

Carbohydrate & Energy Metabolism During Follicle Growth & Oocyte Maturation

What Can Be Learned From IVM & Follicle Culture?

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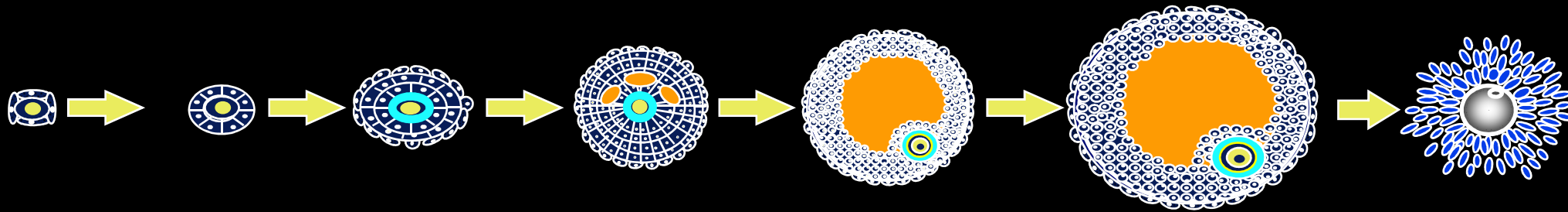
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The Biology Of Follicle & Oocyte Development

Primordial follicle (30-40 μm) **Primary follicle** (50 μm) **Secondary follicle** (50-200 μm) **Pre antral-antral follicle** (>220 μm) **Early antral-antral follicle** (0.5-2.0 mm) **Graafian follicle** (2.0- \geq 17 mm) **Ovulation**



A-cyclic recruitment

Cyclic recruitment

Selection

Gonadotrophin responsive

Gonadotrophin dependent

Molecular regulation: transcription factors, growth factors, peptides, steroids

Cell-cell interactions & signalling for follicle & oocyte growth & development

Follicular Fluid And Granulosa Cell Markers Of Follicle And Oocyte Development

Nutrients
Growth factors
Hormones
Oxygen tension
Granulosa cells



1.5 mm Antral Follicle

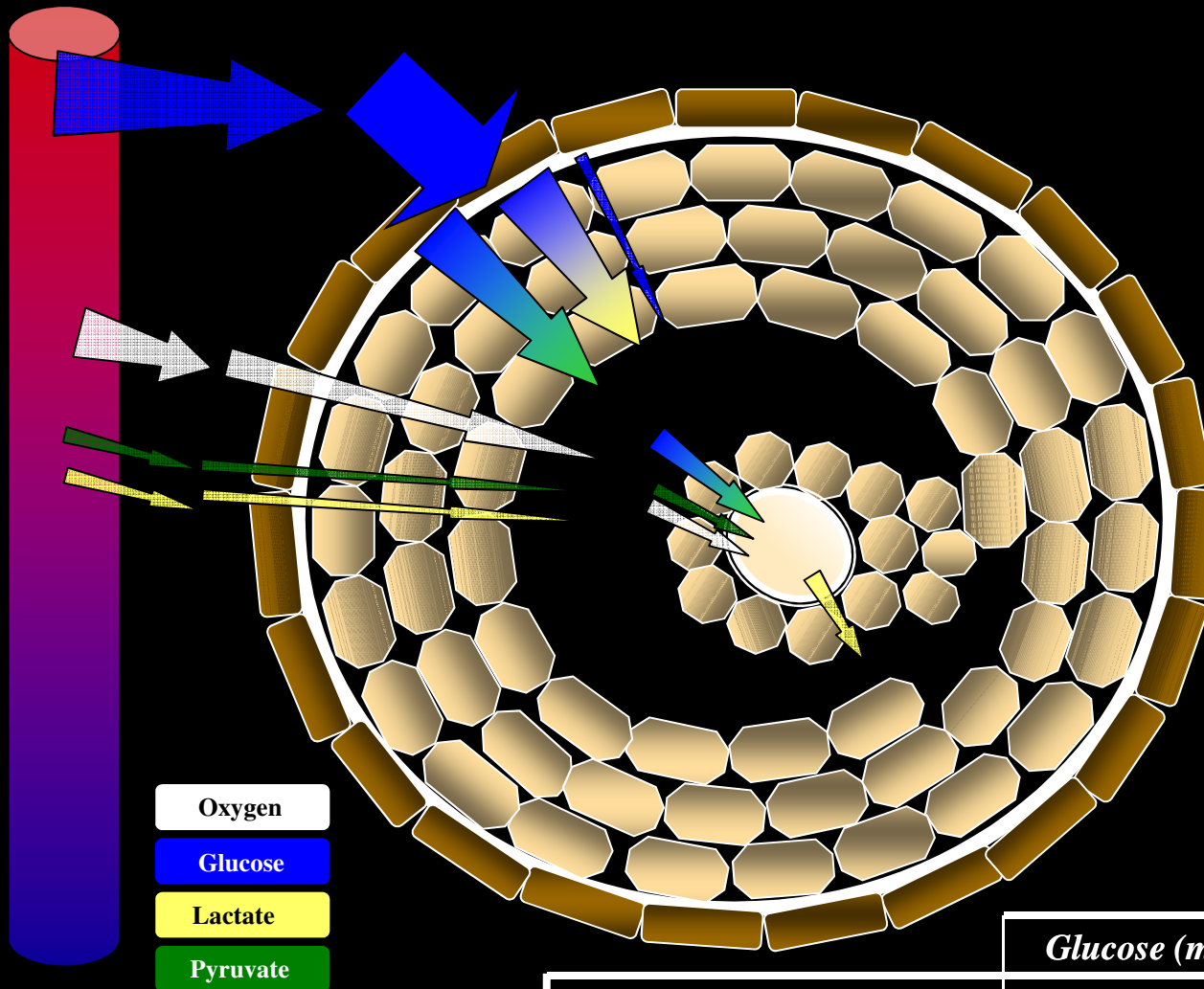


18 mm Ovulatory Follicle

Metabolic requirements change as:

- i) Follicle size and vascularity \uparrow
- ii) Follicle differentiation \uparrow
- iii) Follicle exposure to LH/ hCG

Nutrient Origin & Consumption During Follicle & Oocyte Growth *In Vivo*



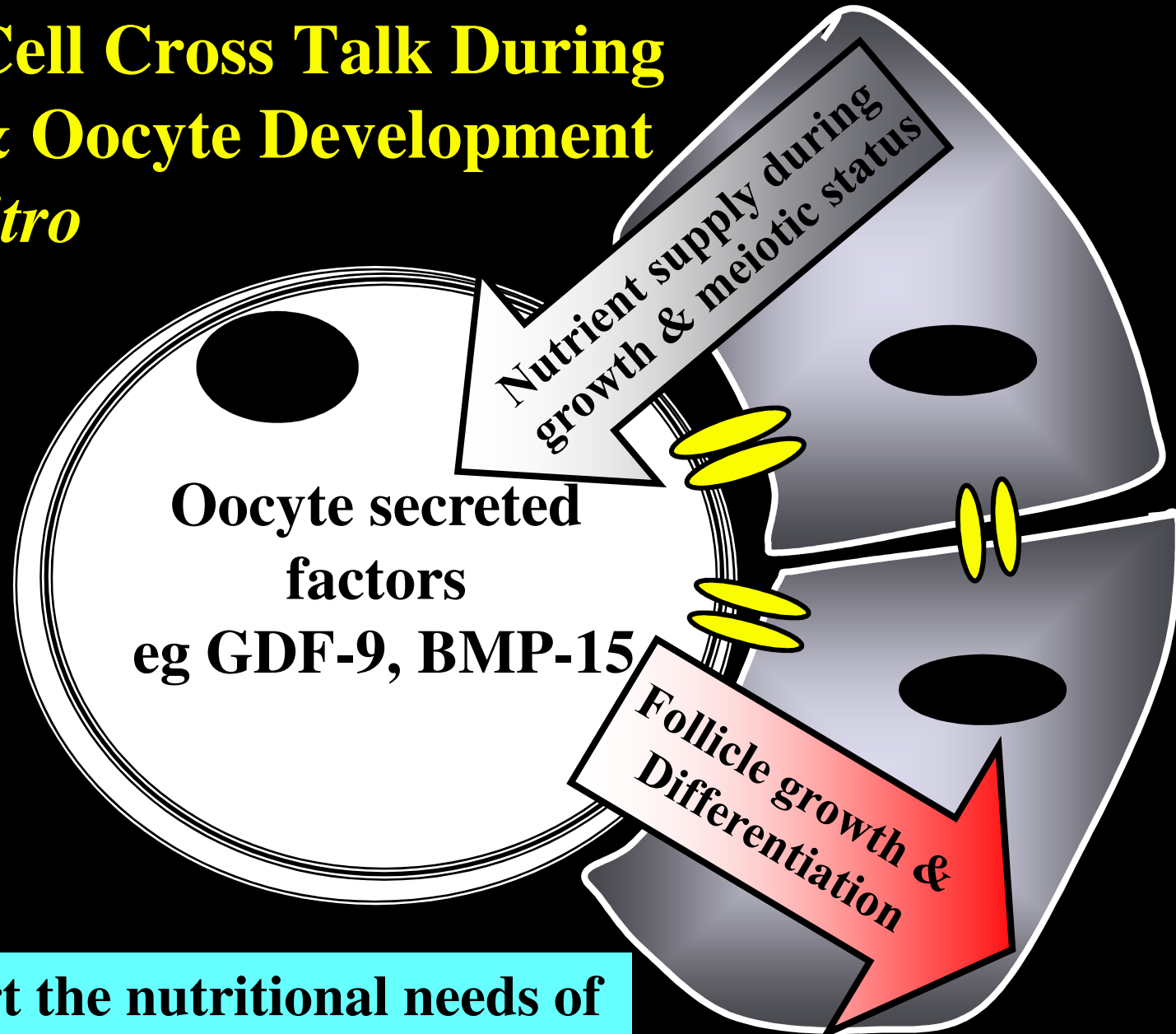
	<i>Glucose (mM)</i>	<i>Pyruvate (mM)</i>	<i>L-Lactate (mM)</i>
<i>Follicular fluid (mouse)</i>	0.46	0.38	17.3
<i>Oviduct fluid (mouse)</i>	1.09 (+ CCs)	0.37 (+ CCs)	10.9 (+ CCs)
<i>Plasma (mouse)</i>	11.7	0.16	4.8
<i>Follicular fluid (human)</i>	3	0.26	6.06
<i>Oviduct fluid (human)</i>	0.5-3.11	0.24-0.32	4.87-10.55
<i>Plasma (human)</i>	5	0.1	0.6

