



Ejaculate analysis

Semen volume

Sperm motility

Sperm vitality

Sperm concentration

Sperm morphology

Reference values

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Sperm motility

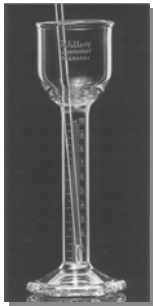
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Semen volume: WHO5-recommended methods



Direct volume measurement in collection vessel

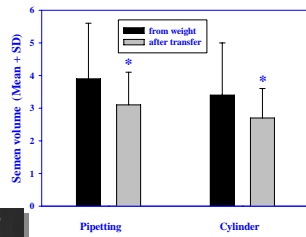


Indirect volume measurement from weight, assuming semen density of 1 g/ml (it is 1.014 g/ml)

- Weigh the vessel (with label) (g)
- Weigh the vessel with semen (g)
- Calculate the weight difference (g = ml)

Semen volume: WHO5-not recommended methods

Transfer from the collection vessel



Transfer from the collection vessel by pipetting or decanting to a cylinder leads to:

- falsely low volumes
- -0.4 to -0.8 ml (14%)

First microscopic evaluation

Liquefaction at RT or 37°C (30 min)

Phase contrast microscope ± heated (37°C) stage



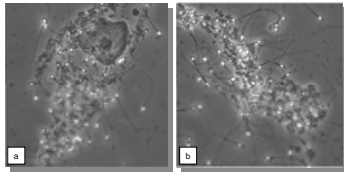
Positive-displacement pipette, 10 µl, 22 mm x 22 mm coverslip



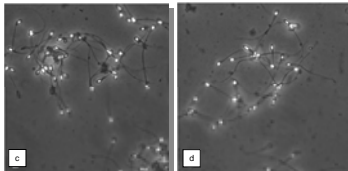
- Aggregation, Agglutination
- Non-sperm cells
- Judge dilution for sperm concentration
- Sperm motility

Sperm aggregation and agglutination

Aggregation - immotile sperm bind to each other
 motile sperm bind to non-sperm cells or debris



Agglutination - motile sperm bind to other motile sperm



Antisperm antibodies?

WHOS: Degrees of sperm agglutination

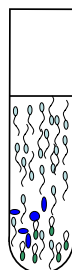
Pairs involved	Degree of agglutination			
	1. Isolated (x 10 sperm) agglutinate, many free sperm)	2. Moderate (10-50 sperm) agglutinate, free sperm)	3. Large (agglutinate > 50 sperm, some sperm still free)	4. Gross (all sperm aggluti- nated, and agglu- tations intercon- nected)
a. Head to head				
b. Tail-to-tail				
c. Head-to-head and tail-to-tail				
d. Mixed (some head-to-head and tail-to-tail agglutinations)				

Semen analysis: representative aliquots

Semen is viscous

Area 1 high sperm concentration
 fast sperm
 good morphology

Area 2 low sperm concentration
 slow sperm
 bad morphology



- Mix the sample well
- Take 2 aliquots
- Assess 2 x 200 sperm (when possible)
- Compare duplicates

Poisson Distribution (sperm/leukocyte numbers)
 Binomial Distribution (% Motility, % Morphology, % Vitality)

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Sperm motility: chamber depth

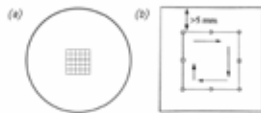
Assess at RT or 37°C in ~20 µm deep chambers, to permit free motion

Depth (D, µm) is volume (V, µl = mm³) divided by area (A, mm²)

22 x 22 10 µl	18 x 18 6.5 µl	21 x 26 11 µl
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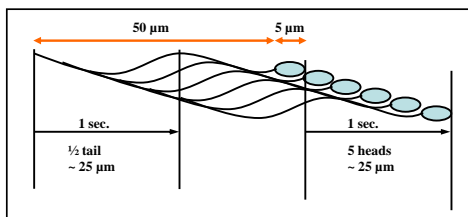
Assess 2 x 200 sperm (if possible) in at least 5 different fields

use an eyepiece graticule to limit the area examined
avoid the edges of the slide

