

Assessment of Carbohydrate and Amino Acid Metabolism in Oocytes/Embryos

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Metabolism/Nutrition Profiling

- Ovarian follicles
- Oocytes (immature and mature)
- Embryos

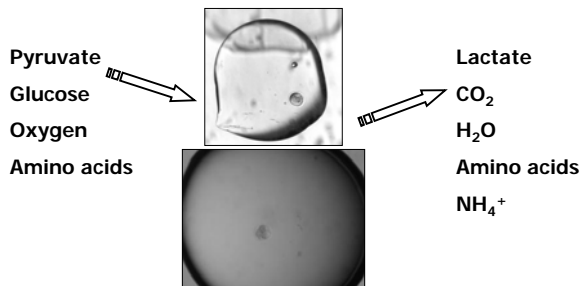
Metabolic Bio-markers of Quality

- Carbohydrates, amino acids, oxygen.
- More complete understanding of metabolism profiles of follicles, oocytes and embryos in healthy and disease states, *in vitro* and *in vivo*.
- Non-invasive metabolic bio-markers are potentially useful to assist selection of embryos produced by ART.
- Metabolism profiling: identification of potential bio-markers.
 - Follicle: *in vitro* growth
 - Oocyte: *in vitro* maturation
 - Embryo: *in vitro* culture

Metabolic Bio-markers of quality

- Several studies.
- Useful research tool.
- Associations of metabolic and biochemical markers with:
 - Meiotic maturation
 - Fertilisation potential
 - Embryo development: slow, arrested
 - Live birth rate
 - Chromosome abnormalities

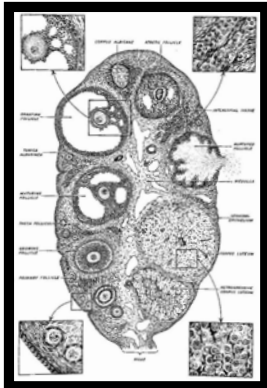
Non-invasive metabolism analysis



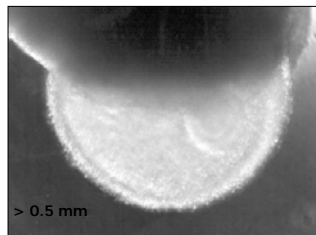
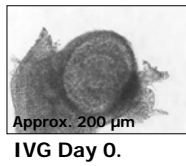
- Prospect of long-term follicle culture for production of oocytes:
 - Cancer patients – unsuitable to re-graft potentially malignant tissue
 - Source of oocytes for egg donation therapy
 - Source of oocytes for SCNT, stem cell production
- Metabolism:
 - Basic biology – involvement in selection processes?
 - Optimisation of culture protocols
 - Non-invasive bio-markers of quality?

Ovarian follicle development

- Complete follicle growth requires several months (approx. 4-5 weeks: mouse; approx. 6 months: human).
- Changing nutritional and metabolic needs of growing follicle and oocyte.
- Follicle metabolism is influenced by signals from the oocyte.
- Oocyte maturation is regulated by cumulus metabolism.



Long-term follicle culture



Possible to culture follicles for 30 or more days

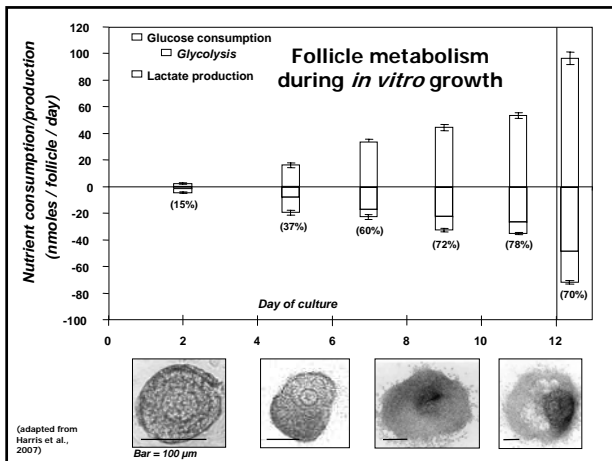
Follicle metabolism profiles change with development

Diffusion of nutrients across small distances: primordial follicles utilise a variety of carbohydrate energy substrates (Harris, 2002).