Nutrient Composition Of Body Fluids

Fluid Source	Glucose (mmol l ⁻¹)	Lactate (mmol l ⁻¹)	Pyruvate (mmol l ⁻¹)	Species
	3 - 3.39	3.17- 6.06	0.26	Human (Leese & Lenton, 1990; Gull et al. 1999)
	0.46	17.3	0.38	Mouse (Harris et al. 2005)
	4.8	5.	0.03	Cow (Orsi et al. 2005)
Oviduct	0.5 - 3.11	4.87 - 10.55	0.24	Human (Gardner et al. 1996)
	0.53 - 1.1	5.4 - 8.58	0.14 - 0.17	Human (Tay et al. 1997, Dickens et al 1995)
	0.59	5.71	-	Pig (Nichol et al. 1992)
	0.07	-	-	Cow (Carlson et al. 1970)
	1.65	11.7	0.17	Mouse (Harris et al. 2005)
Plasma	5	0.6	0.1	Human (Borland et al. 1980)
Blood	3.5 – 11.7	0.56 - 5.64	0.16-0.18	Mouse (Wang et al. 2003, Sanni et al. 2001, Harris et al. 2005)

Oocyte-Somatic Cell Cross Talk During Normal Follicle & Oocyte Development *In Vivo* And *In Vitro*

Bidirectional communication between the oocyte & GCs via **gap junctions**



Gap junctions support the nutritional needs of follicle & oocyte growth *in vivo* & *in vitro*?

The Biology Of Follicle & Oocyte Development

Primordial follicle
(30-40 μm)

Primary follicle
(50 μm)

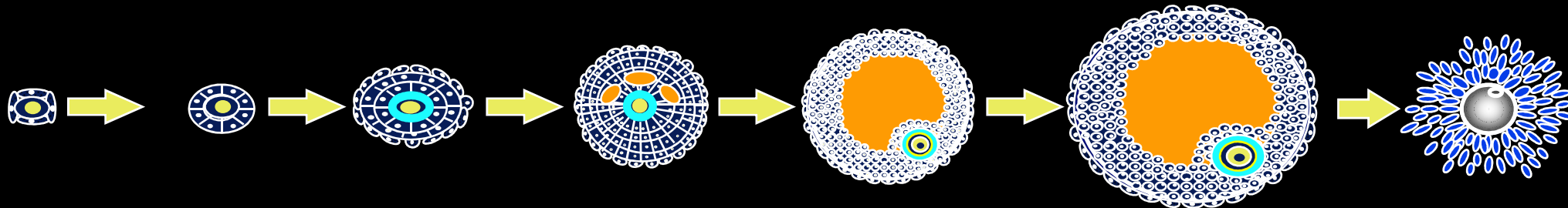
Secondary follicle
(50-200 μm)

Pre antral-antral follicle
($>220 \mu\text{m}$)

Early antral-antral follicle
(0.5-2.0 mm)

Graafian follicle
(2.0- $\geq 17 \text{ mm}$)

Ovulation



A-cyclic recruitment

Cyclic recruitment

Selection

Gonadotrophin responsive

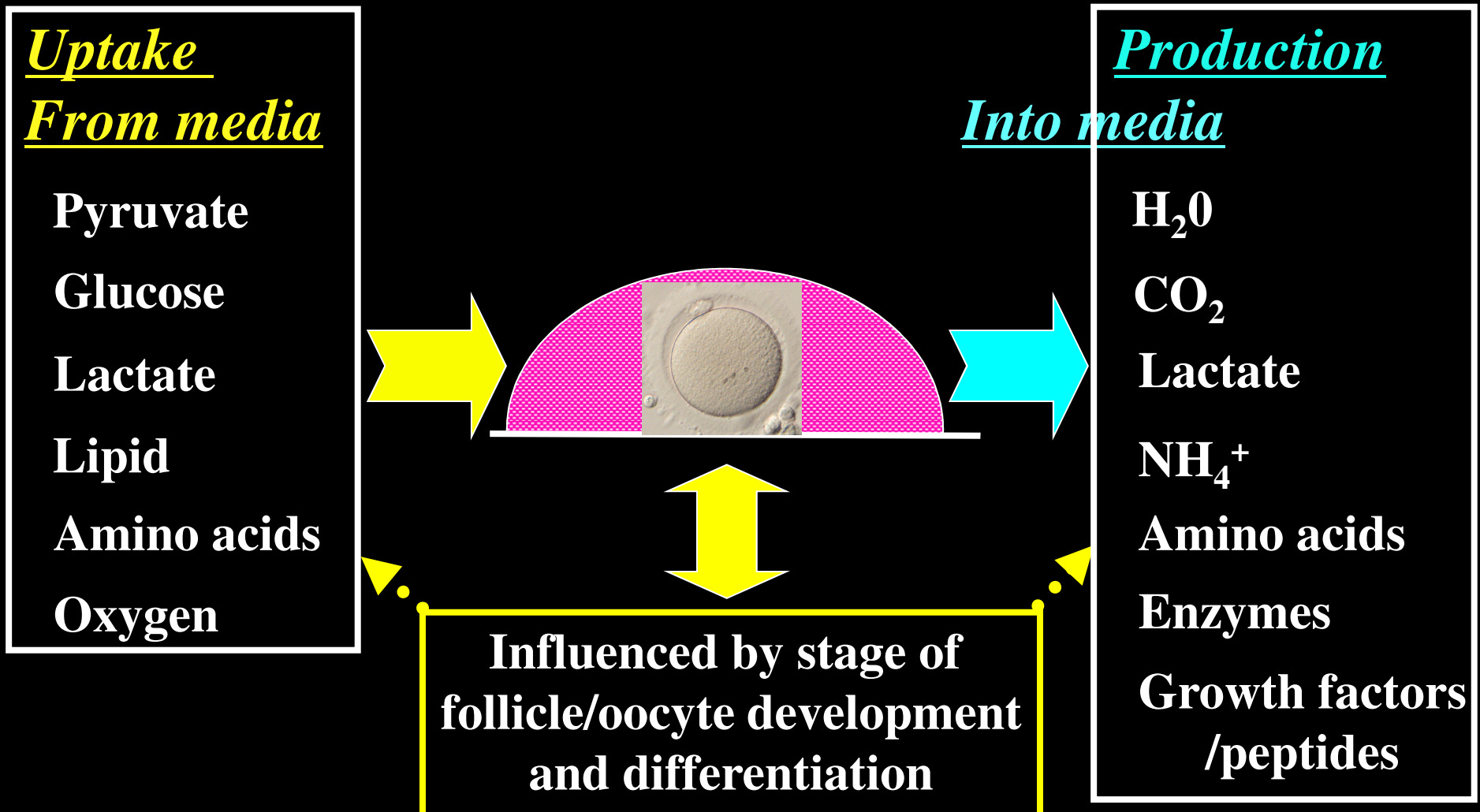
Gonadotrophin dependent

Molecular regulation: transcription factors, growth factors, peptides, steroids

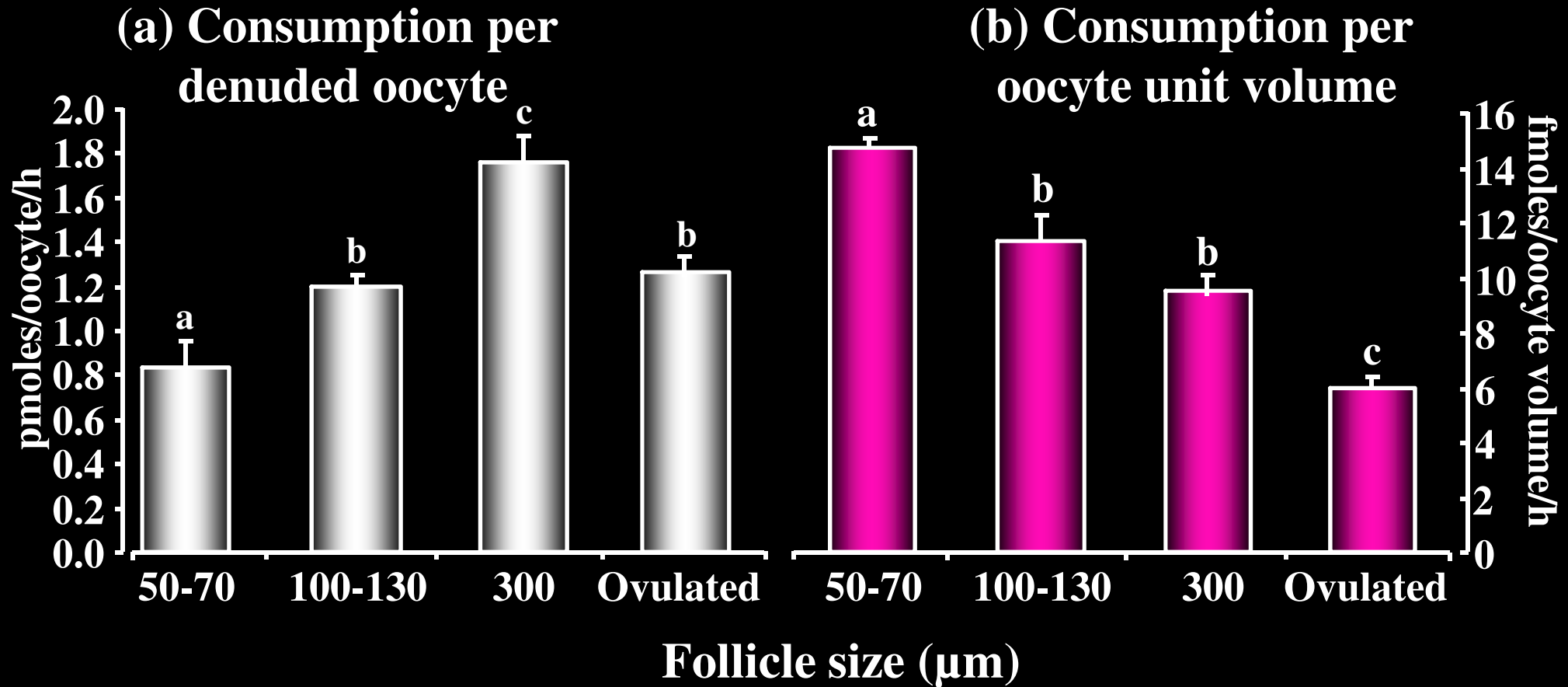
Cell-cell interactions & signalling for follicle & oocyte growth & development

Metabolic indices & energy requirements for oocyte development?