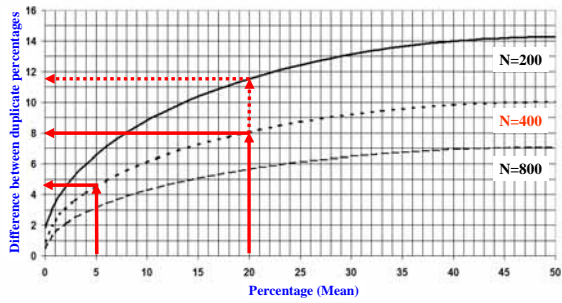


Sperm motility: grading

Is assessed as:	WHO4	WHOS
Immotile	Grade d	IM
Non-progressive	Grade c	NP
Progressive		PR
slow	Grade b	
fast (> 25 µm/s at 37°C)	Grade a	



% errors: Binomial Distribution



e.g. for 400 assessed sperm: the difference between duplicates for a mean of 20% should be $\leq 8\%$; for a mean of 5% should be $\leq 5\%$
 If >this, prepare and assess two more wet preparations

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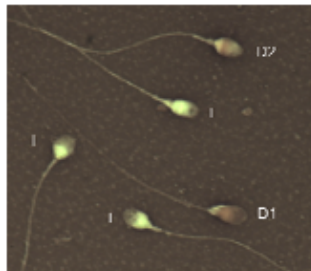
WHO5 sperm vitality: one-step method

If <40% PR, detection of intact sperm head membranes

Intact sperm head membrane: impermeant dyes excluded

e.g. Eosin-Nigrosin
 Eosin = impermeant dye
 Nigrosin = background

- Red head = Dead (D1)
- Pink head = Dead (D2)
- White head = Alive (L)
- Pink neck = Alive (L)



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WHO5: sperm counting chambers

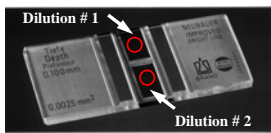
Not recommended:

shallow, small volume chambers: too few sperm
capillary-filled chambers: uneven filling
unfixed samples: motile sperm

Recommended:

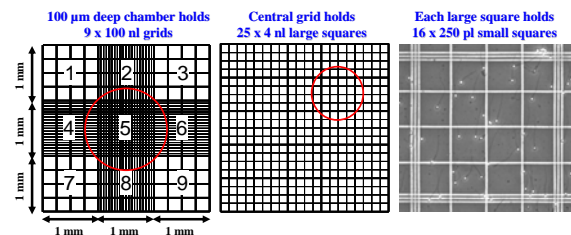
haemocytometer (100 µm deep)
fixed, diluted samples

Sperm concentration: Neubauer Improved Chamber



- Make duplicates from two separate dilutions
- Do not fill both chambers from one dilution
- Do not count the same chamber twice

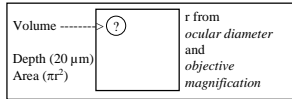
- Sampling error
- Pipetting error
- Diluting error



Sperm concentration: semen dilutions

Dilution for the exact measurement is based on a rough assessment of the number of sperm in the volume observed

The volume observed depends on the area of the microscopic field and the depth of the chamber



For a x40 objective and x10 ocular of 20 mm aperture

The HPF has a diameter of 20 mm/40 = 500 μm, the radius r = 250 μm

So $r^2 = 62500 \mu\text{m}^2$ and the area = $\pi r^2 = 196,375 \mu\text{m}^2$

The volume = area x depth = $196,375 \mu\text{m}^2 \times 20 \mu\text{m} = 4,064,962 \mu\text{m}^3 = 4 \text{ nl}$

