time)
- 3+ Abnormal fertilization

2. Size of pronuclei

(in case of 2 PN fertilization)

- Optimal Normal Size and Equal Size
- Suboptimal Larger/Smaller Size and/or Unequal Size

3. Nucleoli (in case of 2 PN fertilization)

Number of nucleoli – in each pronuclei

Size of nucleoli (small or medium or large – in each of the two pronuclei)

Polarization of nucleoli

- 1. POLARIZED: when all the nucleoli were aligned on the side of the pronucleus near to the other pronucleus
- 2. non-POLARIZED: when the nucleoli were dispersed (or not completely aligned) in the pronucleus

4. Aspect of Polar Bodies

- 1. Single non-fragmented Polar Body
- 2. Single fragmented Polar Body
- 3. Two Polar Bodies (fragmented or non-fragmented) - the presence of 2 PB is required for normal fertilization

5. Distance between the Polar Bodies (in case of the presence of 2 PB)

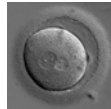
- Optimal Close
- Suboptimal Distant

6. Concentration of cytoplasmic organelles

For each oocyte the % of cytoplasmic organelles concentration/oocyte diameter (%C) is estimated (usually this is between 10% and 20%)

7. Aspect of the zygote cytoplasm

- Suboptimal presence of vacuoles
- Suboptimal presence of the refractal body



Nagy, RBA

Early Stage Morphological Characteristics

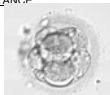
EMBRYO EVALUATION CRITERIA

RELATIVE IMPORTANCE

1. Number of blastomeres

| | Day 2 | Day 3 |
|---------|-----------|-----------|
| Optimal | 4-6 cells | 7-8 cells |
| Medium | 3 cells | 5-6 cells |
| Poor | 2 cells | 2-4 cells |

STRONGLY



2. Dimension of blastomeres – Day 2 and Day 3

- Optimal similar or equal size blastomeres
- Suboptimal different size blastomeres

MODERATELY/STRONGLY

3. Proportion of anucleate fragments – Day 2 and Day 3

- Optimal between 0-10%
- Good between 10-30%
- Medium between 30-50%
- Poor more than 50%

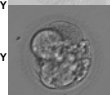
STRONGLY



4. Quality of the cytoplasm

- Optimal: normal appearance
- Suboptimal presence of cytoplasm abnormalities (granulated, vacuoles, refractile bodies)

MODERATELY

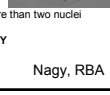


5. Multinucleation of blastomeres

The evaluation of multi-nucleated blastomeres is easier at the 2-4 cells stage.

- Optimal Number of cells with a single nucleus
- Suboptimal Number of cells with multinucleation - Cells with two equal-sized nuclei
- Poor Number of cells with multinucleation - Cells with two unequal sized nuclei or more than two nuclei

MODERATELY



6. Early cell compaction

- Optimal Cells compaction start after 8-10 cell stage embryo (end of day 3 or day 4)
- Optimal Cell compaction is not observed (or not strong) until the end of day 3
- Suboptimal Cell compaction is strongly present late day 2 or early day 3

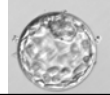
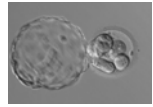
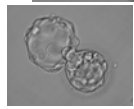
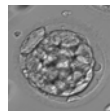
MODERATELY

Nagy, RBA

Blastocyst Stage Assessment

Classification of Blastocyst Development

- Stage A - Hatched blastocyst
- Stage B - Hatching blastocyst
- Stage C - Fully Expanded
 - Thin zona p.
 - Distinct ICM
- Stage D - Distinct Trophoderm
 - A single cavity occupying >50% of the volume of the embryo
 - Regular zona p.
 - The ICM and trophoderm may not be clear
- Stage E - A distinct single cavity 25- 50% of the volume of the embryo.
 - The diameter of the embryo is unchanged.
 - Zona is unchanged



Classification of Blastocyst Quality

ICM

- Grade 1 - Tightly packed, many cells
- Grade 2 - Loosely grouped, several cells
- Grade 3 - Very few cells

Trophoblast

- Grade 1 - Many cells forming a cohesive epithelium
- Grade 2 - Few cells forming a loose epithelium
- Grade 3 - Very few cells

Nagy, RBA

Follicle **Human Oocyte and Embryo Assessment for ART**

Follicular Size

Effect of follicular size on oocyte retrieval, fertilization, cleavage, and embryo quality in in vitro fertilization cycles: a 6-year data collection.

Wittmaack FM, Kreger DO, Blasco L, Tureck RW, Mastroianni L Jr, Lessey BA.

Based on this evaluation of a large number of follicles, follicular size is a useful indicator of oocyte recovery, fertilization, and cleavage in IVF cycles. For optimal results, the follicular fluid volume in gonadotropin- and hCG-stimulated cycles should be > 1 mL, which corresponds to a follicle diameter of > 12 mm, and not larger than 7 mL (24 mm).
Fertil Steril. 1994 Dec;62(6):1205-10.

Cumulus **Human Oocyte and Embryo Assessment for ART**

Assessment of Cumulus

Assessment of human oocyte developmental competence by cumulus cell morphology and circulating hormone profile

Sato, Chikako; Shimada, Masayuki; Mori, Takahide; Kumasako, Youko; Otsu, Eiko; Watanabe, Hirohiko; Utsunomiya, Takafumi

The highest development-supporting competence was observed not with oocytes in grade A COC harvested from natural cycles, but with oocytes in grade B COC from FSH-primed cycles. Hormonal profiles in patients bearing grade B COC were characterized by moderate response in estradiol and progesterone production following FSH, with LH/FSH ratio being below 1.0.
Reproductive BioMedicine Online, Volume 14, Number 1, January 2007, pp. 49-56(8)

Oocyte **Human Oocyte and Embryo Assessment for ART**

Morphology of in-vitro matured oocytes: impact on fertility potential and embryo quality

Anne Lis Mikkelsen and Svend Lindenberg

Significantly more embryos of good quality developed after grade I oocytes [54/144 (37.5%)] compared with those from grade II and grade III oocytes (22/120; P = 0.001). The presence of cytoplasmic abnormalities significantly decreased the cleavage rate (P = 0.04) and also the number of good quality embryos (P < 0.001).

Human Reproduction, Vol. 16, No. 8, 1714-1718, August 2001

| Table III. Fertilization rate and embryo quality according to morphological grading of MII oocytes | | | | |
|--|-----------------|-------------------|------------------------|--------------------------|
| Grading | No. MII oocytes | Fertilization (%) | Cleaved (%) | Good quality embryos (%) |
| Grade I: no anomalies | 144 | 93 (64.6) | 77 (53.5) ^a | 54 (37.5) ^{a,b} |
| Grade II: no anomaly | 97 | 66 (68.0) | 51 (52.6) ^a | 37 (38.1) ^a |
| Grade III: no sperm tail anomalies | 33 | 20 (60.6) | 16 (48.5) | 7 (21.2) ^b |
| Grade IV: grade III | 120 | 66 (55.0) | 49 (40.8) | 22 (18.3) ^b |

MI2 = metaphase II.
Percentages with same superscript in a column differ significantly.
^aP < 0.05, ^bP < 0.005, ^cP < 0.002, ^dP < 0.001.

| Table IV. Fertilization, cleavage and embryo quality according to cytoplasmic and extracytoplasmic abnormalities | | | | |
|--|-----------------|-------------------|------------------------|--------------------------|
| | No. MII oocytes | Fertilization (%) | Cleaved (%) | Good quality embryos (%) |
| No anomalies | 144 | 93 (64.6) | 77 (53.5) | 54 (37.5) |
| Cytoplasmic anomalies | 30 | 12 (40) | 7 (23.3) ^a | 2 ^b |
| Extracytoplasmic anomalies | 86 | 54 (62.8) | 42 (48.7) ^a | 22 (25.6) ^b |

Percentages with the same superscript in a column differ significantly.
^aP < 0.05, ^bP < 0.001.

Current IVF Outcomes

The Need for New/Improved Techniques

Fresh Embryos From Non-Donor Oocytes

| | <35 | 35-37 | 38-40 | 41-42 |
|---|--------|--------|--------|-------|
| Number of cycles | 37,168 | 21,336 | 18,174 | 8,631 |
| Percentage of cycles resulting in live births | 38.8 | 30.6 | 20.6 | 10.9 |
| Percentage of cycles with elective single embryo transfer | 3.3 | 1.9 | 0.7 | 0.3 |
| Implantation rate | 30.8 | 23.5 | 15 | 8.2 |
| Average number of embryos transferred | 2.3 | 2.5 | 2.9 | 3.2 |
| Percentage of live births with twins | 32.3 | 27.7 | 21.4 | 15.5 |
| Percentage of live births with triplets or more | 2 | 1.9 | 1.4 | 0.6 |

https://www.sartcofsonline.com/rt/csr_PublicMultiYear.aspx?ClinicPKID=0 -2006

Estimated IVF Birth Efficiency

- 3-6% of Follicles
- 6-12% of Oocytes
- 10-20% of Embryos (D3)
- 15-40% of Blastocysts

Complications of Multiple Pregnancies

- The incidence of cerebral palsy is increased
 - 8-fold in twins and
 - 47-fold in triplets
- Infant deaths (birth to 1 year) are increased
 - 11.2/1000 live births for singletons
 - 66.4/1000 live births for twins
 - 190.4/1000 live births for triplets and higher order gestations

*Keith et al. Int J Fertil Womens Med 2000;45:206-14.
 *U.S. 1998. DHHS Pub No. (PHS) 90-50212. 24

What is Expected from the Alternative Tests?