Metabolism Measurement During Follicle And Oocyte Development *In Vivo* And *In Vitro*



Pyruvate Consumption By Individual Oocytes Throughout Mouse Oocyte Development

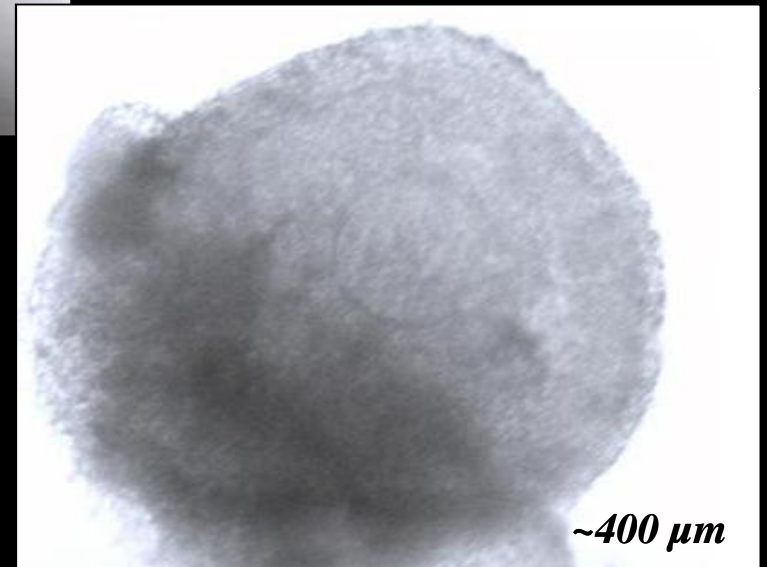
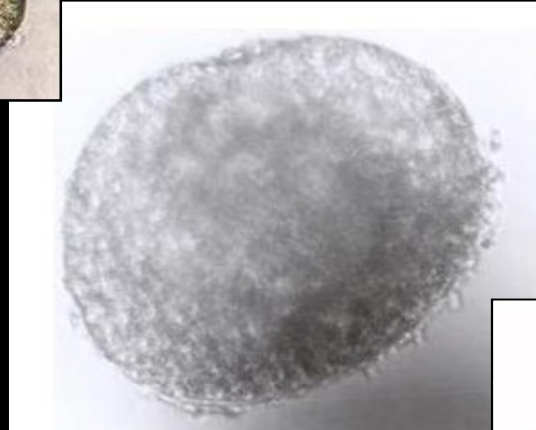


*Different letters are significantly different at $p < 0.05$
Harris et al (2009) Mol Reprod Dev. 76:231*

Mouse Follicle Metabolism *In Vivo*

Small follicles use a combination of glycolytic & aerobic metabolism of glucose

<18 μm



Diffusion of nutrients

across small distances:
primordial follicles

utilise a variety of carbohydrate energy substrates

Large follicles become almost totally reliant on glycolytic glucose consumption

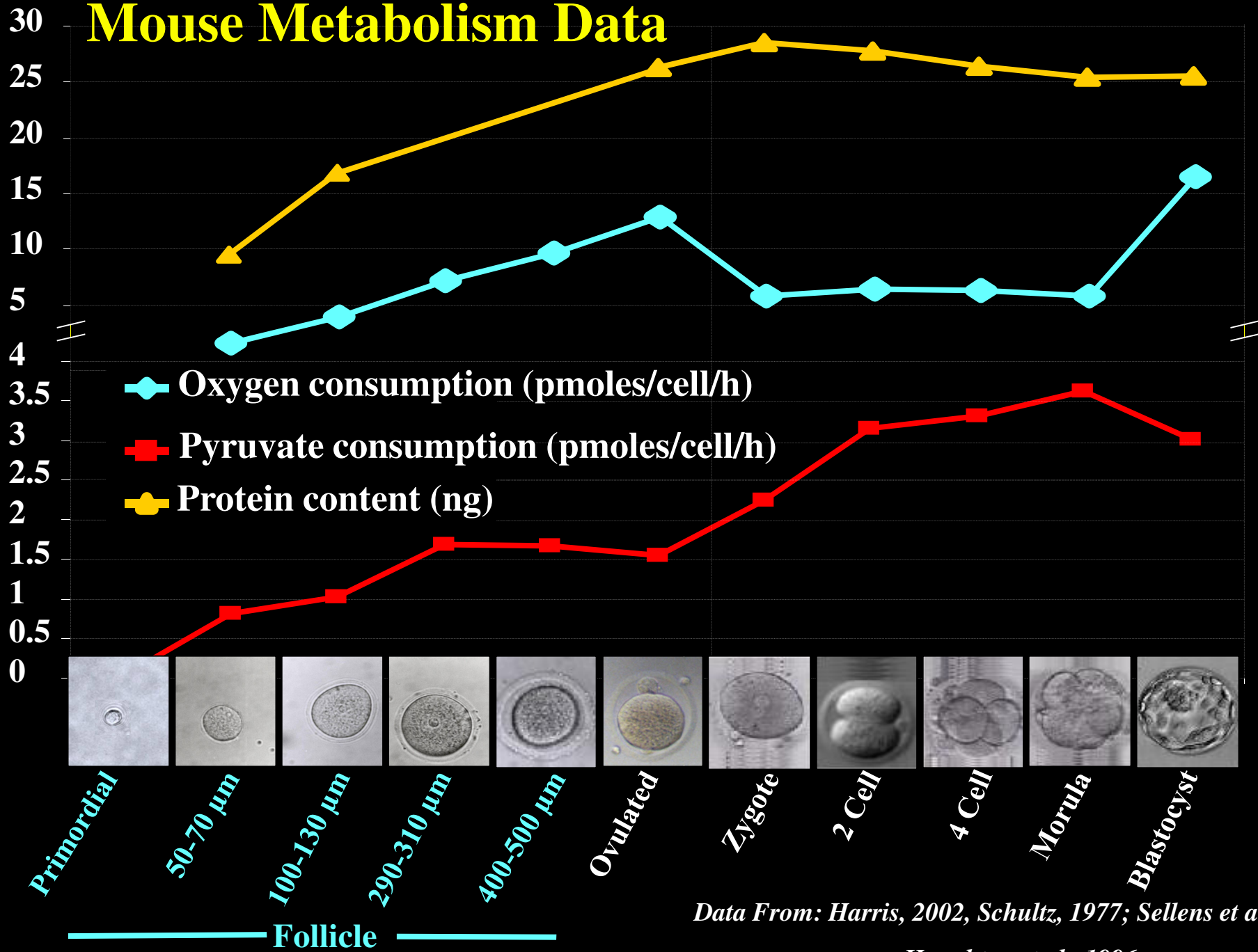
Primordial – Graafian follicle

Mouse: >30,000 –fold increase in volume

Human: >91,000,000 –fold increase in volume

(Harris 2002,
Harris et al., 2007)

Mouse Metabolism Data

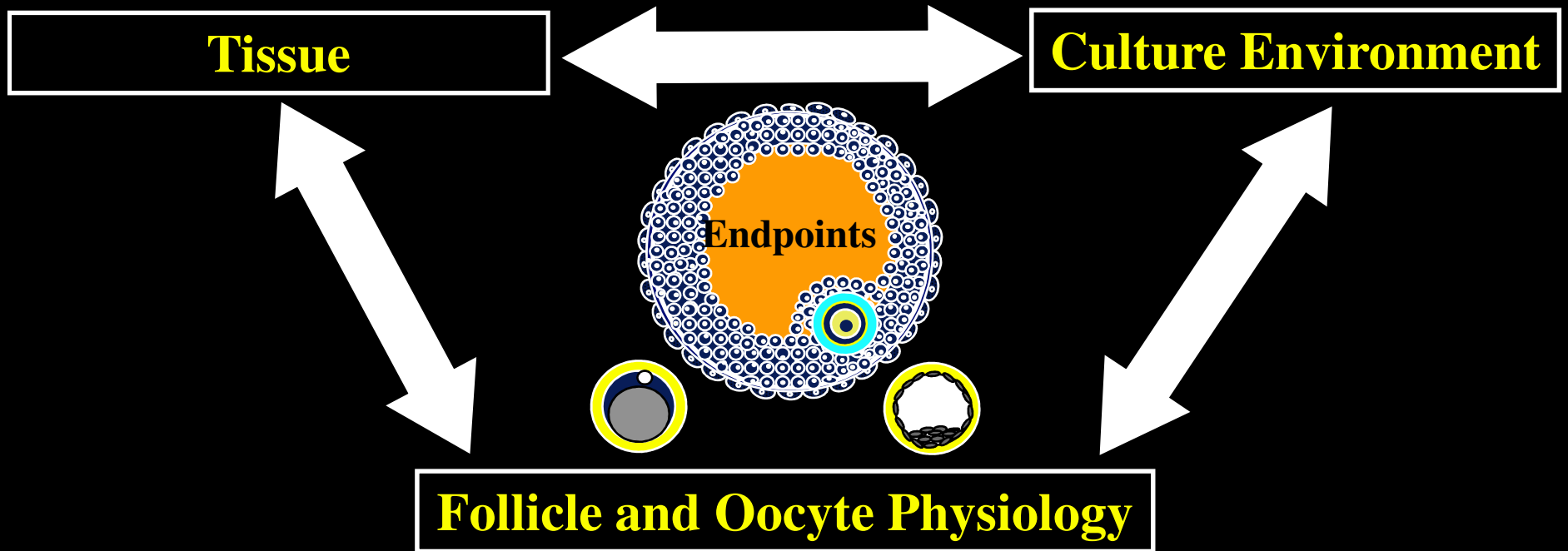


Data From: Harris, 2002, Schultz, 1977; Sellens et al., 1981;