Sperm concentration: semen dilutions

With 50 sperm per HPF (4 nl)
there are 12.5 per nl = 12,500 per μl = 12,500,000 per ml

Dilute 1:20 as recommended in WHO4 = 625,000 per ml

The central grid of the Neubauer chamber holds 100 nl
i.e. there are 63 sperm per grid
- not enough for an acceptable count (<200)

Dilute 1:5 as recommended in WHO5 = 2,500,000 per ml

The central grid of the Neubauer chamber holds 100 nl
i.e. there are 250 sperm per grid
- more than enough for an acceptable count (>200)

Sperm concentration: semen dilutions

	WHO4 More and higher	WHO5 fewer and lower
Dilutions	1+49 1+19 1+9 1+4	1+19 1+4 1+1
N/x400HPF	>200 40-200 15-40 <15	>100 16-100 2-15
Examine:	Central grid (#5) 5, 10, 25 small squares	Central 3 grids (#4, 5, 6) one row (20 nl)
	12 factors rarely 200 sperm	3 factors always 200 sperm

Which sperm to count?

Count all sperm within the central square

The middle line shows the relevant square

Preventing double counting

○ : counted
○ : not counted

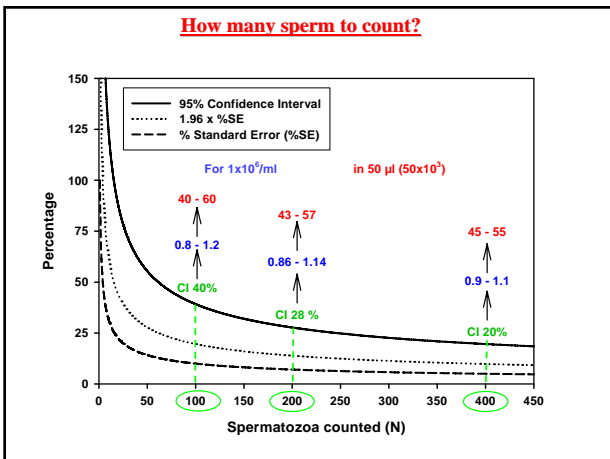
Which sperm to count?

Count sperm on lines only when they are the bottom or left lines ("L")

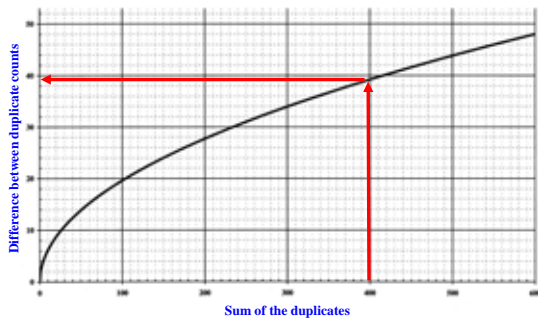
On the left line

On the bottom line

○ : counted
○ : not counted

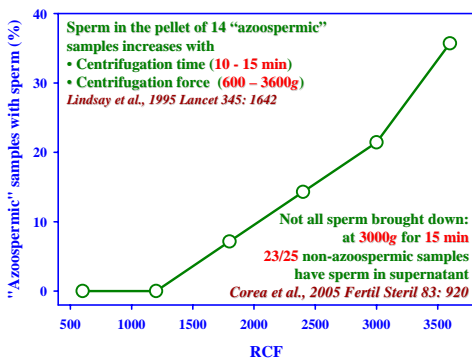


Counting error: Poisson Distribution



e.g. for 400 sperm assessed, acceptable difference is 39
If >39, make two more dilutions

Azoospermia: dependent on RCF



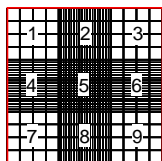
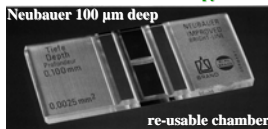
Centrifugation inaccurate!