- Non-invasive (culture medium / fluid / sem. plasma)
- "Easy" (ease of use)
- "Simple" (no need for special knowledge / training)
- Rapid (should take only minutes)
- Cheap (both the instrument and "reagents")
- Reliable / reproducible function
- Fit to the IVF Lab
- Does not interrupt standard routine (dish / culture)
- Provide more useful results than morphology

Does this currently exist? NO

Possible Targets to Use for Alternative Embryo Assessment Approaches

- | | |
|--------------------|---|
| Morphology | - Birefringence (SpindleView) * |
| Metabolic Activity | - Pyruvate/Glucose uptake - Amino acids * - Oxygen consumption (Respirometry) * |
| Constituents | - Genome - Transcriptome - Proteome * - Metabolome |
| Secreted Factors | - PAF - HLA _g * - "Secretome" * |

Polscope / SpindleView

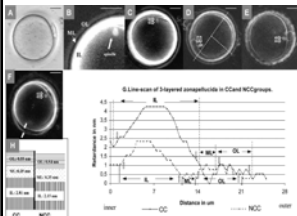


Table III. Mean birefringence magnitude and thickness of the individual zona layers as assessed by Polscope microscopy in oocytes contributing to conception cycle (CC) and non-conception cycle (NCC) groups

| | CC (n=100) | NCC (n=100) |
|-------------------------------|---------------|--------------|
| No. of patients | 23 | 40 |
| No. of oocytes | 403 | 393 |
| Zona inner layer (mean ± SD) | 2.81 ± 0.61* | 2.15 ± 0.41 |
| Birefringence (µm) | 11.23 ± 1.44* | 9.36 ± 1.74 |
| Thickness (µm) | | |
| Zona middle layer (mean ± SD) | 0.35 ± 0.08 | 0.35 ± 0.07 |
| Birefringence (µm) | 3.92 ± 0.76 | 3.66 ± 0.65 |
| Thickness (µm) | | |
| Zona outer layer (mean ± SD) | 0.55 ± 0.18 | 0.55 ± 0.14 |
| Birefringence (µm) | 4.80 ± 1.40 | 5.55 ± 1.05 |
| Thickness (µm) | 19.87 ± 1.92* | 19.58 ± 1.82 |

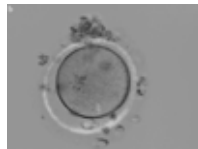
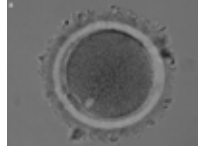
*Significantly different from the NCC group *P < 0.001, **P < 0.01 (t-test).

Shen, Y. et al. Hum. Reprod. 2005 20:1596-1606

Polscope / SpindleView

Table 2. Cycle outcome in relation to oocyte zona birefringence.

| Parameter | ICSI-ZB | ICSI-ZN | ZB-ZN | P-value |
|-------------------------------|--------------|---------------|---------------|---------|
| No. of cycles | 23 | 30 | 43 | - |
| Mean natural age (years) ± SD | 33.9 ± 3.6 | 34.4 ± 4.0 | 34.4 ± 4.1 | NS |
| Mean no. of MI oocytes ± SD | 10.9 ± 3.9* | 7.6 ± 3.0 | 6.7 ± 2.7* | <0.05 |
| ICB oocytes (%) | 73.28 (13.5) | 88.33 (21.3) | 36.43 (8.3)* | <0.001 |
| Fertilization rate (%) | 102.0 (74.3) | 256.37 (87.7) | 296.03 (88.4) | NS |
| Implantation rate (%) | 10.48 (8.0) | 26.10 (26.0) | 17.18 (11.1)* | <0.02 |
| Pregnancy rate (%) | 13.28 (8.0) | 25.90 (26.0) | 16.85 (14.6)* | <0.05 |
| Miscarriage rate (%) | 1.03 (7.7) | 5.21 (20.0) | 3.13 (21.0) | NS |
| Live birth rate (%) | 12.25 (8.0) | 20.90 (26.0) | 13.65 (20.0)* | <0.02 |



ZB = high zona birefringence; ZN = low zona birefringence; MI = multiple MI; NS = not statistically significant.
*P < 0.05, values with the same superscript letter are significantly different. All other comparisons were not significantly different.

This study concludes that oocyte zona birefringence is a good selection criterion and a good predictive criterion for embryo implantation potential. Montag et al., 2007 RBMonlin

Polscope / SpindleView

Light retardance by human oocyte spindle is positively related to pronuclear score after ICSI

Shen Y, Staf T, Mehnert C, De Santis L, Cino I, Tinneberg HR, Eichenlaub-Ritter U. The study suggests that quantitative evaluation of mean retardance of light by the oocyte spindle predicts oocyte health, is related to PN score of the embryo and may be especially useful to assess oocyte quality in countries with legal restrictions to select after fertilization. *Reprod Biomed Online*. 2006 Jun;12(6):737-51.

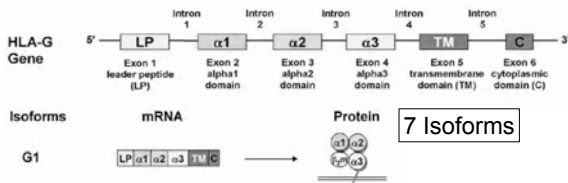
De Santis; Rienzi; Keefe: Spindle and Fertilization / Embryo Development

Evaluation of metaphase II spindle length, retardance and its relationship to embryo quality on day 3 and day 5

T. H. Taylor, T. Elliott, S. A. Gitlin, S. Jones-Colon, H. I. Kort, Z. P. Nagy. Spindle length, as measured by the Oosight, correlates to embryo quality on day 3 and blastocyst quality on day 5. *Fertility and Sterility*, Volume 86, Issue 3, Pages S115-S116

Assessment of Soluble Human Leukocyte Antigen G in Human Embryos

Detection of soluble HLA-G in embryo culture medium has been correlated to pregnancy success in 12 studies, but three studies were not able to detect sHLA-G



- HLA-G expression has been correlated with an increased rate of cleavage division.
- Soluble HLA-G may alter cytokine expression on maternal cells at the site of implantation as well as induce apoptosis in CD8+ maternal T cells to allow successful implantation (ability of the embryo to modulate the maternal immune response and successfully implant into the uterus)
Warner et al., 2008

Assessment of Soluble Human Leukocyte Antigen G in Human Embryos

Soluble HLA-G and Pregnancy Success

Carol M. Warner, Paula W. Lampton, Judith A. Newmark, and Jacques RBMonline; in press

HLA-G Detected in Culture Supernatant: Fuzzi et al.; Roussev et al.; Sher, Noci et al.; Yie et al.; Sher et al.; Criscuoli et al.; Desai; Fisch et al.; Rebmann et al.; Rizzo et al.

HLA-G NOT Detected in Culture Supernatant: van Lierop et al.; Sageshima et al.; Sageshima et al.

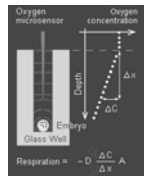
- To be standardized:
- Culture methods
 - Sampling methods
 - Detection methods

There are many questions that remain to be answered before one would have full confidence in using sHLA-G as a selection marker for embryos

Embryo Respiration Unisense

Embryo Respirometry – A novel technique to improve embryo selection in IVF procedures by measuring respiration rates of individual embryos

Henrik Callesen, Lars Hauer Larsen, Lars Damgaard, Ana Sofia Lopes, Torben Greve and Niels Birger Ramsing.
Oral presentation at The XVI Nordic IVF Meeting., January 3-6 2005 in Are, Sweden.

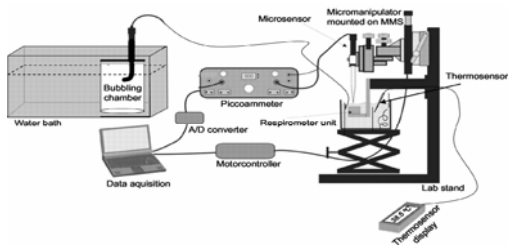


Improved embryo selection using embryo respirometry
Henrik Callesen, Danish Institute of Agricultural Sciences, DK-8830 Tjele, Denmark
Oral presentation, ESHRE June 19-22, 2005, Copenhagen.



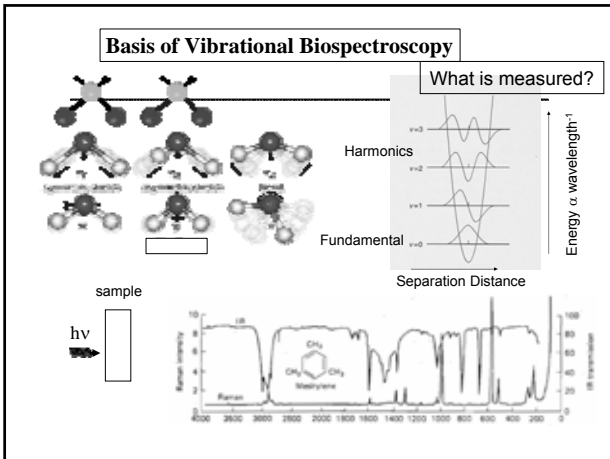
Lopes et al., 2007; Scott et al., 2008

Embryo Respiration Unisense



| Respiratory category | Pregnant | Non-pregnant |
|-------------------------|---------------|--------------|
| High (>1.10 nl/h) | 25% (n = 1) | 75% (n = 3) |
| Medium (0.78-1.10 nl/h) | 100% (n = 13) | 0% (n = 0) |
| Low (<0.78 nl/h) | 48% (n = 11) | 52% (n = 12) |

Respiration rates of individual bovine *in vitro*-produced embryos measured with a novel, non-invasive and highly sensitive microsensor system. Lopes et al., 2005



What is Measured?

- Clinically
 - How the embryo modifies its environment
- Biologically
 - Changes in concentrations of:

| Functional Groups | Constituents |
|-------------------|--------------|
| •CH | •Albumin |
| •NH | •Lactate |
| •OH | •Pyruvate |
| •SH | •Glutamate |
| •C=C | •Glucose |

41

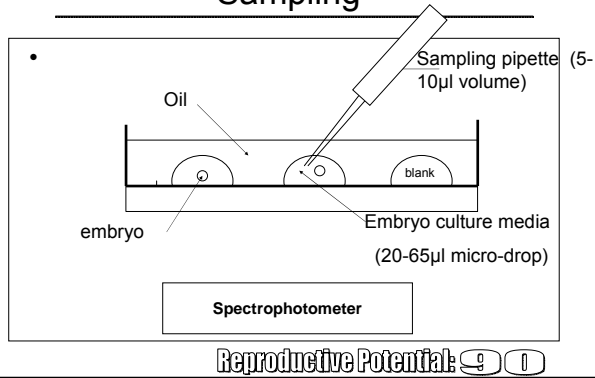
Generation of Embryo Assessment Score

The graph shows the generation of an embryo assessment score. The x-axis is Wavelength (nm) from 900 to 1700. The y-axis is Ratio. A plot shows Log10 Ratio of Sample divided by Blank. Wavelets B₁, B₂, B₃, B₄, B₅ are identified. The formula for the score is: $0.35 + 0.081B_5 - 0.061B_4 - 0.023B_3 - 0.0015B_2 + 0.0026B_1$.

ViaTest-E™ Score

42

Non-invasive Embryo Selection Sampling



Multicenter, Multinational Study Design Single Embryo Transfer

- Standard IVF procedure
- Single embryo culture
- Morphology assessment
- Single embryo transfer: D2, D3, D5
- Media sample plus controls (7µl)
- NIR spectral analysis; Bioinformatics
- Pregnancy confirmed by FCA (Fetal Cardiac Activity)
- 677 samples from 1768 patients analyzed

Accuracy

- A measure of a test's ability to correctly identify positive and negative FCA pregnancy from a complete IVF patient population.