Large follicles become almost totally reliant on glycolytic glucose consumption (Boland et al., 1994; Harris et al., 2007).

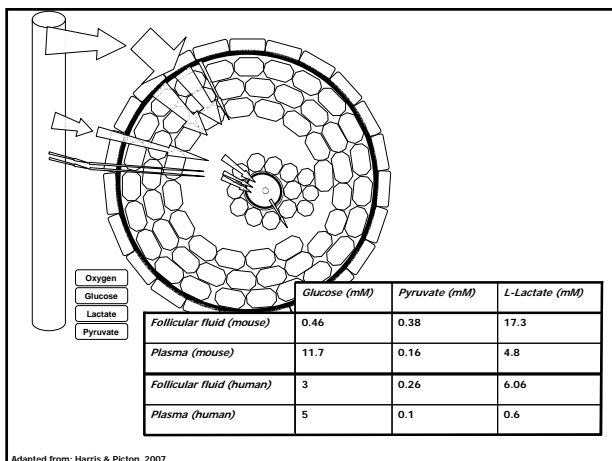
Primordial – Graafian follicle
 Mouse: >30,000 –fold increase in volume.
 Human: >91,000,000 –fold increase in volume.

Note: The image shows a series of follicles increasing in size from left to right. A scale bar indicates <math>< 18 \mu\text{m}</math> for the smallest follicles and 'Approx. 400 μm' for the largest follicle.



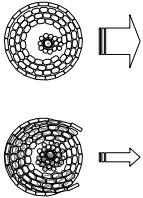
What pushes the metabolic switch?

- Small follicles use a combination of different pathways to metabolise glucose (glycolysis and aerobic metabolism). Sufficient oxygen reaches the oocyte.
- As follicles get bigger, diffusion of nutrients and oxygen to the centre becomes limited (Gosden & Byatt-Smith, 1986).
- To compensate, the follicle develops an antrum, increasing the effective surface area for diffusion over the follicle surface.
- The oocyte secretes factors (including BMP15 and FGFs) which promote follicular glycolysis (Suglura et al., 2005).
- Oxygen is able to diffuse to the follicle core.



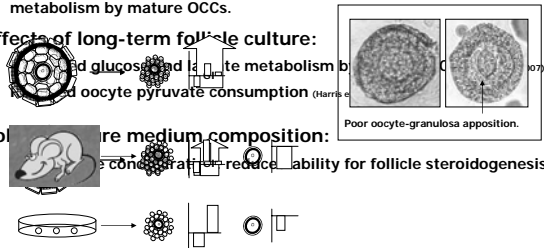
- ***In vitro* growth of follicles: useful time period over which to assess developmental characteristics:**

- Follicle lactate production during growth *in vitro* is higher by follicles not destined to ovulate (Harris et al., 2007).



Culture environment influences follicle, oocyte and cumulus metabolism

- **Quality of follicles cultured long-term influences metabolism of the mature mouse oocyte-cumulus complex** (Harris et al., 2007).
 - Reduced follicle theca coverage: increased glycolytic index in mature OCCs (cell stress).
 - Reduced follicle oocyte-granulosa apposition: reduced lactate metabolism by mature OCCs.
- **Effects of long-term follicle culture:**
 - Increased glucose and lactate metabolism by mouse OCCs (Harris et al., 2007).
 - Reduced oocyte pyruvate consumption (Harris et al., 2007).
- **Follicle culture medium composition:**
 - Low glucose concentration reduces ability for follicle steroidogenesis (Boland et al., 1994).