Houghton et al., 1996

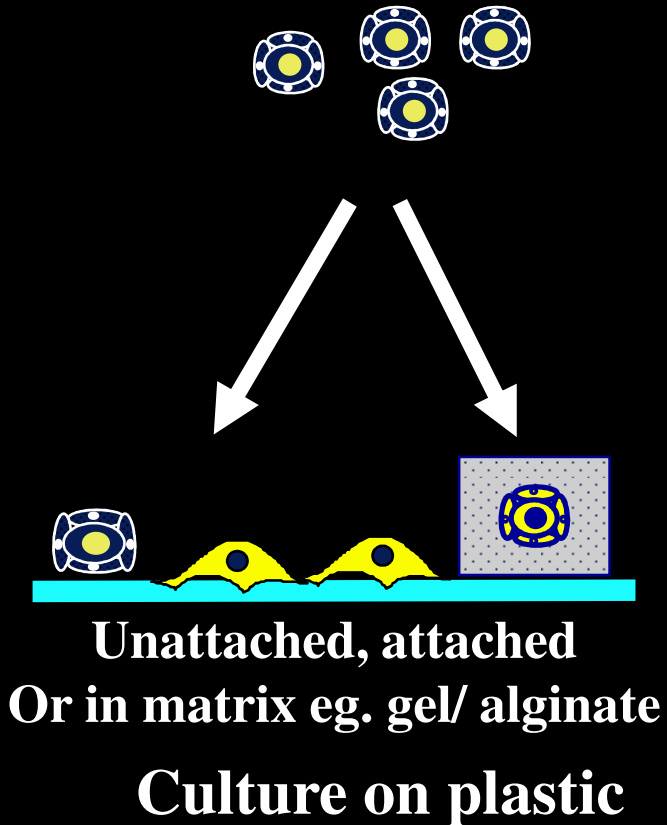
What Can Be Learned From IVM & Follicle Culture?

Rodent, Ruminant And Human

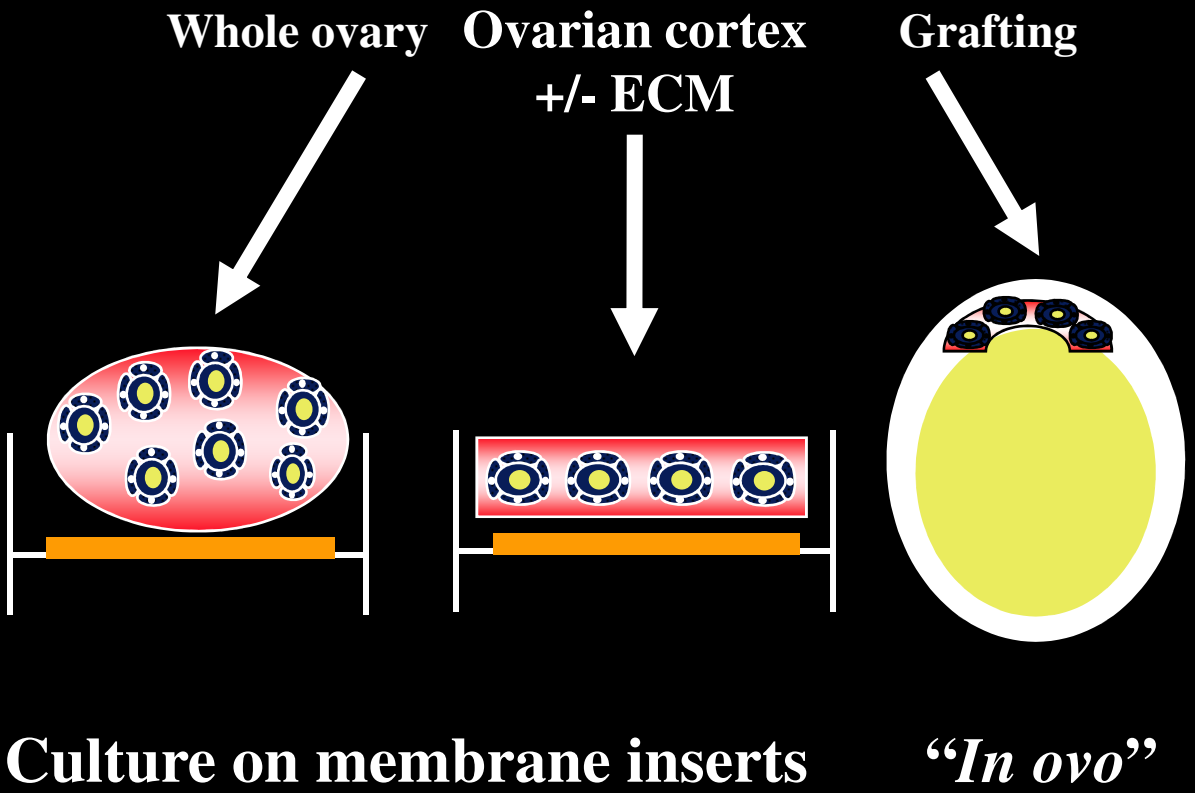


Culture Systems For Primordial And Primary Follicles

Isolated Follicle Culture



In situ Culture



Culture Systems For Preantral Follicles

**Manual Dissection
(theca enclosed)**

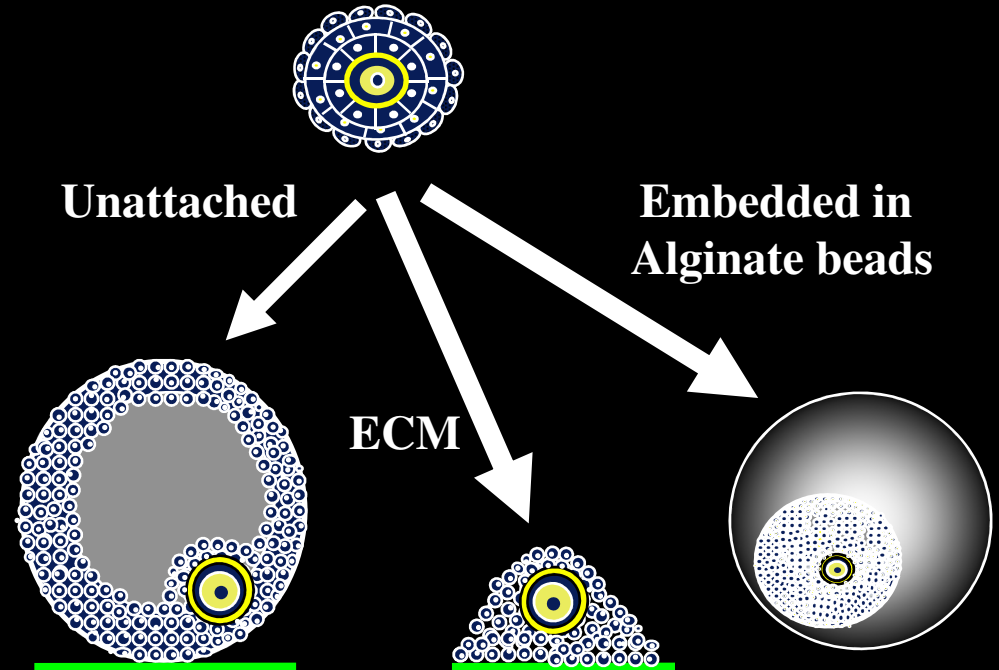
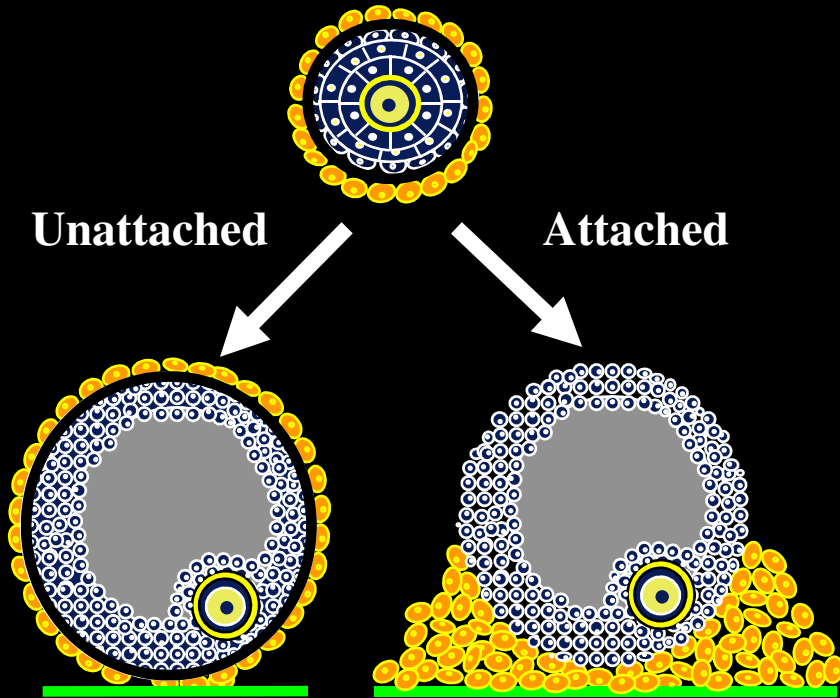
**Collagenase + Dissection
(theca free)**