- The accuracy of a test can be determined by calculating:

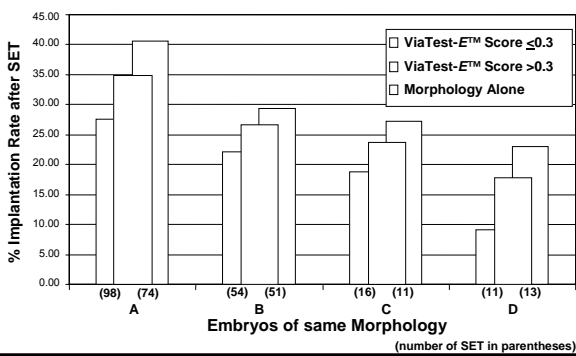
$$\frac{TP + TN}{TP + FN + TN + FP}$$

- where TP = true positive
- TN = true negative
- FP = false positive
- FN = false negative

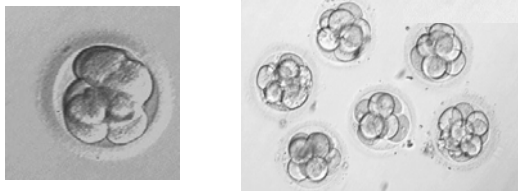
Accuracy of ViaTest-E™ vs. Morphology

| Day of Transfer | % Accuracy | |
|-----------------|------------|------------|
| | Morphology | ViaTest-E™ |
| Day 2 | 31.9 | 71.3 |
| Day 3 | 55.0 | 74.0 |
| Day 5 | 48.3 | 79.2 |
| Global Average | 45.1 | 74.8 |

SET Implantation Rates of Day 3 Embryos Comparing the Same Morphology Grade and a ViaTest-E™ Score of \leq or $>$ than 0.3



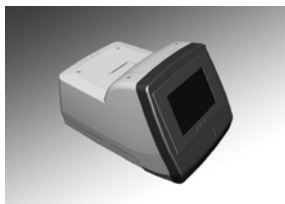
...so the challenge remains



Objective: Singleton ongoing pregnancy

How to select the best Embryo???

Where will we end up.....?



VS



Conclusions

- The current "traditional" evaluation methods have limited capability to accurately and reliably assess gamete / embryo viability, and developmental potential.
- There is a clear need to improve ART efficiency – higher pregnancy rate and lower (or no) risk for multiple gestation.
- To achieve these goals, a new, reliable (alternative or additional) embryo / gamete assessment technology is required.

Conclusions

- This new system should be a non-invasive, simple, rapid, reliable "on site" system that can fit easily in any laboratory and in any routine.
- Currently, there are a number of techniques under development / investigation that in the future may become useful tools to help to assess gamete / embryo potential / viability.
- These systems have to be evaluated in a prospective, randomized (blinded if possible) fashion to demonstrate their benefit.

**An Update on Embryo Culture for Human Art:
Media, Performance and Safety**

**Thomas B. Pool, Ph.D.
Fertility Center of San Antonio
San Antonio, Texas USA**

Commercial Relationships

- Irvine Scientific, Inc.
- Incept Biosystems, Inc.

Learning Objectives

Following this presentation, attendees should be able to:

1. Identify the nutritional strategies employed in contemporary human embryo culture media.
2. Describe the basis of concerns for potential health hazards introduced by embryo culture.
3. List the cautions to be taken in evaluating data from animal and human studies with respect to the introduction of potential health hazards.
4. Discuss the rate-limiting aspects of contemporary embryo culture and new approaches to alleviating culture-induced stress.

Somatic Cell Media Used for Human IVF and Embryo Culture

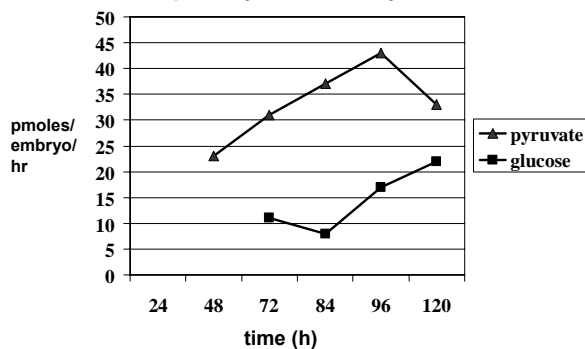
| Year | Investigator | Medium | Use |
|------|---------------|-----------|---------------|
| 1880 | Ringer | salt soln | amphib. heart |
| 1907 | Tyrode | salt soln | intestine |
| 1943 | Earle | salt soln | somatic |
| 1950 | Morgan et al. | Med 199 | somatic |
| 1956 | Eagle | MEM | somatic |
| 1963 | Ham | F-10 | somatic |

Dawn of Specific Media for Human Embryo Culture

Menezo *et al.*, 1984

Quinn *et al.*, 1985

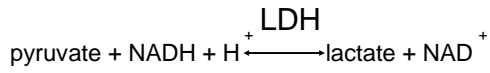
Uptake by Human Embryos



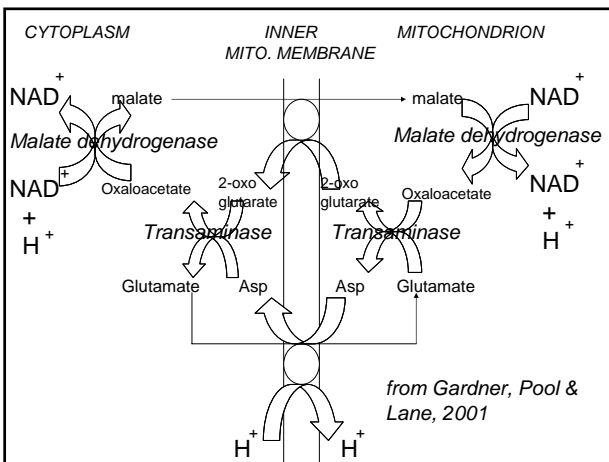
redrawn from Hardy, 1993

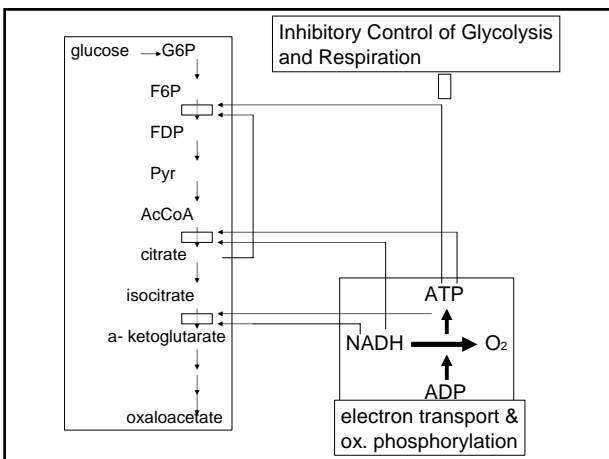
Further Metabolic Considerations

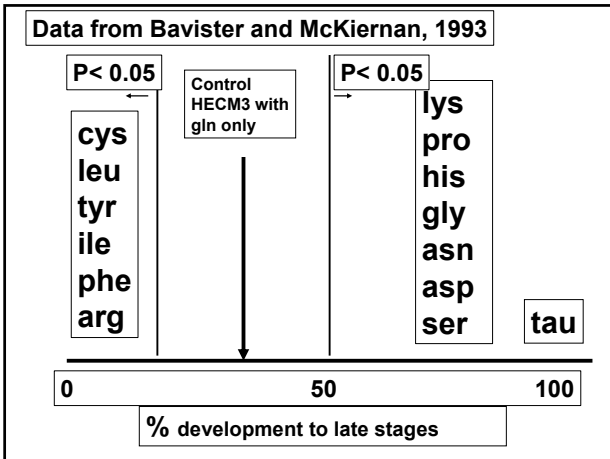
Pyruvate and lactate:



1. Conversion regenerates NAD for glycolysis under anaerobic conditions.
2. Cytoplasmic and mitochondrial pools of NADH are not shared.
3. Transfer of reducing power between compartments requires a specific shuttle.







From Gardner and Lane, 1993; Lane and Gardner, 1997

| zygotes → 8 cell | > 8 cell |
|------------------|----------|
| mMTF | mMTF |
| NEGLN ← | NEGLN |
| | ESS |
| | 20AA ← |
| ESS | NEGLN |
| | ESS |
| | 20AA |
| 20AA | NEGLN |
| | ESS |
| | 20AA |

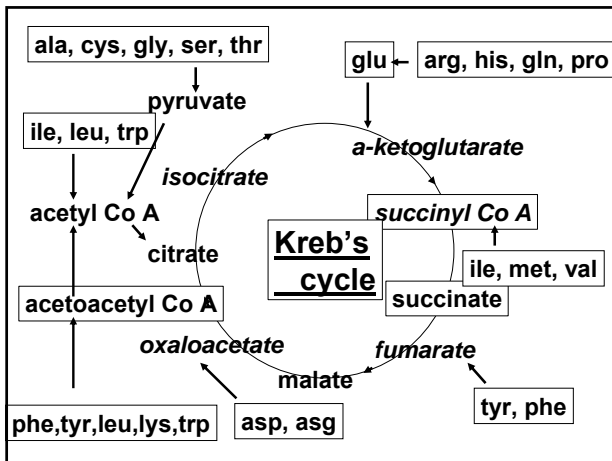
Methionine:

“Essential” amino acid, thus omitted in first interval.

Fuels methylation through synthesis of s-adenosyl methionine

SAM synthesis occurs before genomic activation in humans.