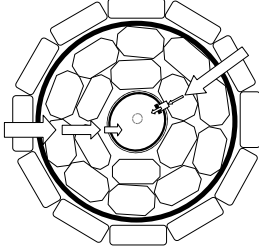


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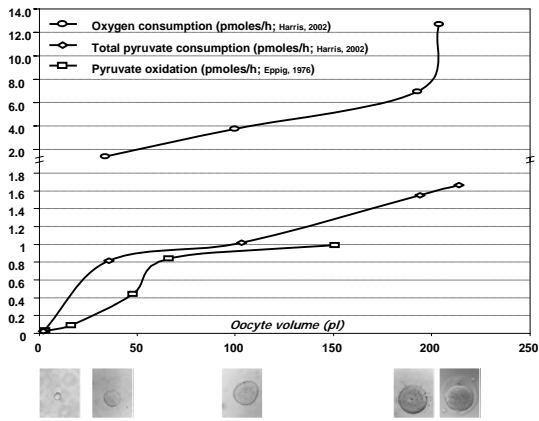
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Oocyte Metabolism and Nutrition

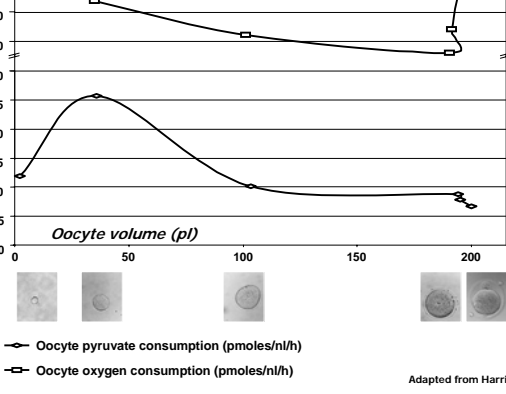
- During oogenesis, the oocyte obtains metabolites by direct uptake of metabolites from interstitial fluid/follicular fluid.
- Metabolites and molecules can also pass directly to the oocyte via gap junctions with granulosa/cumulus cells.

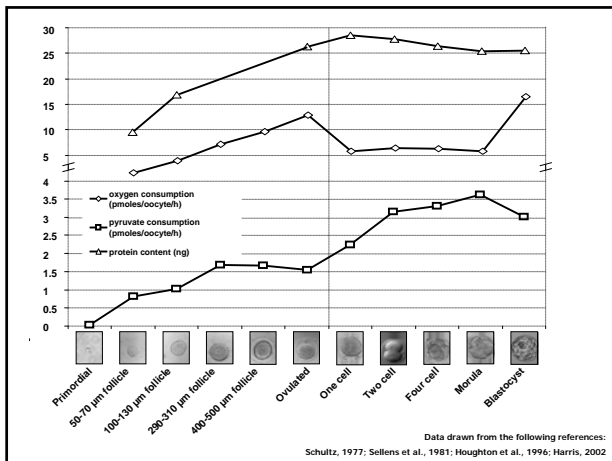


Nutrient consumption by growing mouse oocytes



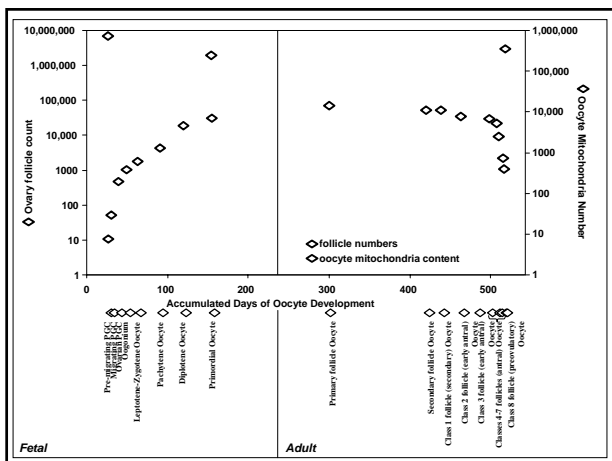
Nutrient consumption by growing oocytes, standardised for cytoplasm volume





Mitochondria: role in oocyte quality

- Oocyte and preimplantation embryo metabolism: oxidative.
- Requirement for adequate cohort of good quality mitochondria.
- Bottleneck:
 - Mitochondria Restriction: expansion of a very small cohort of mitochondria.
 - mtDNA Segregation: expansion of a subset of segregated mitochondria.