Unattached

Attached

Unattached

**Embedded in
Alginate beads**



**Nayudu-Gosden-Spears
(mouse)**

**Smitz-Cortvrindt
(mouse)**

**Newton-Picton-Gosden
(sheep & human)
Telfer (bovine & human)**

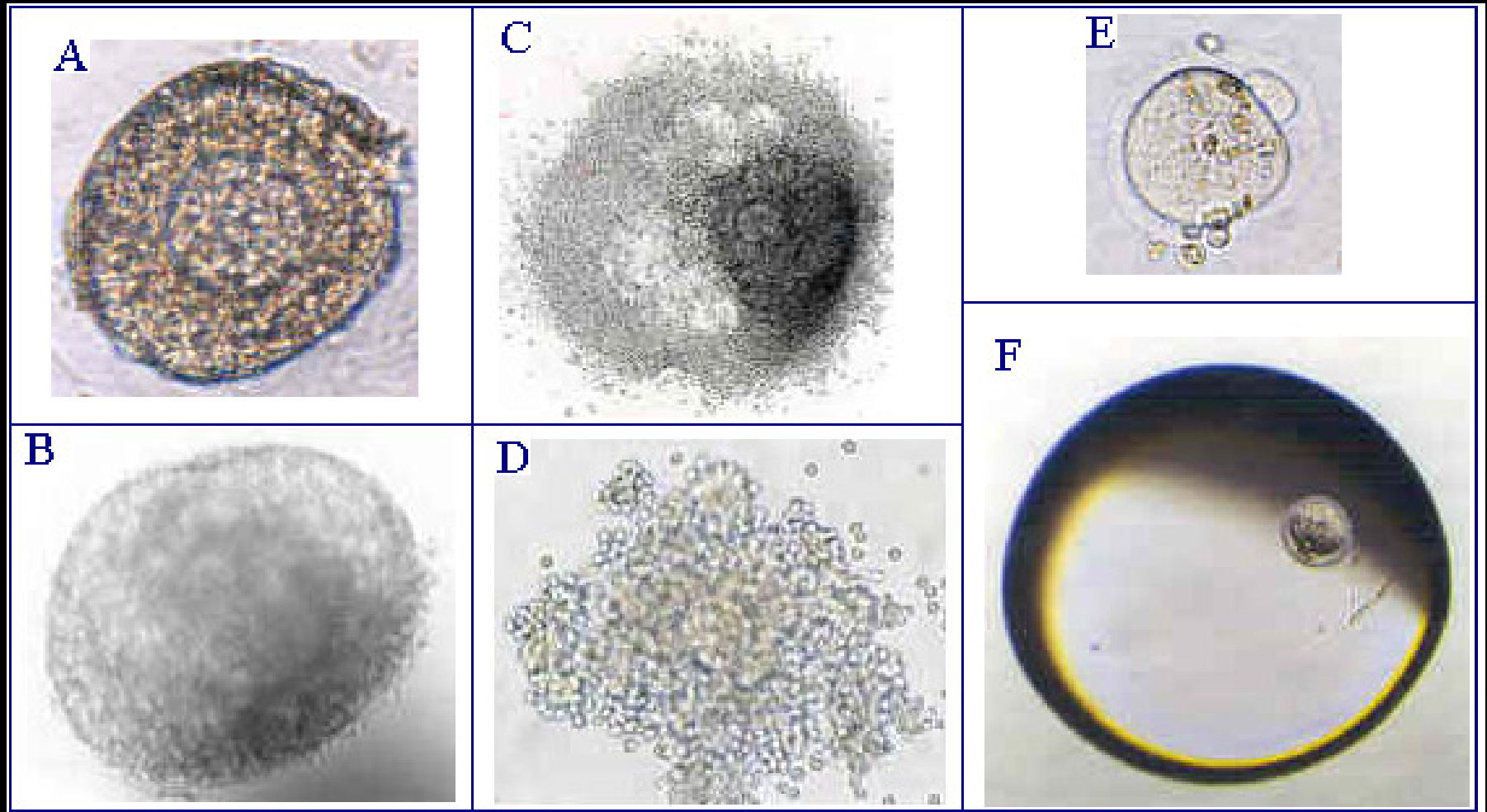
**Eppig
(mouse)**

**Kreeger et al
Heise et al
(mouse, bovine,
primate & human)**

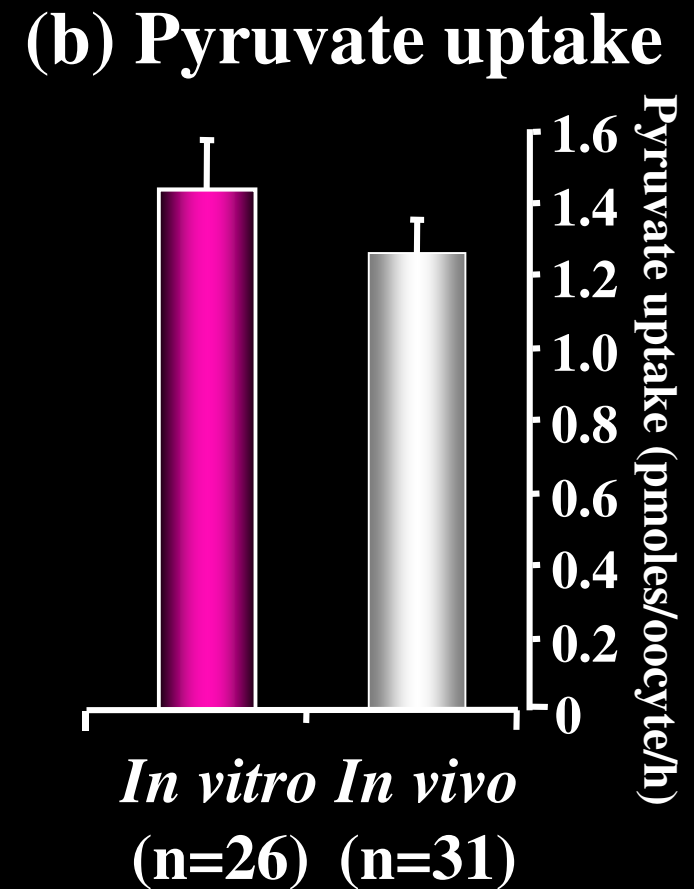
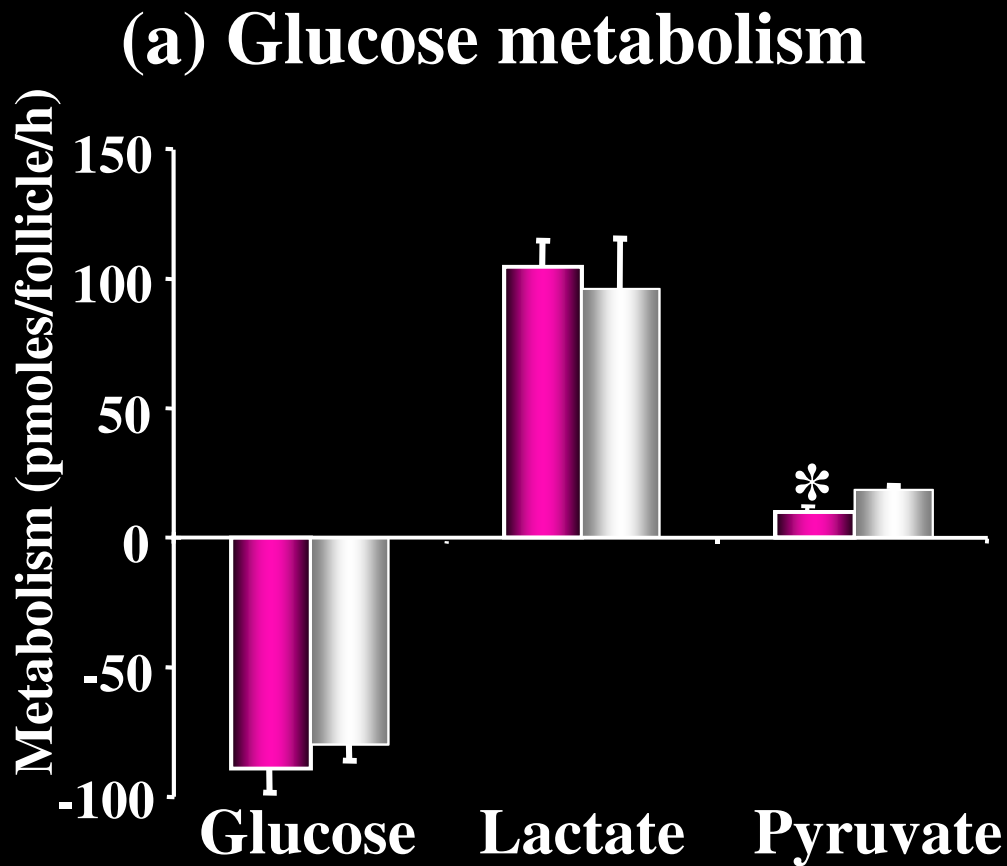
⊙ Granulosa cells ● Theca cells

In Vitro Grown Mouse Oocytes

(Follicles cultured according to the method of Cortvrintdt & Smitz, 1996)