Menezo, YJR. RBM Online 12:616-621, 2006



Adverse effects of ammonium

Ammonium production in embryo culture
Gardner and Lane, 1993

Ammonium in culture induces exencephaly
in F1 hybrid mouse
Lane and Gardner, 1994

Ammonium induces multiple problems for
mouse blastocyst; ammonium build-up
preventable with dipeptide alanyl-gln
Lane and Gardner, 2003

Adverse effects of ammonium

Exposure of mouse embryos to ammonium
chloride induces abnormalities, but at a lower
rate than previously reported
Sinawat et al., 2003

Reports of effects of ammonium discrepant;
studies have produced a biased model of
glutamine effects
Biggers et al., 2004

Replacement of glutamine with glycyl-gln
enhances mouse preimplantation development
Biggers et al., 2004

EFFECTS OF WHOLE SERUM

Vesicular trophoblast in blastocysts cultured from zygotes in serum:
Gardner, 1994; Gardner et al., 1994
Dorland et al., 1994; Thompson et al., 1995

Abnormal mitochondria, reduced oxidative capacity
Dorland et al., 1994; Thompson et al., 1995

EFFECTS OF WHOLE SERUM

Birth of abnormally large offspring after transfer of embryos grown in serum (LOS in ruminants)

Macromolecular Supplementation

Serum albumin
enriched fraction produced by cold alcohol precipitation

recombinant

Serum albumin plus alpha & beta globulins

Serum albumin plus non-serum polyhydroxylated species (dextran)

Problem: proteins in solution versus oligosaccharides emanating from fixed sites via O-linkage to ser/thre

Common Threads: 1st interval

Sequential culture

1. Reduced glucose with a subset of amino acids (no glucose & taurine or 0.5 mM glucose plus NEAA with glycolytic suppressor)
2. 21 mM lactate
3. 0.33 mM pyruvate

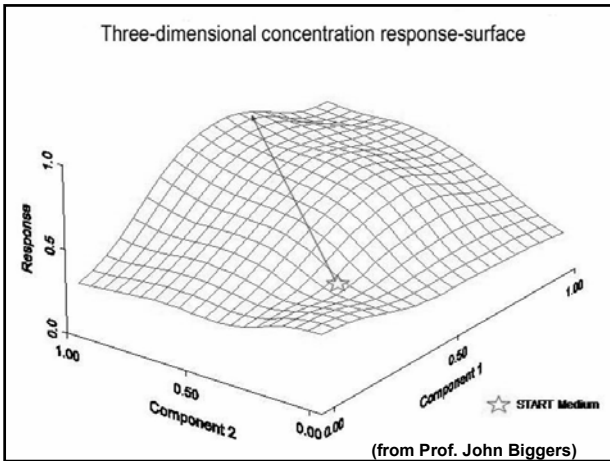
Common Threads - 2nd interval

Sequential culture

1. Elevated glucose (~3 mM) with a complex array of amino acids.
2. Reduced pyruvate (0.1 mM?).
3. Reduced lactate (12 mM?).

Return to Monoculture

Use a complete medium containing electrolytes and energy substrates derived via simplex optimization .



in either case,

Ensure that glutamine is present in the form of a dipeptide.

Components of a culture system:

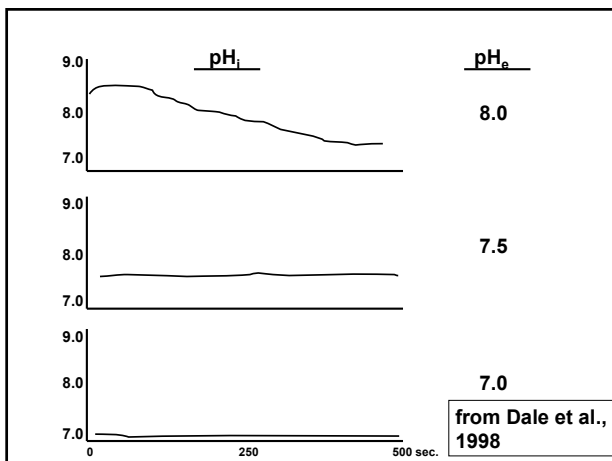
- Culture media
- Macromolecules
- Culture vessel
- Oil overlay
- Incubator
 - temperature
 - humidity
 - gas phase

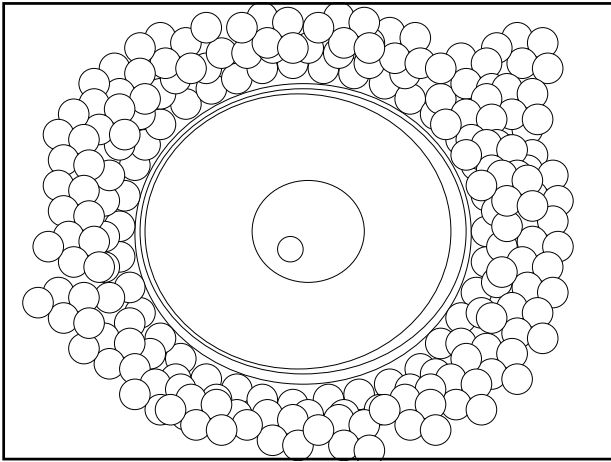
Take Home Lesson:

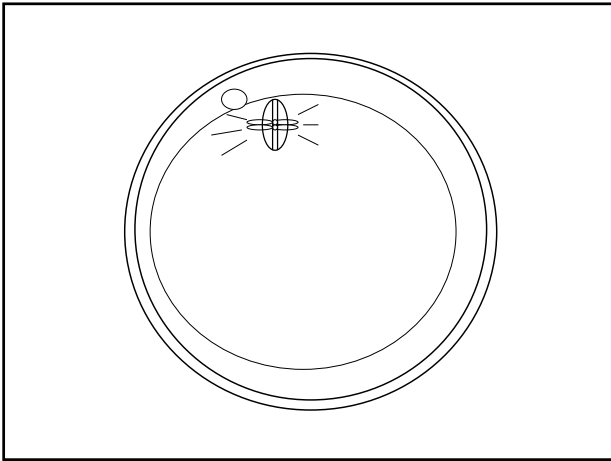
One cannot consider the effects upon outcome of a culture medium formula without simultaneously considering the influence of all other elements of the culture system.

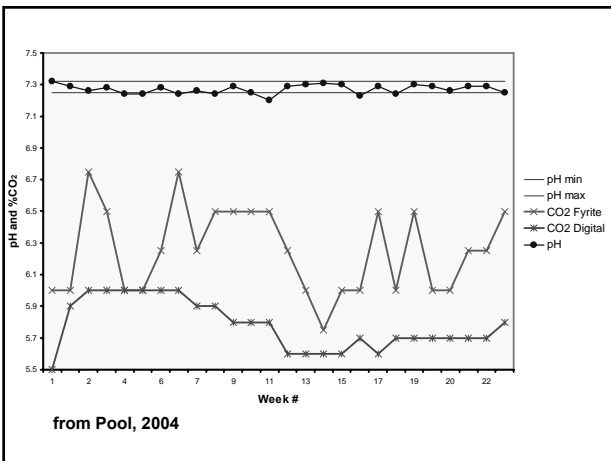
Enhanced and Reproducible Performance from Embryo Culture

- 1. Monitor temperature**
- 2. Understand, measure, monitor and manage pH.**

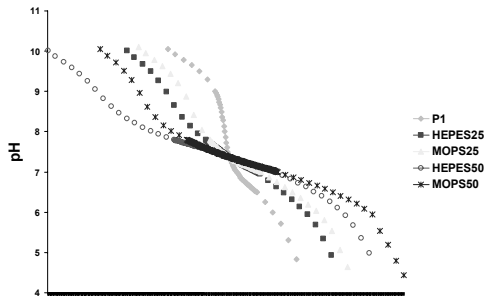




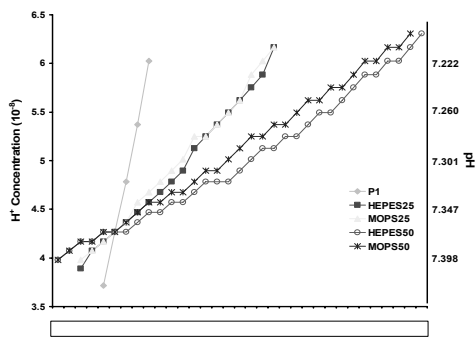




Dose Titration



Dose Titration



The problematic in-vitro embryo in the age of epigenetics

M.H. Johnson, RBMOnline 10 (Suppl. 1):88-96, 2005

**Adapting means getting past the
cumulative stresses of the culture
environment.**

**Might the embryo accumulate non-
lethal defects that later have
inappropriate fetal, neonatal or adult
consequences?**

“primum non nocere”

Epidemics: “As to diseases, make a habit of two things – to help, or at least to do no harm”

Epigenetic deregulation of genomic imprinting

Allele-specific expression of imprinted genes:

- depends on inheritance from mother or father
- controlled by DNA methylation in ICR
- epigenetic “life cycle” (germline erasure, germline establishment, somatic maintenance) disrupted in some diseases

Arnaud P. and Feil R., Birth Defects Res.,75:81-97, 2005

Epigenetic deregulation of genomic imprinting

Examples of such diseases:

- Beckwith-Wiedemann syndrome (BWS)**
- Prader-Willi syndrome (PWS)**
- Angelman syndrome (AS)**
- Hydatiform mole**
- Rett syndrome (ICR mediation of imprinted expression perturbed)**

Arnaud P. and Feil R., Birth Defects Res.,75:81-97, 2005

What has precipitated the concern?

Ruminant LOS.

H19 imprinting errors induced in mouse models with certain media.

Altered gene expression in bovine embryos.

Elevated incidence of certain imprinting disorders after ART.

Imprinting disorders inherited in ESC.

Cautions for interpreting model data:

Faulty media formulations induce errors.

Inappropriate medium supplements can induce errors.

Inappropriate gas phase produces errors.

Genetic strain influences imprinting errors.

Ammonium induces imprinting errors.

Was superovulation used in the study?

“primum non nocere”

Epidemics: “As to diseases, make a habit of two things – to help, or at least to do no harm”

“pluralitas non est ponenda sine neccesitate”

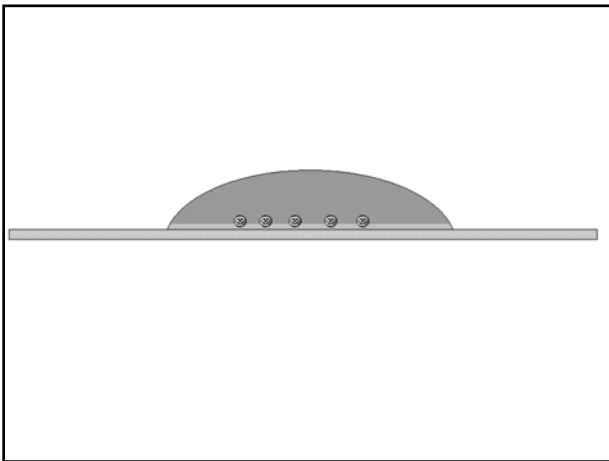
“plurality should not be posited without necessity”

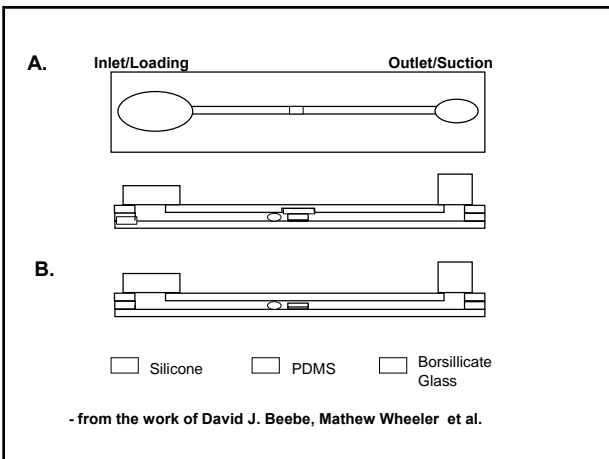
What's wrong with the microdrop?