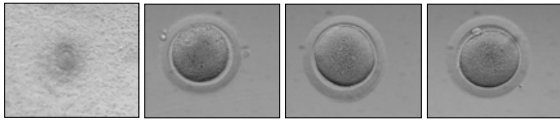


Mitochondria quality

- **Accumulation of mtDNA mutations** (Hsieh et al., 2002).
- **Mitochondria activity** (Thouas et al., 2004).
- **Spindle assembly** (Wilding et al., 2003; Eichenlaub-Ritter et al., 2003) – **aneuploidy risk**.
- **Mitochondria morphology** (Heng-Kien et al., 2005).
- **MtDNA gene expression** (Hsieh et al., 2004).
- **Mitochondria density** (May-Panloup et al., 2005; Santos et al., 2005).
- **Repeated ovarian stimulation associated with mitochondria mutations in mouse oocytes** (Chao et al., 2005).
 - **Ageing** (Wilding et al., 2001).
 - **Ovarian failure** (May-Panloup et al., 2005).
 - **Diabetes, insulin resistance** (Sheratt et al., 1999; Kyu-Lee et al., 2005; Maassen et al., 2005).
 - **Cryopreservation** (Jones et al., 2004).
- **Identification of markers of mitochondria quality would be beneficial!**

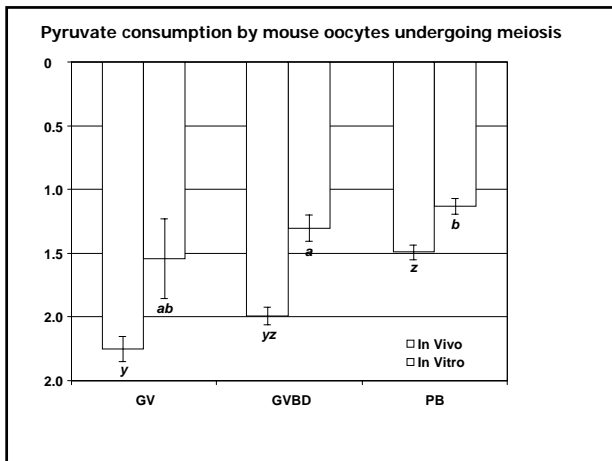
- **Metabolic profiling: identification of possible bio-markers of quality:**
 - **Follicle: *in vitro* growth**
 - **Oocyte: *in vitro* maturation**
 - **Embryo: *in vitro* culture**

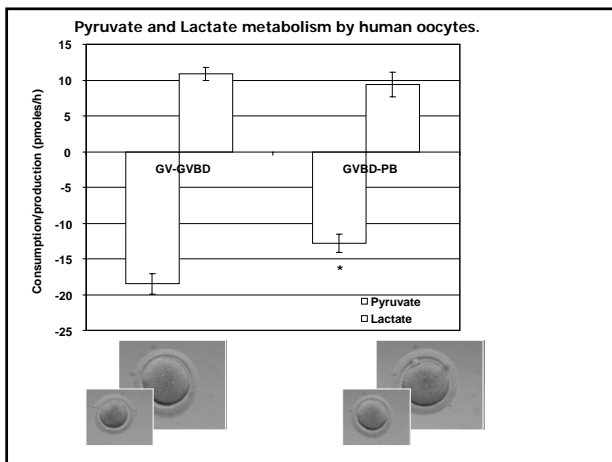
Oocyte meiotic maturation



Metabolism during meiotic maturation

- **FSH stimulates PI3-K pathway. Translocation of GLUT-4** (Roberts et al., 2004).
- **Ovulation induction stimulates glucose metabolism by the follicle** (Botland et al., 1994; Harris et al., 2007) **and cumulus cells** (Downs & Utsch, 1999).
- **More glucose diverted to the pentose phosphate pathway** (Downs & Utsch, 1999).
- **Generation of purine precursors.**
- **Glucose hyaluronic acid synthesis – cumulus expansion** (Sutton-McDowell et al., 2004).
- **Mouse oocyte pyruvate consumption rate changes during meiotic maturation** (Downs et al., 2002).

