09.00 – 09.30 The search for excellence in IVF laboratories: towards "the best" ESHRE Campus symposium Bologna, Italy 23-24 January 2009

Zygote morphology as a reflection of embryo polarity

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Learning Objectives

- Define polarity in mammalian oocytes and embryos
- Define zygote morphology, what it means biologically
- Relate polarity and zygote morphology to continued development
- Discuss how Polarity and Zygote morphology can affect clinical outcome



Published Literature

- *Gardner 1996/ Edwards 1997* suggested that it does exist in oocytes which dictates all further polarity and axes and that an animal-vegetal pole exists, directed by the polar body position
- Zernika-Goetz stated "there are no essential components that are localized uniquely to the animal or the vegetal pole 1998
- Zernika-Goetz, 2002 suggest that the axis is set up by Sperm Entry site = first axis
- *Hiiragi and Solter, 2004, 2005:* Alternately, it is the plane separating the 2 pronuclei as they move to the center of the oocyte which sets up the first axis (in the mouse)
- Hansis and Edwards, 2003/2005: The only report on polarity in human 4-cell embryos, not repeated, but its not related to the oocyte or zygote

Empiric observations

- Uneven cleavage at the first mitotic division is not compatible with delivery
- Muti-nucleation in blastomeres at the 2-4 cell stage is not compatible with delivery
- Off-Center Nuclei at the 1 cell stage is not compatible with delivery





Investigating Human Oocyte Polarity

- Look at human oocytes and embryos and elucidate impact of any observed polarity in cleavage on outcome using common/ known parameters of determining embryo morphology as predictors of outcome measures.
- The ideal method is Time-Lapse videos or frequent sequential still observations
- Relate this to known morphological features that impact outcome

Time Lapse of Human Oocytes and Embryos

- Immature GV and M1 oocytes, fresh
- 1PN and 3 PN abnormal fertilized oocytes, fresh
- Thawed 2PN embryos donated to research
 Thawed 2-8 cell embryos, donated to research
- 2 Systems were utilized:
- Octax, in continual streaming video within a closed chamber
 Unisense-EmbryoScope in a dedicated incubator system with time lapse.
- All Oocytes and Embryos were set up as individuals in both systems and monitored for 24-120 hours continually

GV to M1 Maturation

- 15/19 GV oocytes matured to MI with
- Disillusion of the GV membranes, considerable movement of the cytoplasm, including a pulsing as the oocyte appeared to slightly shrink and reexpand
- Polarity? It would be hard to tell unless there were fluorescent markers to indicate a top and bottom in the oocyte

MI to MII

28/35 MI matured to MII accompanied by

- ➤ the extrusion of the polar body (PB)
- > which was accompanied by deep shrinking followed by expansion, but never to its original size, with formation of the perivitaline space.
- Where the oocyte did not shrink, the PB was small or appeared to be sequestered into the oocyte (Abnormal??)







Origin of Centrioles

- Mice: Centrioles are formed in the oocyte
- In humans they come from the sperm
- Can this be a source of difference in patterning and polarity between mouse and human oocytes?
 - » Review Chatzimeletiou, K et al. 2008































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2 PN Oocytes

- Cleavage was generally on the Pronuclear Plane
- The first cleavage division is very dynamic and can only be observed with *rapid* time-lapse (short interval) or continual video
- Blastomeres undergo pulsing, blebbing and rearrangement but settle into a 2-cell embryo by +/-2 hours
- The polar body rapidly rotates into the cleavage plane
- This pattern is repeated at the second/third division and then no more
- At the 2 cell stage, multinucleated blastomeres can explode into multiple cells



Polarity: Relevance to ART?

(this will be more evident in the second lecture)

- MII Oocytes with abnormal PB and no PVS are likely to be dysfunctional even after fertilization
- Fertilized oocytes with PNs not centrally located are abnormal, can not adhere to polarity planes of cleavage
- Day 2 scoring is essential (in the absence of time-lapse) to eliminate cleavage errors
- The timing of Day 2 scoring is important and needs to be empirically defined per lab due to the dynamic nature of the first mitotic division

Fertilized Oocyte Scoring-Zygote Morphology

Looks at a part of nucleoli in the early embryo

Nucleoli

- > Found in all actively dividing cells
- \succ Sites of rRNA synthesis
- > Develop on the DNA where the genes for ribosomes are located, rDNA
- These points on the DNA = NORs
 Nucleolar Organizing Regions

Nucleoli

- During mitotic cell cycles the nucleoli fuse as the chromatin condenses
- Between 5-7 in human cells
- Always equality between mitotic daughter cells- size, number, fusion patterns
- Nucleoli are sites of protein synthesis, some mitogenic factors and growth regulatory proteins
- In oocytes the nucleoli are disaggregated at the GV stage and reform after fertilization during the first 2-3 mitotic divisions

Human NORs

- 5 pairs of NOR-bearing chromosomes
 13, 14, 15, 21, 22 (acrocentric)
- * Generally 5-7 NORs in human cells
- Activation of the NORs induces pol 1 transcription
 NOR's are clustered and this is dependent on
- Nors are clustered and this is dependent on heterochromatin adjacent to rDNA genes
 Heterochromatin is *not* inactive and may be
- involved in developmental control (Dimitri, 2004)
- Transcription of rDNA results in 3 functional parts:

- Dense fibrillar component (DFC)
 - Required for transcription of DNA
- Pre-rRNA + proteins
 Fibrillar component (FC)
 - <u>Structural center</u> for transcription, storage of inactive transcription factors
 - Surrounded by DFC
- Granular component (GC)
 - Pre-ribosomal particles- cytoplasmic





Sperm and NPB's

- Sertoli cells are very mitotically active
- This means they have de-constructed nucleoli during mitosis
- There are +/- 6 NPB (FC regions) per daughter cell easily visible
- This is what should be brought to the oocyte in the sperm



Male factor identified by PN morphology

































Conclusions

- PN morphology may reflect the state of chromatin condensation onto the spindle
- May reflect the cell cycle and synchrony of the male and female nuclei
- Has a direct impact on early scoring parameters and blastocyst developmet
- Is correlated with euploidy/aneuploidy
- Has an impact on delivery rates