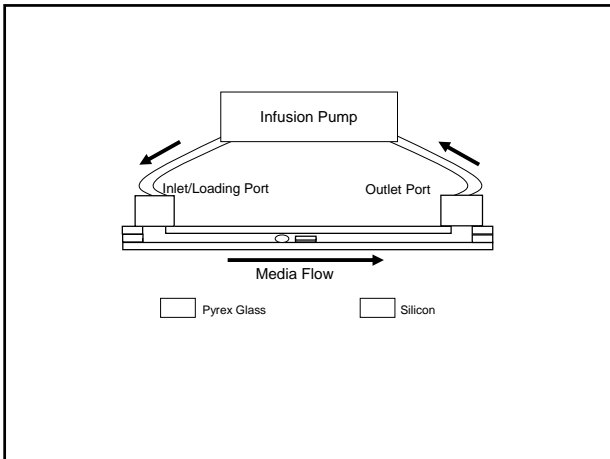
Maximum concentration of solutes obtainable is determined by solubility.

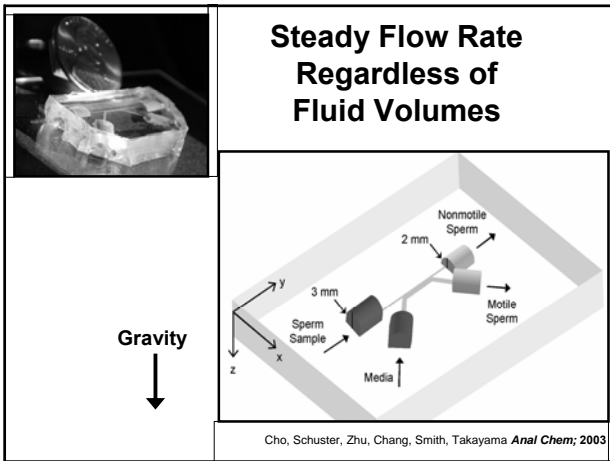
Functionally static

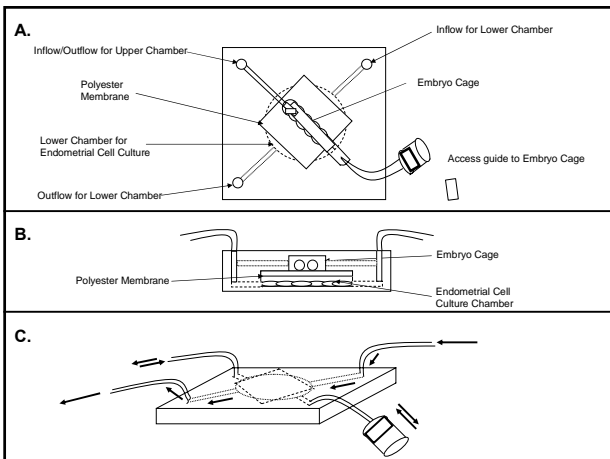
Third dimension not available for surface chemistry.











Embryo Culture in Microchannels

Raty *et al.*, Theriogenology, 2000

| Stage | μF | μdrop |
|----------------|---------------|------------------|
| 16c/mor. – 24h | 23.5* | 4.7 |
| Blast. – 48h | 17.6** | 2.4 |
| Blast. – 72h | 72.9** | 42.9 |
| Hatched – 72h | 4.1* | 0 |
| Hatched – 96h | 26.5** | 8.8 |

* $p < 0.05$; ** $p < 0.01$

In each arm, $n = 170$

Microfluidics For Sperm, Eggs, and Embryos

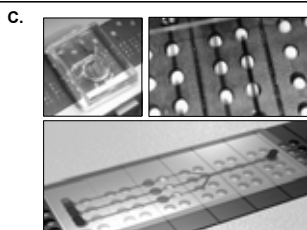
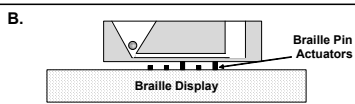
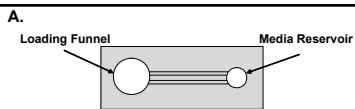
University of Michigan Interdisciplinary Team



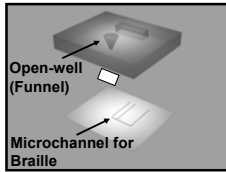
Shuichi Takayama, Ph.D.
Departments of Biomedical Engineering & Macromolecular
Science and Engineering
Specializes in Microfluidics for Cell Culture and Analysis



Gary D. Smith, Ph.D.
Department of OB/GYN, Urology, and Physiology
Laboratory Director:
University of Michigan Fertility Clinic
Huntington Reproductive Center of Brazil



Current Embryo Culture Chip

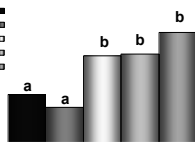


- Easy in, easy out
- 100% recovery
- Quick (reduced pH shifts)
- User friendly

Courtesy of G. Smith / S. Takayama

Microfluidic Embryo Culture

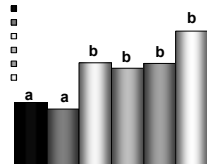
% Hatching Blastocyst



- Improved mouse blastocyst hatching

• Increased blastocyst cell #

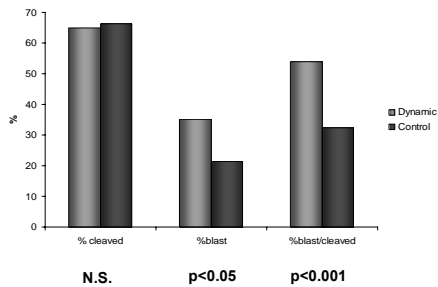
Blastocyst Cell #



Cabrera et al. 2006

Bormann et al., 2007

Bovine zygotes, produced in vitro, randomized into 50 μ l of KSOM w/ 3 mg/mL BSA in either static drops (n=145) or dynamic culture (n=97) in a microfluidic device, cultured 10 per group.



Potential Applications of Microfluidics

Micro-culture volumes with flow.

Medium change by gradient.

High macromolecular:solvent ratios.

Fixed-site, 3-dimensional presentation.

Micro-environmental self-regulation.

Integration of real-time analytical capabilities (metabolome, secretome) with flow controlled solid-phase, micro-volumetric culture in a single platform.

Acknowledgments

Postdoctoral fellow:
Jason E. Swain, M.S., Ph.D.

Laboratory:
Ginny Ord
Darleen Taylor,
Veronica Sessamon
Donna Navarro

Nighthawk Bay
Laguna Madre
South Texas

References

- Arnaud P, Feil R. Epigenetic deregulation of genomic imprinting in human disorders and following assisted reproduction. *Birth Defects Res* 2005; 75:81-97.
- Biggers JD, Racowsky C. The development of fertilized human ova to the blastocyst stage in KSOM (AA) medium: is a two-step protocol necessary? *Reprod Biomed Online* 2002; 5(2); 133 – 40.
- Biggers JD, McGinnis LK, Lawitts JA. Enhanced effect of glycyl-L-glutamine on mouse preimplantation embryos in vitro. *RBM Online* 2004; 9:59-69.
- Biggers JD, McGinnis LK, Summers MC. Discrepancies between the effects of glutamine in cultures of preimplantation mouse embryos. *RBM Online* 2004; 9:70-73.
- Bormann C, Cabrera L, Heo YS et al. Dynamic microfluidic embryo culture enhances blastocyst development of murine and bovine embryos. *Biol Reprod* 2007; Suppl 1: Abstr. 62, pp 89-90.
- Bormann CL, Wheeler MB, Beebe DJ, Takayama S and Smith GD. 2007 *Microfluidics for Assisted Reproductive Technologies*. In: "Micro- and Nanoengineering of the Cell Microenvironment: Technologies and Applications" Artech House Publishing Inc.
- Cabrera LM, Heo YS, Ding J, et al. Improved blastocyst development with microfluidics and braile pin actuator enabled dynamic culture. *Fertil Steril* 2006; 86 Suppl 2: 43.
- Dale B, Menezo Y, Cohen J, et al. Intracellular pH regulation in the human oocyte. *Hum Reprod* 1998; 13:964-970.
- Dorland M, Gardner DK, Trounson AO. Serum in synthetic oviduct fluid causes mitochondrial degeneration in ovine embryos. *J Reprod Fertil Abstract Series* 1994; 13: 70.
- Dumoulin JCM, Meijers CJJ, Baras M et al. Effects of oxygen concentration on human in-vitro fertilization and embryo culture. *Hum Reprod* 1999; 14:465-469.
- Duranthon V, Watson AJ, Lonergan P. Focus on Mammalian Embryogenomics Preimplantation embryo programming: transcription, epigenetics, and culture environment. *Reproduction* 2008; 135:141-150.
- Fernandez-Gonzalez R, Ramirez MA, Bilbao A, et al. Suboptimal In Vitro Culture Conditions: An Epigenetic Origin of Long-Term Health Effects. *Molecular Reproduction and Development* 2007; 74:1149-1156.
- Gardner DK. Mammalian embryo culture in the absence of serum or somatic cell support. *Cell Biol Int* 1994; 18: 1163-79.
- Gardner DK, Lane M. Embryo culture systems. In: Trounson A, Gardner DK (eds) *Handbook of In Vitro Fertilization*, 2nd edn. CRC Press, Boca Raton, FL, USA, 2000. pp. 195-254. Gardner DK, Lane M, Johnson J et al. Reduced oxygen

tension increases blastocyst development, differentiation and viability. *Fertil Steril* 72 (suppl.1); S30-S31.

Gardner DK, Lane M. Ex-vivo early embryo development and effects on gene expression and imprinting. *Reprod. Fertil Dev* 2005; 17: 361-70.

Gardner DK, Pool TB, Lane M. Embryo nutrition and energy metabolism and its relationship to embryo growth differentiation, and viability. *Seminars in reproductive Medicine* 18: 205 – 218.

Hardy K. Development of human blastocysts in vitro. In: (Bavister B., ed) *Preimplantation Embryo Development*, Springer-Verlag, New York, NY 1993, pp 184-199.