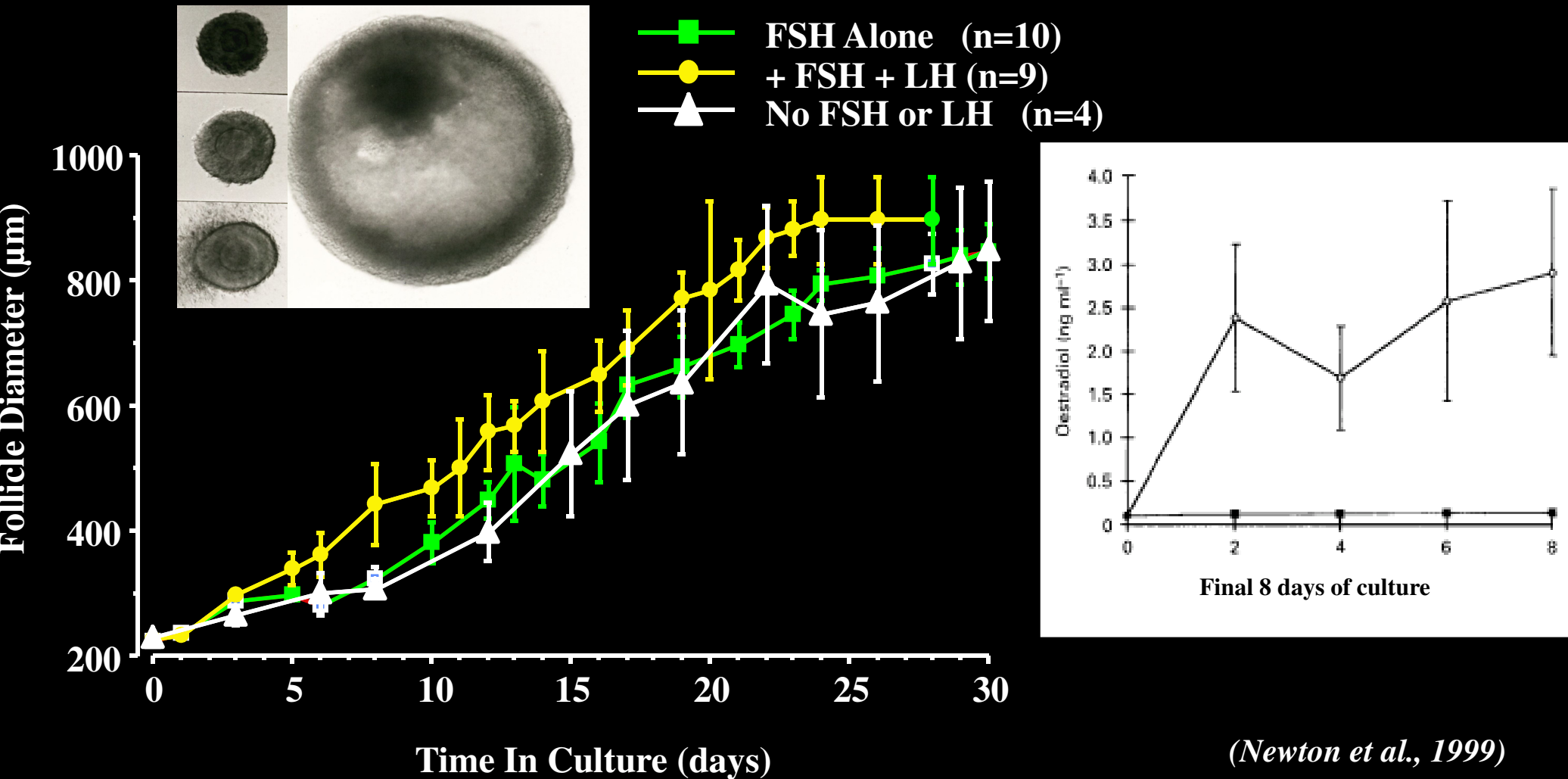


Energy Metabolism By Individual Metaphase II Mouse Oocytes Grown *In Vivo* & *In Vitro*



Gonadotrophic Regulation Of Preantral Ovine Follicle Growth *In Vitro*

(Preantral follicles: 180-200 μm diameter)

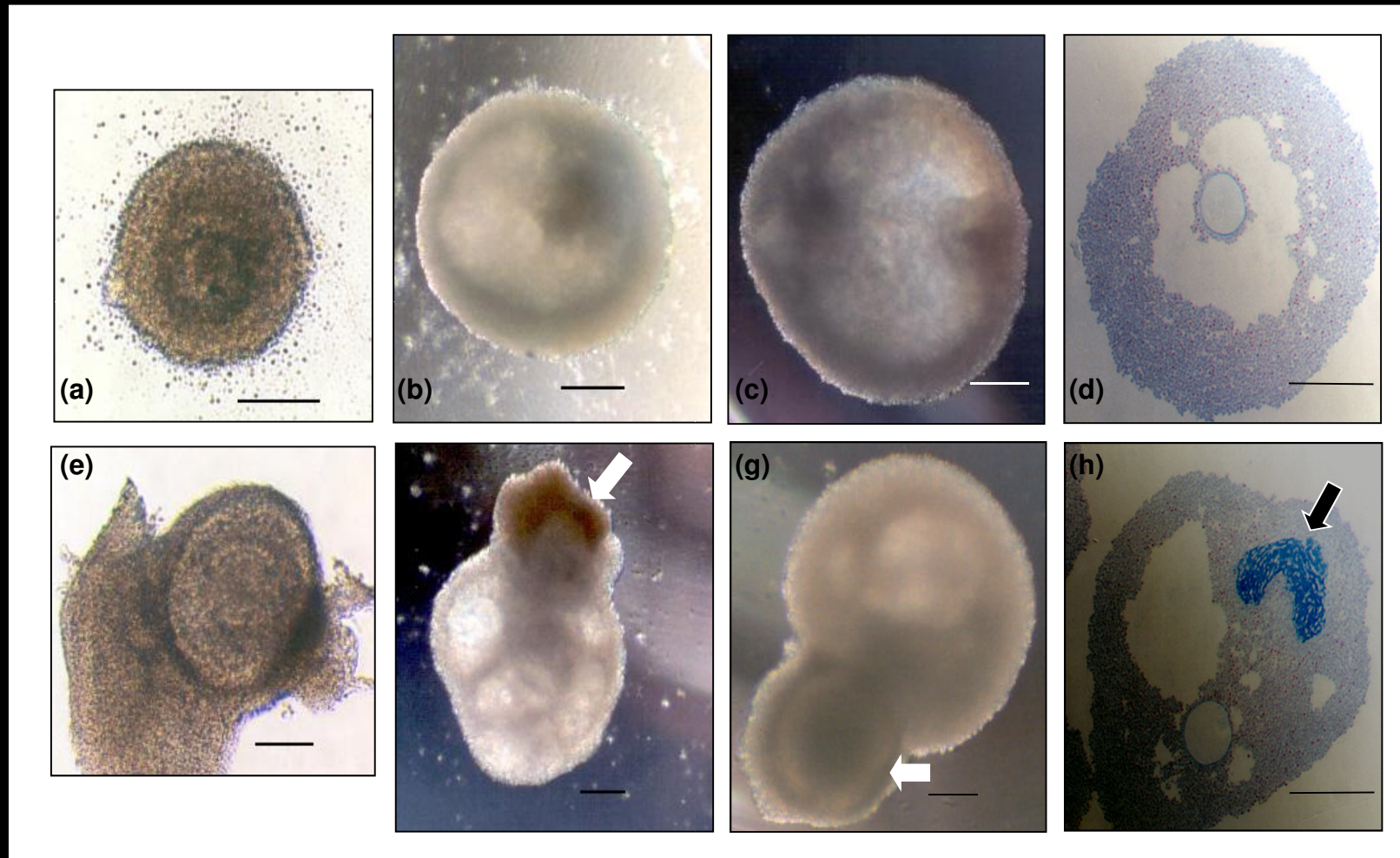


(Newton et al., 1999)

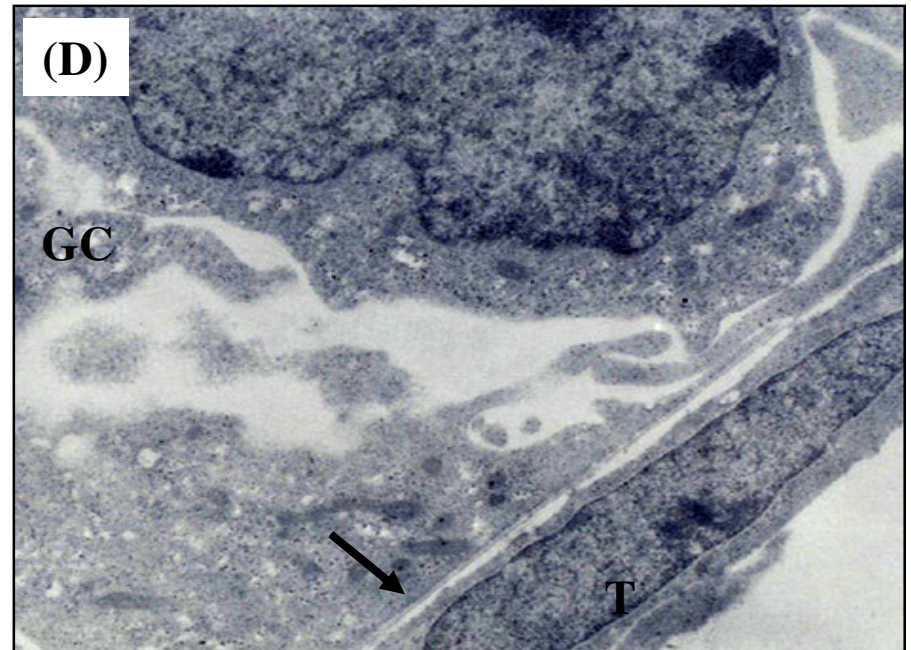
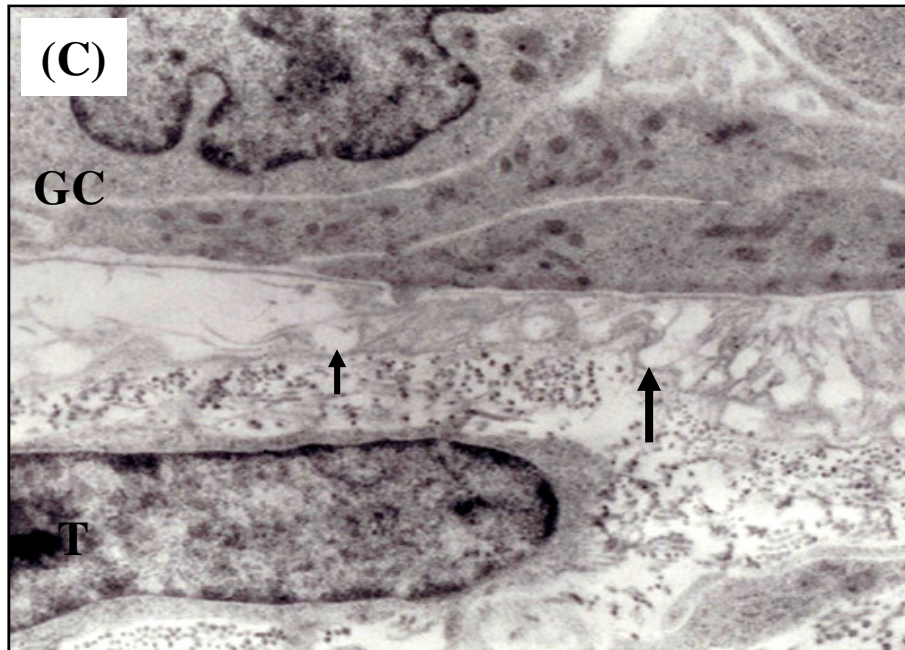
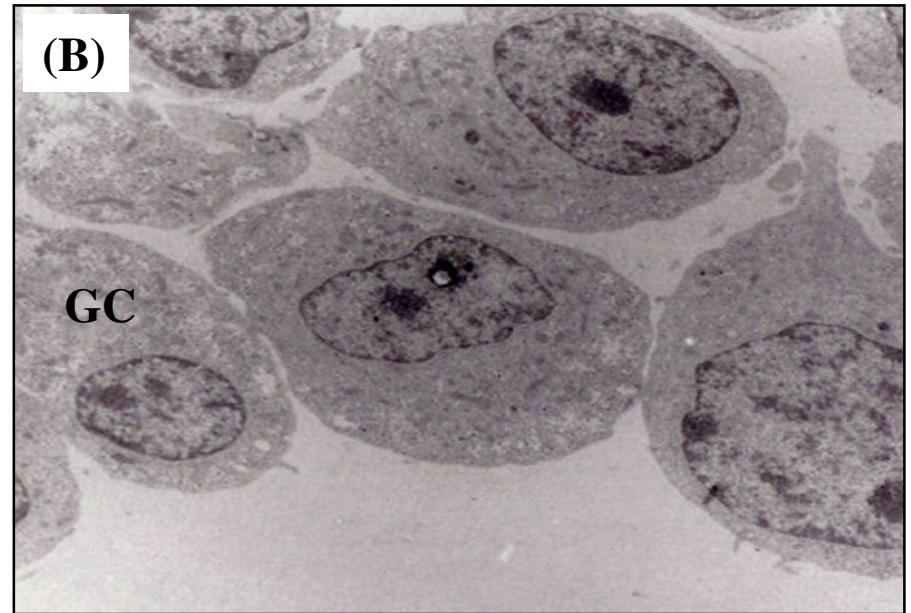
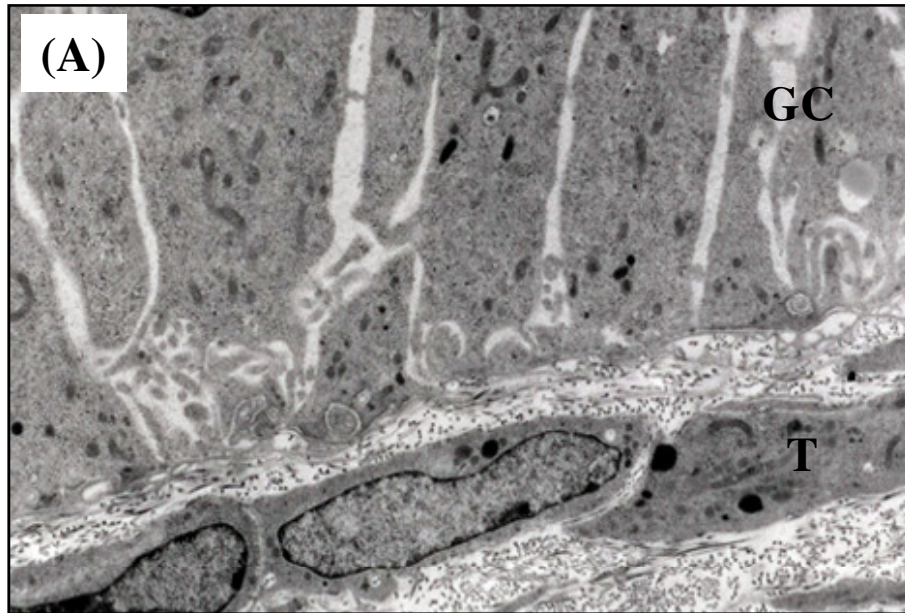
Manipulation Of *IVG* of Sheep Preantral Follicles