Low sperm numbers: accurate assessment required

Dilute semen less and read larger volumes

Dilute semen 1+1 with Formalin
Phase contrast microscopy

Dilute semen 1+1 mit Formalin
+ Hoechst 33342 dye
Fluorescence microscopy



Read all 9 grids
= 900 nl

Sensitivity
56,000 per ml

Read all the chamber
= 25,000 nl

Sensitivity
2,000 per ml

If <25 spermatozoa observed, state:
"<56,000" per ml" or "<2,000 /ml"

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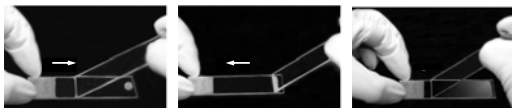
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Sperm morphology: semen smears



- Pull the semen droplet behind the slide
- Do not push it in front of it

Semen smear thickness depends on:

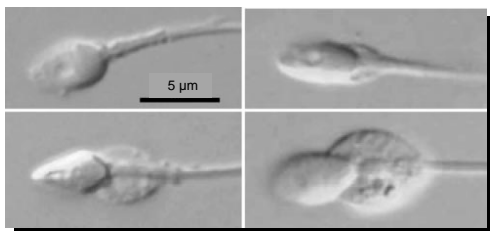
- aliquot volume (10 µl) smaller → separated sperm
- angle of dragging slide (45°) greater → thinner smear
- speed of smear (~1 sec) higher → thicker smear

Disadvantages of fixing air-dried semen smears:

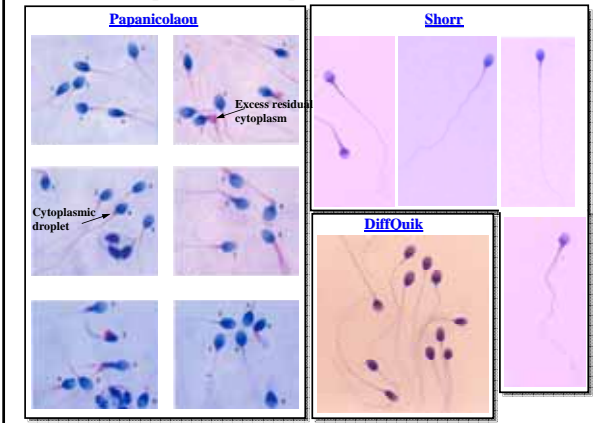
- dehydration smaller than in wet preparations
- shearing forces expansion of immature sperm heads
- osmotic insults loss of cytoplasmic droplet
- retention of excess residual cytoplasm

Sperm morphology: cytoplasmic droplets

- Cytoplasmic droplets (midpiece vesicles)
- Present on motile sperm in semen, mucus, medium
- Osmotically sensitive: shrink on air-drying

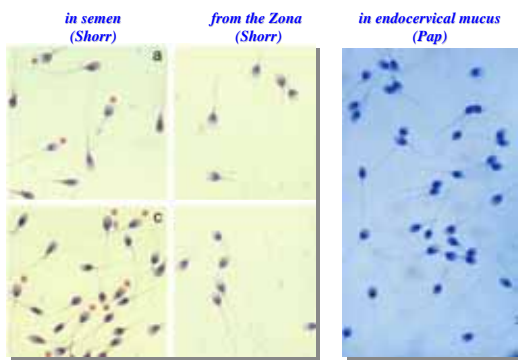


Sperm stains: Pap, Shorr, DiffQuik



“Normal Sperm“: concept

Identify a subpopulation of potentially fertilising spermatozoa (normal/ideal)
Biologically selected by cervical mucus or the zona pellucida



Sperm morphology

Normal heads?



Abnormal heads?



Normal tails?