Johnson MH. The problematic in-vitro embryo in the age of epigenetics. *RBM Online*, 2005 10 suppl 1:88-96.

Lane M, Gardner DK. Increase in postimplantation development of cultured mouse embryos by amino acids and induction of fetal retardation and exencephaly by ammonium ions. *J Reprod Fertil* 1994; 102:305-312.

Lane M, Gardner DK. Ammonium induces aberrant blastocyst differentiation, metabolism, pH regulation, gene expression and subsequently alters fetal development in the mouse. *Biol Reprod.* 2003; 69:1109-1117.

Lawrence LT, Moley KH. Epigenetics and Assisted Reproductive Technologies: Human Imprinting Syndromes. *Seminars in Reproductive Medicine* 2008; 26:143-152.

Menezo YJR. Paternal and maternal factors in preimplantation embryogenesis: interaction with biochemical environment. *RBM Online* 2006; 12:616 – 21.

Menezo Y, Testart J, Perone D. Serum is not necessary in human in vitro fertilization and embryo development. *Fertil Steril* 1984; 42:750.

Pool TB. An update on embryo culture for human assisted reproductive technology: media, performance, and safety. *Semin Reprod Med* 2005; 4:309 - 18.

Pool TB, Blastocyst development in culture: The role of macromolecules. In: *ART and the Human Blastocyst*, Springer-Verlag, New York, NY, Inc., pp. 105-117.

Pool TB. Optimizing pH in clinical embryology. 2004; *Clinical Embryologist* 7:1-17.

Pool TB. Recent advances in the production of viable human embryos in vitro. *RBM Online* 2002; 4:294-302.

Pool TB, Atiee SH, Martin JE. Oocyte and embryo culture. Basic concepts and recent advances. In: May JV (ed.) *Assisted reproduction: laboratory considerations. Infertility and Reproductive Medicine Clinics of North America*, 1998; 9:181-203.

Pool TB, Martin JE. The production of viable human blastocysts: the evolution of sequential culture systems. In: Wolf DP, Zelinski-Wooten M (eds) *Endocrinology:*

assisted fertilization and nuclear transfer in mammals. Humana Press, Totowa, NJ, USA, pp. 137-157.

Quinn P, Kerin JF, Warnes GM: Improved pregnancy rate in human in vitro fertilization with the use of a medium based on the composition of human tubal fluid. *Fertil Steril* 1985; 44: 493.

Raty S, Davis JA, Beebe DJ et al. Culture in microchannels enhances in vitro embryonic development of preimplantation mouse embryos. *Theriogenology* 2001; 55:241.

Rinaudo PF, Giritharan G, Talbi S, et al. Effects of oxygen tension on gene expression in preimplantation mouse embryos. *Fertil Steril* 2006; 86(Suppl 3): 1252-65.

Sinawat S, Wei-Chih H., Flockhart JH et al. Fetal abnormalities produced after preimplantation exposure of mouse embryos to ammonium chloride. *Hum Reprod* 2003; 18:2157-2165.

Steele W, Allegrucci C, Singh R et al. Human embryonic stem cell methyl cycle enzyme expression: modeling epigenetic programming in assisted reproduction? *RBM Online* 2005; 10: 755-66.

Suh RS, Phadke N, Ohi DA, et al. Rethinking gamete/embryo isolation and culture with microfluidics. *Hum Reprod Update* 2003; 9:451-61.

Summers MC, Biggers JD. Chemically defined media and the culture of mammalian preimplantation embryos: historical perspective and current issues. *Hum Reprod Update* 2003; 9(6): 557–82.

Swain JE, Pool TB, Takayama S, Smith GD (2008) Microfluidics in ART: Current Progress and Future Directions. In *Textbook of ART*. Eds: Gardner DK, Howles C, Weissman A, Shoham Z. Informa Publishing, London.

Thompson JG, Gardner DK, Pugh PA, et al. Lamb birth weight following transfer is affected by the culture system used for pre-elongation development of embryos. *J Reprod Fertil Abstract Series* 1994;13:69.

Thompson JG, Kind KL, Roberts CT, et al. Epigenetic risks related to assisted reproductive technologies. Short and long-term consequences for the health of children conceived through assisted reproduction technology: more reason for caution? *Hum Reprod* 2002; 17: 2783-2786.

Thompson JG, Mitchell M, Kind KL. Embryo culture and long-term consequences. *Reproduction Fertility and Development* 2007; 19: 43-52.

Walker SK, Heard TM, Seamark RF. In vitro culture of sheep embryos without co-culture: successes and perspectives. *Theriogenology* 1992; 37:111-26.

Weathersbee PS, Pool TB, Ord T. Synthetic serum substitute (SSS): a globulin-enriched protein supplement for human embryos culture. *J Assist Reprod Genet* 1995;12:354-60.

**24th Annual Meeting of ESHRE
Barcelona - 2008**

**PRE-CONGRESS COURSE:
The Human IVF Lab in 2008 and beyond**

**Safe and Efficient vitrification methods
for human oocytes: how to do it?**

Laura Rienzi, Rome, Italy

Learning objectives

1) SAFETY

- Possible injuries to the oocyte during cryopreservation
- Possible contamination of the oocyte

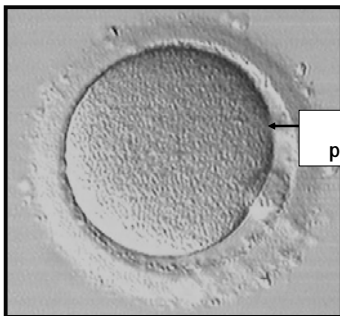
2) EFFICIENCY, WHERE ARE WE?

- oocyte cryosurvival
- oocyte/embryo development post vitrification

3) HOW TO IMPROVE IT?

- Factors that may influence the efficiency


Possible injuries




membrane permeability

g.en.e.r.a.


Membrane permeability



60-65%




35-40%




75-80%

Survival rates of human oocytes frozen with the same slow freezing protocol
(Lassalle et al., 1985)

Aquaporin-9, a protein channel that can transport water and other solutes through the plasmalemma is expressed in rat GV-stage but not mature oocytes (Ford et al., 2000)



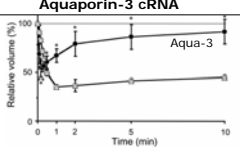
+



+

Permeability
Aquaporin-9
expression

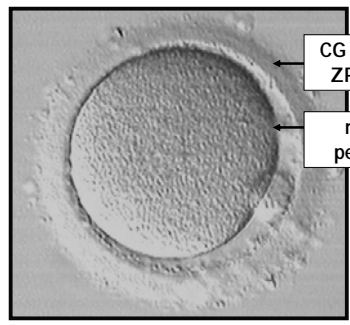
Osmotic response to glycerol of mouse oocytes injected with Aquaporin-3 cRNA



Edashige et al., 2003

g.en.e.r.a.

Possible injuries

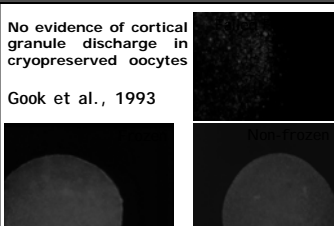


g.en.e.r.a.

Cortical granules release

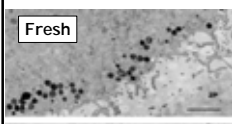
No evidence of cortical granule discharge in cryopreserved oocytes

Gook et al., 1993

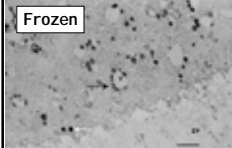


“The immunostaining examination for CG of the frozen–thawed oocytes did not reveal evidence of the premature release of CG.”

Li et al., 2005



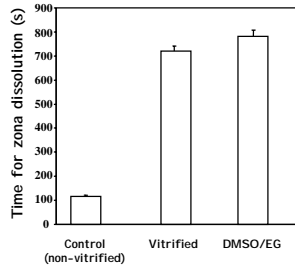
Fresh



Frozen

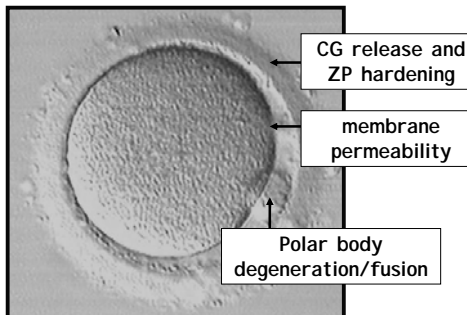
Ghetler et al., 2006

Zona Pellucida Hardening



Larman et al., 2006 n = greater than 60 oocytes per treatment with 3 replicates

Possible injuries



Aneuploidy and PB retention

Early reports on failure of PBII extrusion and increase of aneuploidy in thawed mouse oocytes

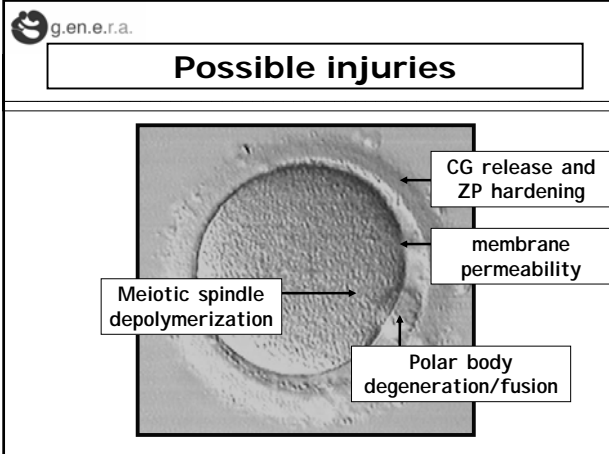


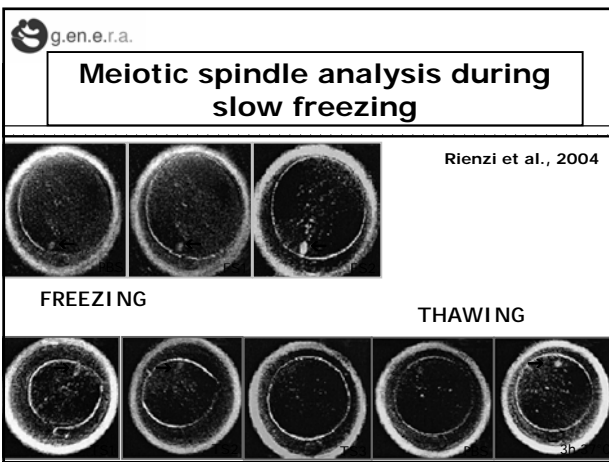
Glenister et al, 1987; Carroll et al., 1989

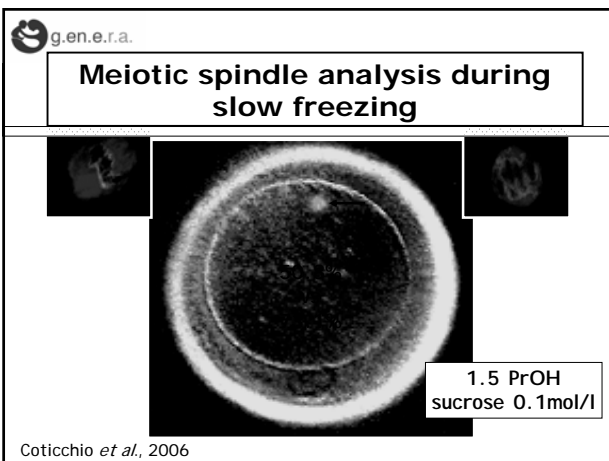
| Frozen | No. of Oocytes (%) | | |
|--------|--------------------|--------------|----------------|
| | Scored | % Aneuploidy | % Retention PB |
| + | 352 | 6.4 | 2.6 |
| - | 218 | 8.0 | 4.4 |

No increase in the rates of aneuploidy/digyny in parthenogenetically activated mouse oocytes after cryopreservation with DMSO/slow freezing

Bos-Mikich and Whittingham, 1995







g.en.e.r.a.

