







Selection of mature oocytes with high developmental competence to obtain good embryos and/or use oocyte scores to obtain additional information on embryo quality

- 1. Oocyte with normal genetic constitution (faithful separation of chromosomes on the meiotic spindle)
- 2. Cytoplasmic mature oocyte (chromatin epigenetically remodelled during growth; full recruitment of maternal RNA and proteins, e.g. zona proteins; high number of functional mitochondria)
- 3. Oocyte from healthy follicle (efficient paracrine signalling to provide optimal conditions for oocyte maturation but also all aspects of ovulation and remodelling of extracellular matrix which may support oocyte/sperm interactions and early development and implantation)

Normal Genetic Constitution and Potential to Support Chromosome Segregation in the Preimplantation Embryo:

Aberrant spindles are a hallmark of low quality human metaphase II oocytes (e.g. from aged patients which are frequently aneuploid)







Non-invasive analysis of spindle morphology and formation

Eichenlaub-Ritter et al., RBMOnline, 2004 Jan;8(1):45-58.









- 2. Scoring by Analysis of Kinetics and Maturity at in vitro Maturation
- 1. Identify the oocytes from a cohort that have the "highest" quality and developmental potential (without chromosomal aberrations)
- 2. Identify factors (e.g. stimulation protocol, handling/culture conditions) that may contribute to obtain ,healthy' oocytes
- 3. Obtain information on whole cohort/treatment cycle to predict success and/or councel patients

















Absence of a birefringent spindle is frequently associated with disturbed spindle organization and chromosome congression failure



















Timing of progression to meiosis II is dependent on maturation conditions/ components in culture media and this may significantly affect developmental potential and quality of embryos!			
MediumA: PB start at ≈7+3hrs	Medium B: PB start at ≈7+1.5hrs		
Octax - Pol Circularly polarised light/ elec	arisation Microscopy: ronically controlled LC polarizing optics		

Dynamics (timing and progression) of anaphase I and polar body formation/ metaphase II arrest can be assessed, for instance in relation to source, handling or culture conditions of oocytes and in particular for selection of fully mature oocytes and embryos with high developmental compentence

Influence of absence/presence of a birefringent spindle on fertilization and development

	n % + spindle		%Fertilized		Embryo
			+ spindle	-spindl	le quality
Wang et al. (2001) Fertil.Steril.	533	61.4	61.8	44.2ª	
Wang et al. (2001) Hum.Reprod.	1544	82.0	69.4	62.9ª	
Rienzi et al. (2003) Hum. Reprod.	532	91.0	74.8	33.3 ^b	
Cooke et al. (2003) Hum.Reprod.	124	92.7	70.4	n.d.	
Moon et al.(2003) Hum.Reprod.	626	83.6	84.9	75.7ª	(62.9/35.9)
Cohen et al. (2004) Hum.Reprod.	770	76.0	70.6	62.2ª	
Konc et al. (2004)J.Ass.Reprod.Gene	428	74.8	73.4	n.d.	
Shen et al. (2006) RBM Online	1369	83.9	88.5	66.4 ^b	
Chamayou et al., RBM Online	967	42.9(?)	n.d.	n.d.	(43.5/48.5)
Rama Raju et al., (2007)RBM Onl.	205	88	82.5	31.1 ^a	(48.5/14.3ª)





There may be less aneuploidy in embryos with good PN-score compared to bad PN-score:

Balaban et al., 2004, RBMOnline 8, 695-700.

Gianarolli et al., 2003, Fertil. Steril. 80, 341-349.

Kahraman et al., 2002, Hum Reprod. 17,3193-3200.

















Mean retardance of light and spindle length correlate to PN-Score				
	n	Retardance (nm)	Length (µm)	
PN-Score A,B	180	1.72 ± 0.43	12.7 ± 1.8	
PN-Score C	51	$1.53\pm0.40^{\ast}$	12.5 ± 1.6	
PN-Score D	324	$1.52 \pm 0.44 ^{**}$	12.6 ± 1.7	
PN-Score E and Abnormals	121	1.39 ± 0.46^{stst}	11.7 ± 1.7**	
Significantly different to score A,B; * p < 0.05; **p < 0.001.				























Magnitude of retardance and length of the human oocyte spindle may be used to identify non-invasively oocytes from a cohort forming embryos with good PNscore, which may be chromosomally normal and possess high developmental potential (e.g. high number and activity of mitochondria)

Mean magnitude of retardance and length of the human oocyte spindle may be used to identify indivisual patients/cycles with higher or lower chance to conceive



Quantitative Imaging of the Zona Pellucida of Human Oocytes (Pelletier et al. 2004, Fertil. Steril):

3 Layers; retardance initially increases at fertilization; zona then becomes thinner during early development

Quantitative Imaging of the ZP prior to fertilzation by ICSI (Shen et al., 2005, Human Reproduction, 20:1596-1606)

•Analysis of Mean Thickness of the Layers of the Zona Pellucida

•Analysis of Mean Retardance of Layers of the Zona Pellucida

•Comparison of Data between Oocytes used in Transfer in Conception Cycles (CC) versus Non-Conception Cycles (NCC)







No significant difference between conception cycle (CC) and non-conception cycle (NCC) in: & Patient age & Maternal smoking & Number of attempts & Peak Oestradiol levels & Number of follicles & Number of follicles & Number of oocytes & Fertilization rate & Oocytes with birefringent spindle & Number of embryos transferred

Thickness (pm)	2.81:46.60' 11.25:1.441'	2.15±0.41 9.36±1.74	Mean retardance of zona inner layer about 30% higher in CC compared to NCC!









Study by Munne et al. 2007: Embryo grading is not very predictive for presence of chromosomal aberrations

Chromosomal aberrations in good grade (1) embryos: 30%

Chromosomal aberrations in low grade (4) embryos: 44%

Qualitative and quantitative analysis of spindle and zona retardance has additional predictive value to assess chromosomal aberrations of the oocyte and may help to identify those ,best' oocytes with presumably full nuclear and cytoplasmic maturity related to good developmental potential leading to implantation, conception cycles and birth of a healthy child. Delayed progression into anaphase I, polar body formation and establishment of a metaphase II spindle can be an indicator of a mutation or adverse exposure during oocyte maturation!



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MTG: Octax Polarisation Microsc.