(a) - (d) **theca-free** follicles harvested by enzyme digestion

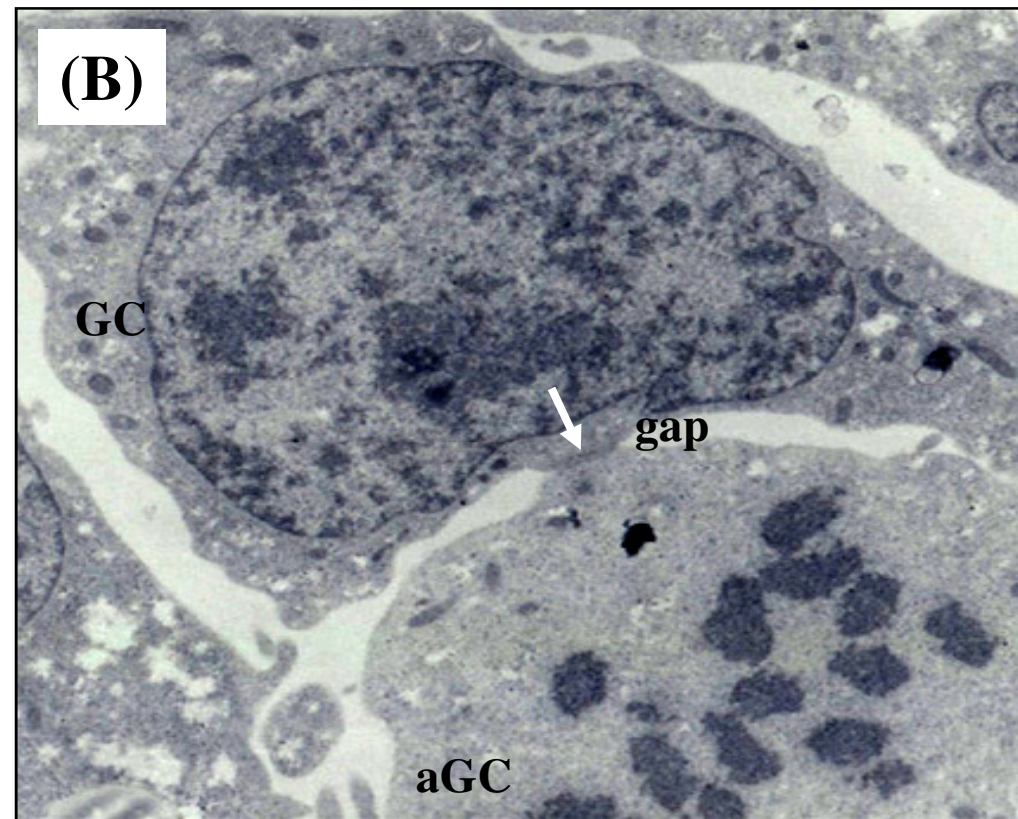
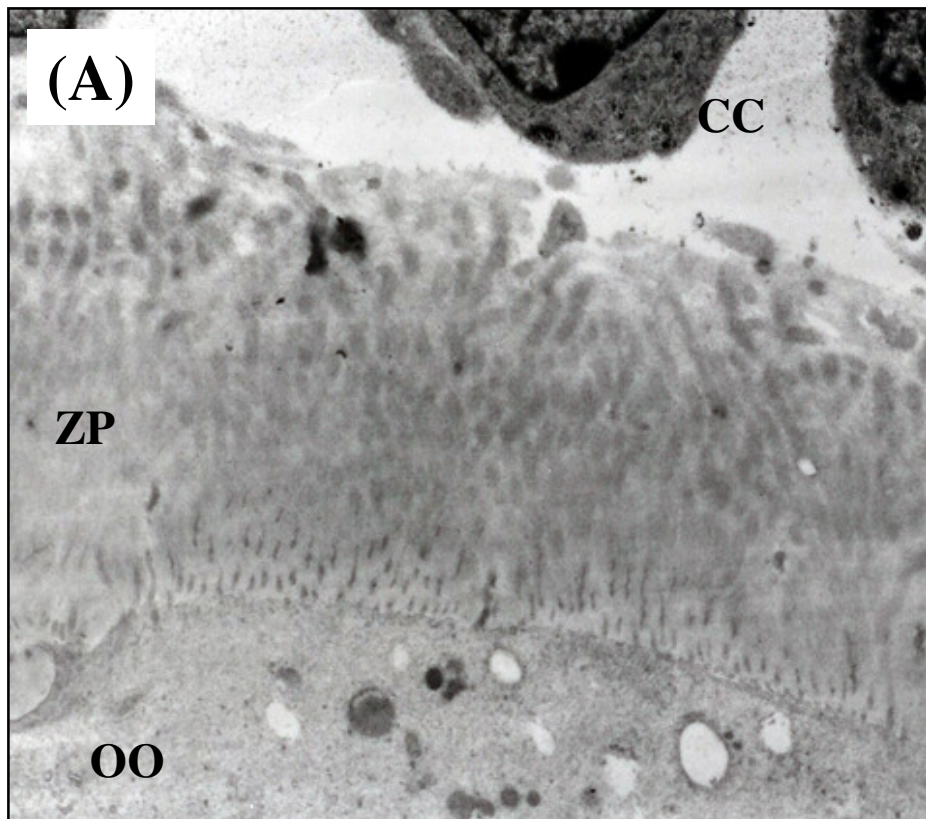
(e) - (h) **theca-enclosed** follicles harvested by mechanical isolation



TEMs Of Sheep Follicles Cultured For 30 Days



TEMs Of Cultured Sheep Follicles



Follicles cultured for 30 days in serum-free medium containing ascorbate

Picton et al 2006

Summary Of Metabolism Measurements During *In Vitro* Growth Of Follicles and Oocytes

Oocyte Physiology

Growth rates
Growth initiators & regulators
Gene transcription & translation
Epigenetic regulation
Metabolic requirements

Culture Environment

- Media composition
 - Growth factors?
 - Substrates & energy?
 - Paracrine production?
- Culture system & duration
 - sequential, perfusion, O_2

Endpoints

A central diagram showing the stages of follicle development. At the top is a large, multi-layered follicle with a yellow center, labeled 'Endpoints'. Below it are two smaller follicles, one with a grey center and one with a white center, both with yellow outer layers. A large blue arrow points from the 'Culture Environment' box to the 'Oocyte Physiology' box, and another large blue arrow points from the 'Culture Environment' box to the 'Endpoints' diagram. A large blue arrow also points from the 'Endpoints' diagram to the 'Metabolic indices & energy requirements for oocyte development in vivo' box.