Possible injuries

Cytoplasmic and Cytoskeleton damage

Meiotic spindle depolymerization

CG release and ZP hardening

membrane permeability

Polar body degeneration/fusion

g.en.e.r.a.

Osmotic toxicity

Coticchio et al., 2004

SLOW FREEZING

VITRIFICATION

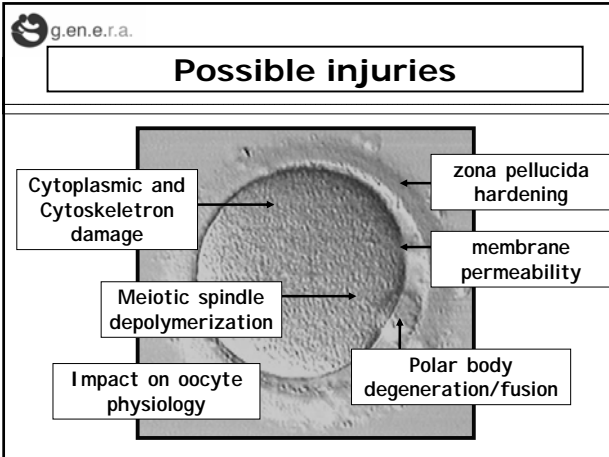
g.en.e.r.a.

Osmotic toxicity

OOCYTE OSMOTIC TOLERANCE AND OOLEMMA PERMEABILITY

- Temperature of exposure influence shrinking (swelling) patterns
- Oocyte shrinkage tolerance is about 30% of their initial volume
- At 22°C, EG has a lower permeability coefficient relative to DMSO and PG
- The membrane is more selective for EG and DMSO than for PG (mean reflection coefficient Sigma lower for PG)
- Permeability coefficients of individual oocytes varied substantially (inherent biological variability)

Van den Abbeel et al., 2007



g.en.e.r.a.

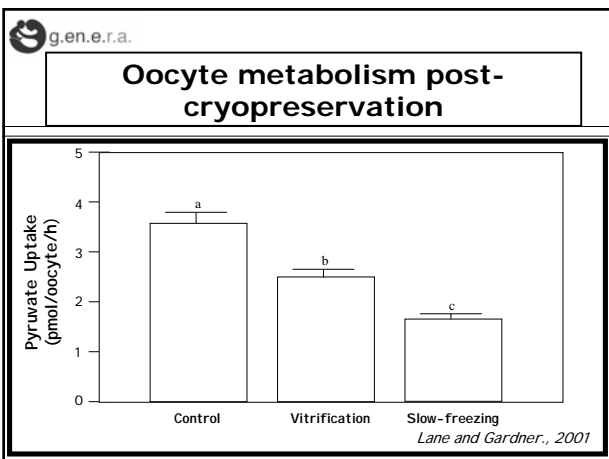
Oocyte metabolism post-cryopreservation

METABOLISM MONITORING THROUGH PYRUVATE UPTAKE (mouse oocytes):

Mouse oocytes and developing embryos following slow freezing were metabolically impaired compared with those that were vitrified

...although vitrification was also associated with a decrease in nutrient utilization by the oocyte compared to controls the decrease was significantly smaller than that induced by slow freezing.

Lane and Gardner., 2001; Lane et al., 2002



Oocyte protein profile post-cryopreservation

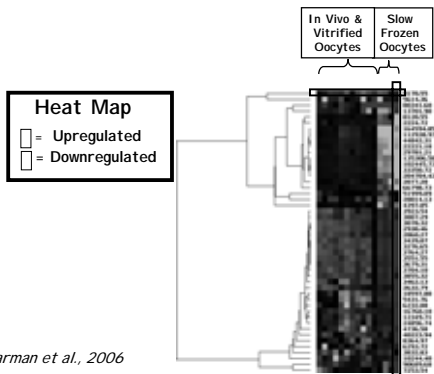
PROTEOMIC ANALYSIS OF OOCYTE PROTEIN PROFILES (mouse oocytes) by SELDI-TOF MS:

Mouse oocytes following slow freezing revealed major alterations compared with those that were vitrified.

Vitrified oocytes appeared to be similar to the non-cryopreserved control oocytes...

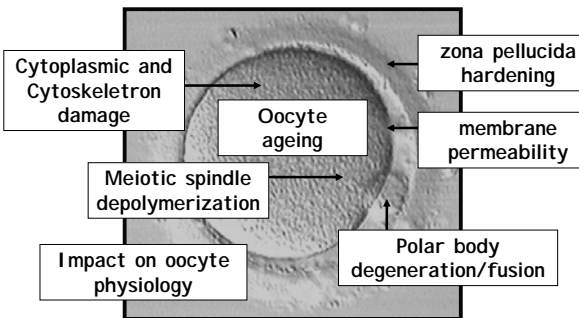
Larman et al., 2006

Hierarchical Clustering of Anionic Protein Profile



Larman et al., 2006

Possible injuries



Learning objectives

1) **SAFETY**

- Possible injuries to the oocyte during cryopreservation
- Possible contamination of the oocyte

2) **EFFICIENCY. WHERE ARE WE?**

- oocyte cryosurvival
- oocyte/embryo development post vitrification

3) **HOW TO IMPROVE IT?**

- Factors that may influence the efficiency

Principles of vitrification

1. High levels of cryoprotectants.
2. Extremely fast rates of cooling.
3. No ice crystal formation or damage;
straight to a glass.

TOOLS

Small volume is necessary for extremely rapid cooling rate (about 20,000°C/min).

- Electron microscopic grids
 - Open-pulled straw
 - Stripper trip
 - Cryoloop
 - Hemi-straw system
 - Cryotop
 - Cryoleaf
- } OPEN SYSTEM
- Cryotip
 - ...

Direct contact with nitrogen

OOCYTE CONTAMINATION:

- Not sterile procedure
- Liquid nitrogen may be contaminated by the surface of straws/cryovials or other tools
- Risk of liquid nitrogen mediated disease transmission

Tedder et al., 1995; Fountain et al., 1997; Berry et al., 1998

SOLUTIONS:

- Use of sealed system to avoid direct contact (ref)
- Cooling in liquid nitrogen vapours

Larman et al. 2006, Cobo et al., 2007

Learning objectives

1) SAFETY

- Possible injuries to the oocyte during cryopreservation
- Possible contamination of the oocyte

2) EFFICIENCY, WHERE ARE WE?

- oocyte cryosurvival
- oocyte/embryo development post vitrification

3) HOW TO IMPROVE IT?

- Factors that may influence the efficiency

Results of vitrification

| Author | Study | Patients | Clinical pregnancies | Abortions | Ongoing pregnancies | Gestational sacs | Freezing protocol |
|-----------------|-------|----------|----------------------|-----------|---------------------|------------------|-------------------|
| Kuleshova, 1999 | ICSI | 4 | 1 | 0 | 1 | 1 | VF |
| Chu, 1999 | ICSI | 1 | 1 | 0 | 1 | 1 | VF |
| Yoon, 2003 | ICSI | 34 | 6 | 0 | 6 | 7 | VF |
| Katayama, 2003 | ICSI | 2 | 2 | 0 | 2 | 2 | VF |
| Chian, 2005 | ICSI | 25 | 11 | | | | VF |
| Kim, 2005 | ICSI | 13 | 7 | | | | VF |
| Ruvacaba, 2005 | ICSI | NA | 8 | | | | VF |
| Okimura, 2005 | ICSI | NA | 12 | | | | VF |
| Lucena, 2006 | ICSI | 73 | 13 | | | | VF |
| Kurayama, 2005 | ICSI | 29 | 12 | | | | VF |
| Kyono, 2005 | ICSI | 1 | 1 | 0 | 1 | 2 | VF CRYOTOP |
| Selman, 2006 | ICSI | 7 | 2 | 0 | 2 | 2 | VF OPS |
| Yoon, 2007 | ICSI | 28 | 13 | 2 | 11 | 17 | VT SN2 grids |
| Cobo, 2007 | ICSI | 23 | 14 | 3 | 11 | 20 | VF CRYOTOP |
| Antinori, 2007 | ICSI | 120 | 36 | 8 | 28 | 39 | VF CRYOTOP |
| Chang, 2008 | ICSI | 2 | 2 | 0 | 2 | 3 | VF CRYOTOP |
| Chen, 2008 | ICSI | 1 | 1 | 1 | 1 | 2 | VF |

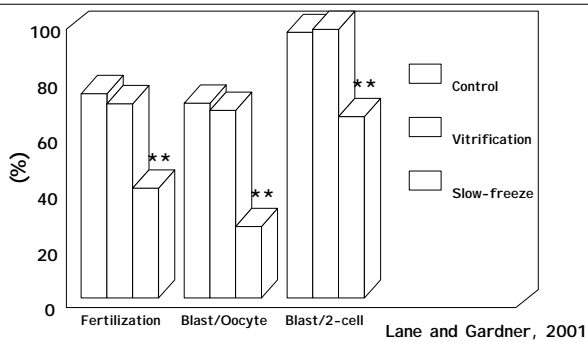
117 ongoing pregnancies/deliveries
149 sacs/live births