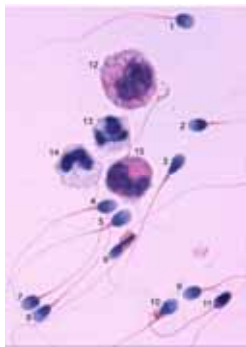
WHOS: sperm micrographs



WHOS: sperm descriptions

	Sperm	Head Shape	Other head comments	Midpiece	Principal piece	Overall	Comments
Plate 7	37	normal				normal	If PP OK
	38	abnormal	round			abnormal	
	39	normal				normal	
	40	normal				normal	
	41	normal				normal	
	42	normal		thick		abnormal	
	43	normal	<40 % acr			abnormal	
	44		out of focus				
	45	abnormal	round			abnormal	
	46	abnormal	round			abnormal	
	47	normal				normal	
	48	normal				normal	If PP OK

Spermatozoa and other non-sperm cells in semen



With Pap: immature germ cells more pinkish, polymorphs more bluish
Concentration: number relative to that of spermatozoa (conc. known)

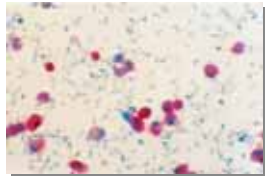
Other cell types in semen

Leucocytes

Peroxidase-positive
Leukocytes
Granulocytes



Pan Leukocyte marker
CD45 Antibody



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WHO5: Reference values for human semen

Reference Population

Fathers (Partners with time to pregnancy of ≤ 12 months)

N > 1600 3 continents 5 centres

Samples

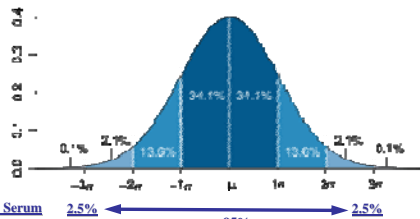
Only 2-7 days of abstinence
one sample per man
complete samples

Methods

collection, preparation, analysis with WHO methodology
IQC + EQC-controlled labs.
only haemocytometer (not CASA, Makler chambers)
Tygerberg morphology

Reference Distribution and Intervals

It is accepted that 95% of the values should lie between the reference intervals



Two-sided: Serum

One-sided: Semen

One-sided Reference Intervals:

- higher values irrelevant
- lower reference limit is the 5th percentile

WHO2-4 "Normal", "reference" values and WHO5 "lower reference limits"*

WHO	2	3	4	5
	1987	1992	1999	2007/8
	normal	normal	reference	LRL (5 PC)
Semen Vol. (ml)	2.0	2.0	2.0	1.5
Total number (M)	40	40	40	39
Sperm concn. (M/ml)	20	20	20	15
Progr. Motility (%)	50	50	50	28
Vitality (%)	50	75	75	59
Normal Forms (%)	50	30	(15)	3

*subject to confirmation by WHO

Summary

Changes to the WHO laboratory manual for the examination of human semen are aimed at:

- increasing the accuracy of analytical results
- providing more experimental details of common methods
- giving hints and details of what to do when QC results are poor

These should help improve:

- standardisation between labs
- the diagnostic value of semen analysis results
- follow up of therapeutic treatments



Low sperm numbers: no accurate assessment required

Rough estimation:
if <4 sperm per x400 HPF (<16 per x200 HPF)
state “< 2x10⁶ sperm /ml”

Are any sperm present?
Centrifuge 1 ml semen at 3000g for 15 min
Leave ~50µl supernatant
Place 2x10 µl pellet under 22mm x 22 mm coverslips (20 µm deep)
Examine both coverslips entirely (~2x480 fields)
If sperm found, state “cryptozoospermia”

Are any motile sperm present?
Place 40 µl semen under 24mm x 50mm coverslip (33 µm deep)
Examine coverslip entirely (~1200 fields)
If motile sperm found, state how many

N.B. All these procedures are inaccurate
Few sperm = large counting error
No sperm seen ≠ no sperm present
The 95% upper CI for 0 = 3.7