Metabolic indices & energy requirements for
oocyte development *in vivo*

In Vitro Maturation Of Oocytes

The *In Vitro* Maturation Of Oocytes

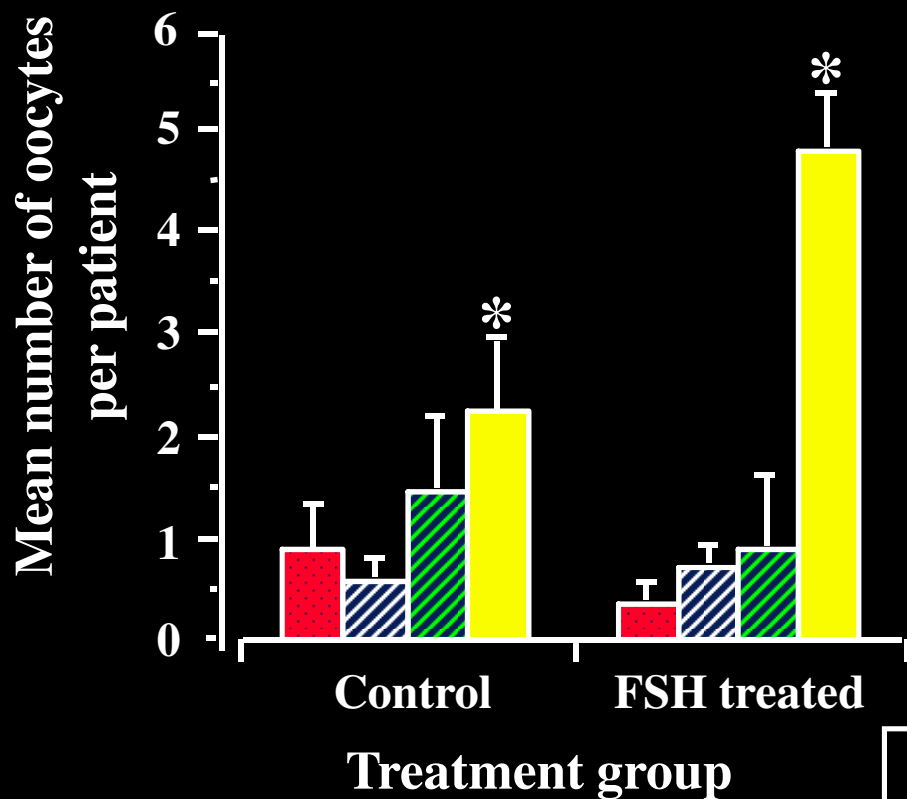


Results in all species indicate that we can:

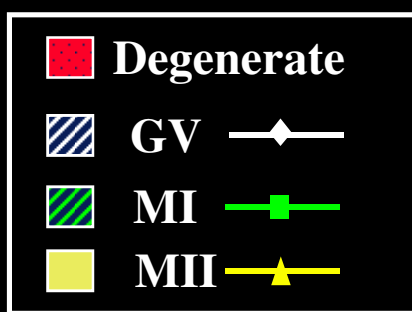
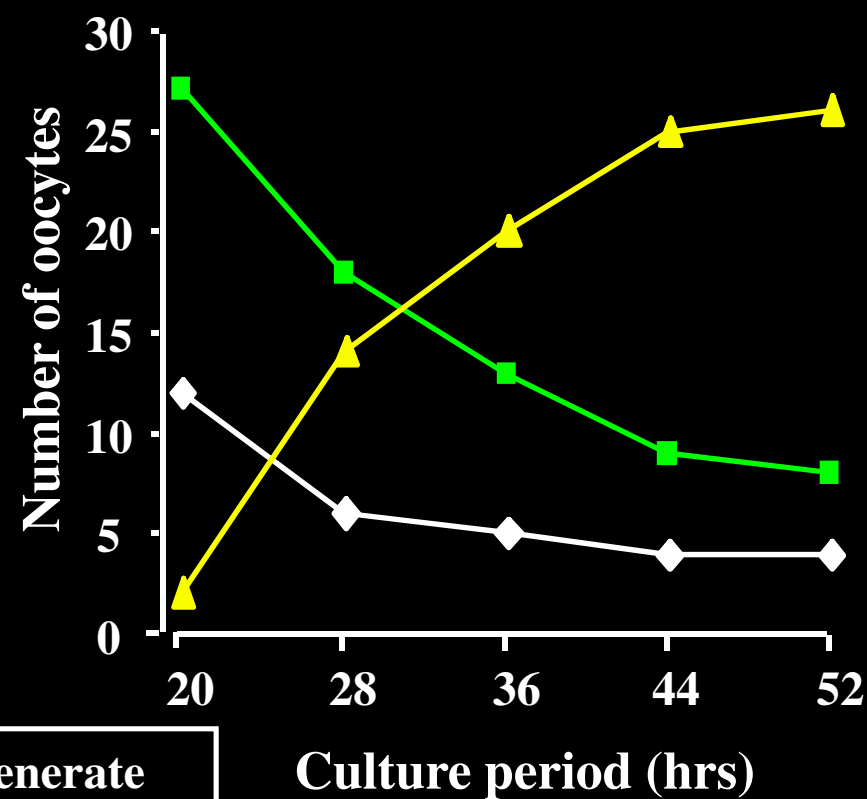
- Induce nuclear and cytoplasmic maturation in cumulus enclosed secondary oocytes to produce MII gametes with variable developmental potential
- More limited understanding of the developmental potential and epigenetic normality of oocytes derived from early stages *in vitro*.
- Clear need to improve efficiency of IVM by quantification of the biological risks and limitations of the technology

Analysis of Oocyte Maturation *In Vitro* In Humans

(a) Effect of FSH Priming on IVM



(b) Timing of IVM



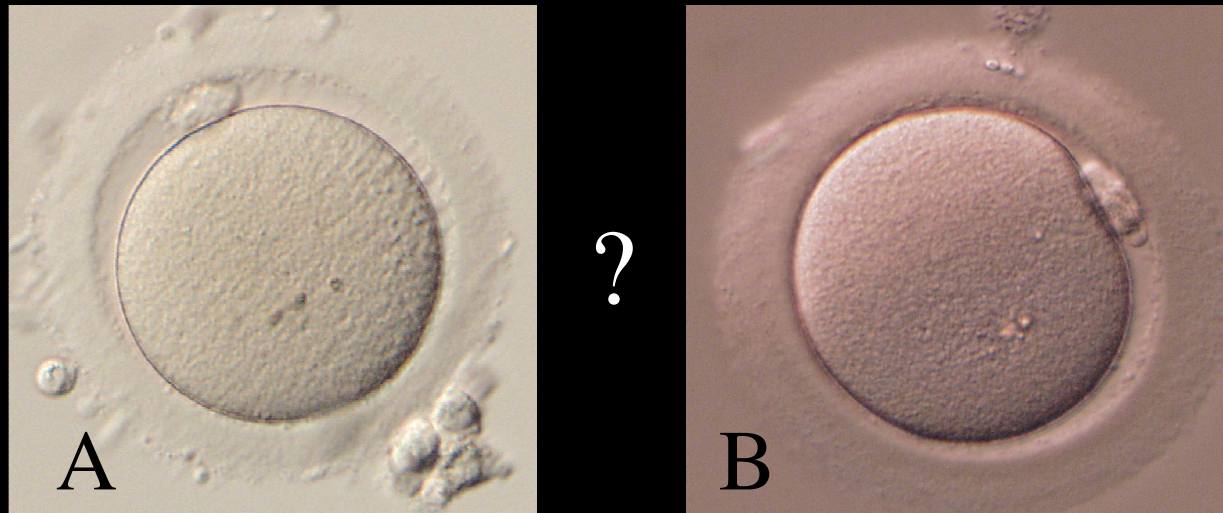
Effect Of Gonadotrophins During IVM On Ovine Embryo Development *In Vitro*

IVM Media	No oocytes inseminated	Embryo cleavage no.	Blastocyst no.
Serum-free + Gn	146	103 (70.5%)	29 (28.1%)
Serum-free - Gn	112	59 (52.6%)*	4 (6.7%)*
10% FCS + Gn	139	105 (75.7%)	24 (22.8%)

* = P < 0.05

Cotterill, Catt & Picton Unpublished

Can Amino Acid Turnover And Energy Metabolism Be Measured In Individual Oocytes ?



1. Do developmentally competent oocytes have a distinct metabolic finger print ?
2. Can the metabolic signature of an oocyte be linked to molecular &/or cytogenetic correlates of developmental competence ?

Physiological Functions Of Amino Acids

- ❖ Building blocks for protein synthesis
- ❖ Energy source
- ❖ Involved in nucleotide synthesis
- ❖ Osmolyte functions
- ❖ Antioxidant functions
- ❖ Involved in pH regulation (micro buffer function)
- ❖ Chelators- working as protection against oxidation
- ❖ Signalling molecule precursors

Amino Acid Profiling

As The Means To Select The Best Embryo

Philosophy of Approach

- **The most viable preimplantation embryos are those with the lowest level of metabolism i.e. the “*quiet embryos*”**

overall metabolism, aa turnover and glycolysis

- **Low metabolism is achieved by reducing the concentration of nutrients in culture media to the levels measured in the female reproductive tract, this encourages the embryo to use endogenous resources.**

Can this approach be applied to individual oocytes?

**Evidence of links between
oocyte metabolism & quality during IVM
in cows**

What about humans?

Summary Of Metabolic Studies Of The Later Stages Of Oocyte Development During IVM

- Amino acid consumption/production is significantly different between individual, developmentally competent bovine MII oocytes and those which fail to fertilise and/or arrest during embryo cleavage.
- Asparagine, glutamine, serine and phenylalanine turnover are potential markers of bovine oocyte developmental competence.
- Carbohydrate and amino acid metabolism by human oocytes are significantly linked to oocyte developmental competence, patient age, aetiology, gonadotrophin dose/treatment and insulin sensitising drug treatment *in vivo*.

Follicle and Oocyte Development In Health & Disease

Metabolic assays
Molecular investigations
Assays of gene function

*Research in rodents, large animals & humans,
In vitro growth and maturation of oocytes & follicles*

Acknowledgements



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Vinay Sharma

Anthony Rutherford

Adam Glaser

Tommy Tang

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