Results of vitrification

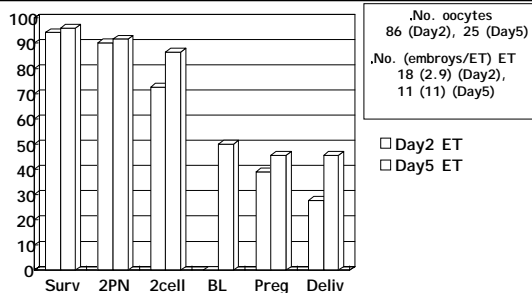
| Variable | Slow Freezing literature 1996-2005 | Vitrification literature 2003-2005 |
|---|------------------------------------|------------------------------------|
| Age, mean | 33.7 | 32.3 |
| Fertilization rate | 64.9 (2,478/3,818) | 74.2 (637/859) |
| Clinical pregnancies per thawed oocyte | 2.3×10^{-2} (153/6720) | 4.5×10^{-2} (61/1354) |
| Clinical Pregnancies per injected oocytes | 4.0×10^{-2} (153/3818) | 7.2×10^{-2} (61/859) |
| Clinical Pregnancies per transfer | 20.6 (153/742) | 45.5 (61/134) |
| Implantation rate | 10.1 (185/1828) | 17.2 (81/473) |

Oktaý *et al.*, 2006

Mouse oocyte vitrification



Masa Kuwayama, ESHRE, 2006



Learning objectives

1) **SAFETY**

- Possible injuries to the oocyte during cryopreservation
- Possible contamination of the oocyte

2) **EFFICIENCY, WHERE ARE WE?**

- oocyte cryosurvival
- oocyte/embryo development post vitrification

3) **HOW TO IMPROVE IT?**

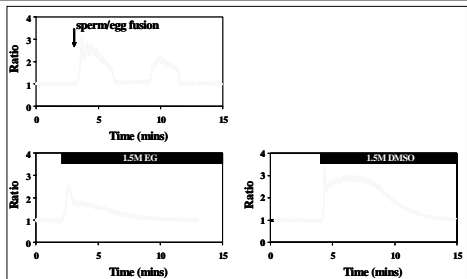
- Factors that may influence the efficiency

Factors that may influence oocyte vitrification efficiency

1) **CRYOPROTECTANTS** and intracellular Ca^{2+}

2) **TEMPERATURE** and meiotic spindle

Cryoprotectant and intracellular Ca^{2+}



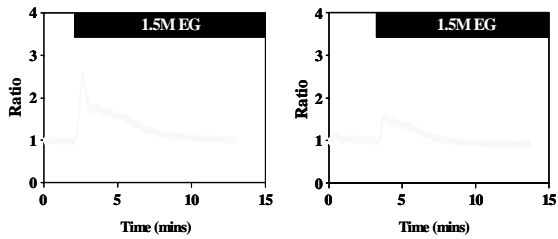
Larman et al., 2006

Cryoprotectant and intracellular Ca²⁺

Increase in intracellular Ca²⁺ triggers activation:

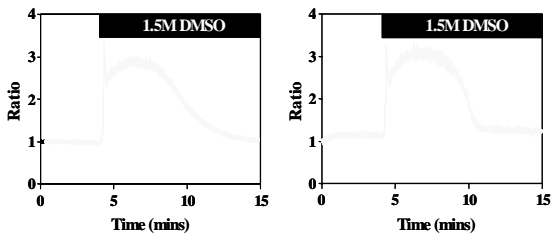
- Block to polyspermy
- Completion of meiosis and start of mitotic divisions
- Down-regulation of cell cycle proteins
- Apoptosis

Effect of removing extracellular Ca²⁺ on EG challenge



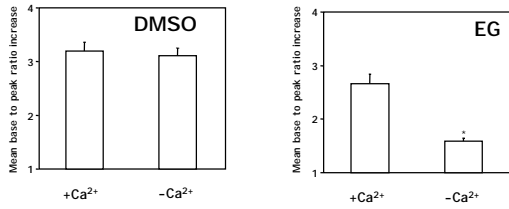
Larman et al., 2006

Effect of removing extracellular Ca²⁺ on DMSO challenge



Larman et al., 2006

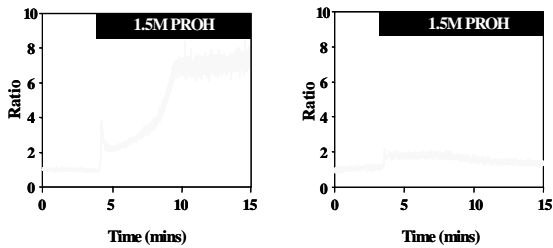
Effect of removing extracellular Ca²⁺ on cryoprotectant challenge



n = 12 for each treatment with 4 replicates *; p = <0.01

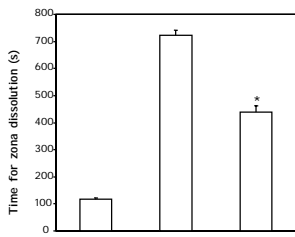
Larman et al., 2006

Effect of removing extracellular Ca²⁺ on PROPANDIOL challenge



Larman et al., 2006

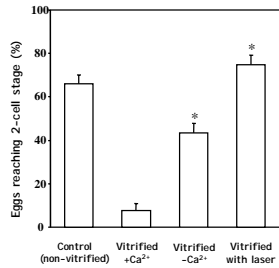
Effect of removing extracellular Ca²⁺ on ZONA HARDENING



Larman et al., 2006

n = greater than 60 oocytes per treatment with 3 replicates *; p = <0.01

Effect of removing extracellular Ca²⁺ on FERTILIZATION RATE



Larman et al., 2006
n = greater than 200 oocytes per treatment with 3 replicates *; p < 0.01

Factors that may influence oocyte vitrification efficiency

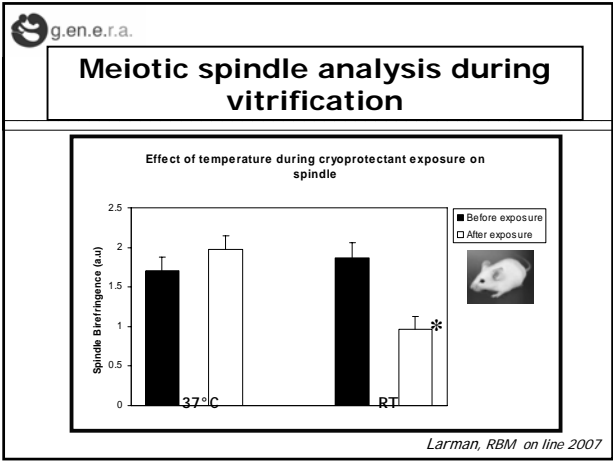
2) TEMPERATURE and meiotic spindle

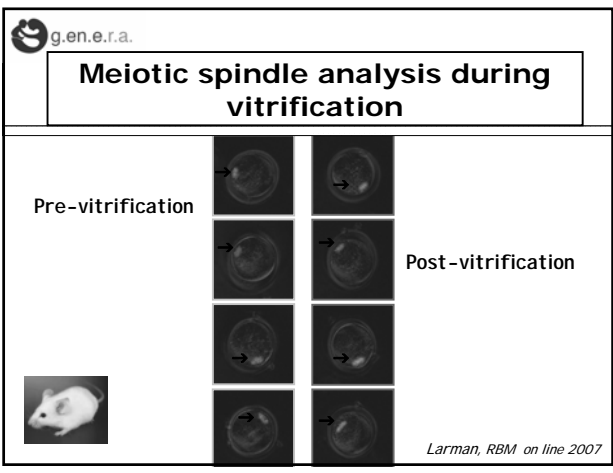
Meiotic spindle analysis during vitrification

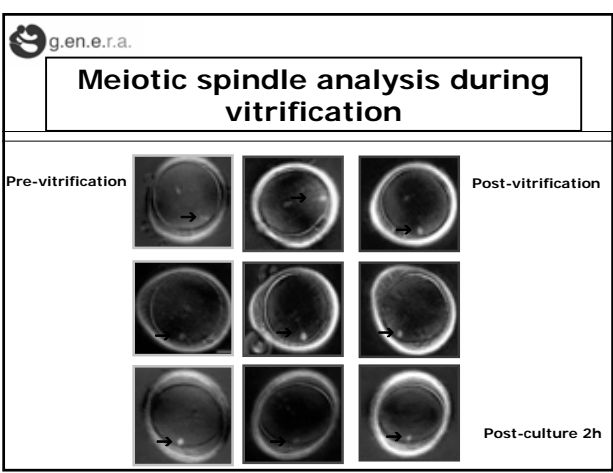
MEIOTIC SPINDLE AND VITRIFICATION TEMPERATURE:

Non invasive visualization of the meiotic spindle in living mouse oocytes following cryoprotectant exposure at room temperature (RT) and 37°C.

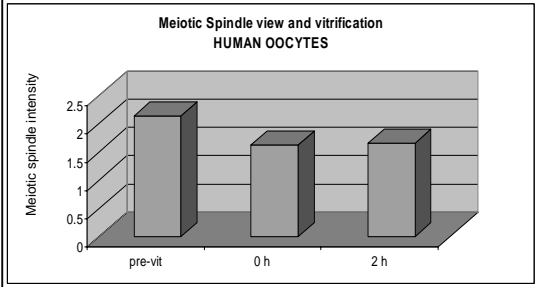
Non invasive visualization of the meiotic spindle in living mouse and human oocytes following vitrification procedure performed at 37°C.







Meiotic spindle analysis during vitrification



Larman, RBM on line 2007

CONCLUSIONS

Oocyte cryopreservation poses certainly specific problems:

- The oolemma and not the size of MII oocyte is the key to explain the low survival rates obtained with slow freezing.
- Release of cortical granules (controversial)
- Chemical toxicity from cryoprotectants (type specific)
- Osmotic toxicity
- Meiotic spindle depolymerization
- Oocyte physiology alteration (metabolism and protein profile)

CONCLUSIONS

Safe and Efficient vitrification methods for human oocytes: how to do it?

- The actual vitrification methods seems already to offer high survival rate. Deriving embryo viability has however to be determined on larger scale.
- The choice of the cryoprotectant is very important to minimize oocyte damage and activation.
- To reduce MS damage, vitrification procedure has been proposed to be performed at 37°C.



CLINICA VALLE GIULIA, Rome



ginecologia
endocrinologia
embriologia
riproduzione
assistita



Specialisti:
Filippo Maria Nicolini
Stefano Bernardini
Stefano Cerasola



Laboratori:
Laura Bernardini
Stefania Bernardini
Stefania Bernardini