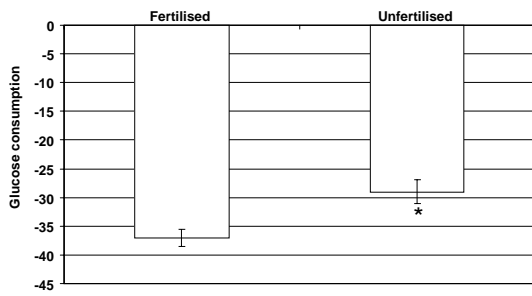




Metabolic markers during IVM

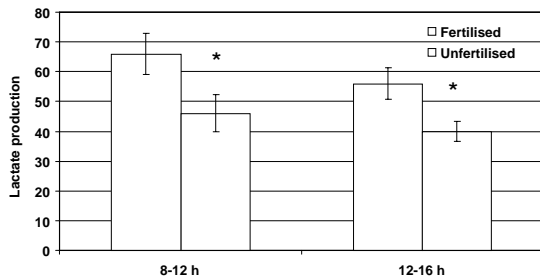
- IVM – time window for metabolism analysis.
- Human oocytes competent to mature to MII have greater glutamine consumption (Vyjayanthi et al., 2007).
- Human oocytes with intermediate pyruvate consumption more likely to implant (Turner et al., 1994).
- Higher rates of mouse OCC glucose and lactate metabolism during maturation associated with oocyte fertilisation (Preis et al., 2005).

Glucose consumption by mouse OCCs during hours 8-12 of 16 h IVM



Preis et al., 2005

Lactate production by mouse OCCs during hours 8-12 and 12-16 of 16 h IVM

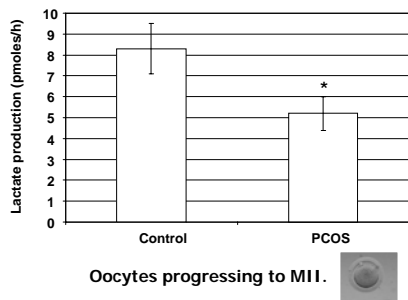


Preis et al., 2005

Metabolic disturbances during IVM

- **Reduction in mouse cumulus PPP glucose metabolism in diabetes – reduced oocyte maturation rate** (Colton et al., 2003) •
- **Altered oocyte carbohydrate metabolism in polycystic ovary syndrome (PCOS)** (Harris et al., in prep.) •

Lactate production by control and PCOS oocytes undergoing PB formation.



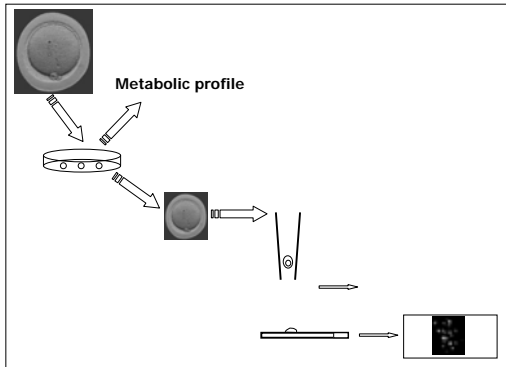
Harris et al., in prep.

- Altered metabolism observed in oocytes from patients with different aetiologies (e.g. PCOS).
- Focus on certain patient groups?
- Is metabolism disturbed in other cases?
- Chromosome abnormality?

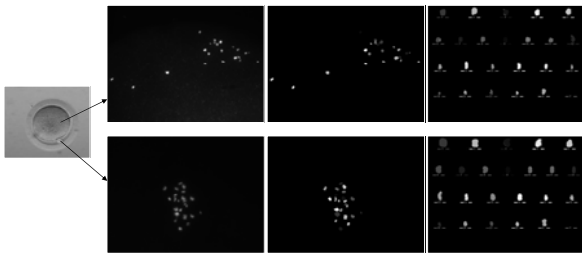
Oocyte chromosome abnormalities

- High proportion of mature human oocytes harbour chromosome abnormalities.
- Arise from errors in meiosis I:
 - Nondisjunction
 - Premature Separation of Sister Chromatids (PSSC).
- Aneuploidy in embryo.
- Chromosome segregation and polar body formation are energy-dependent.
- Age-related increase in chromosome abnormalities.
- Associated with age-related decrease in mitochondria quality? (Schon et al., 2000).
- Non-invasive markers to predict chromosome abnormality?

Investigation of oocyte chromosome abnormalities



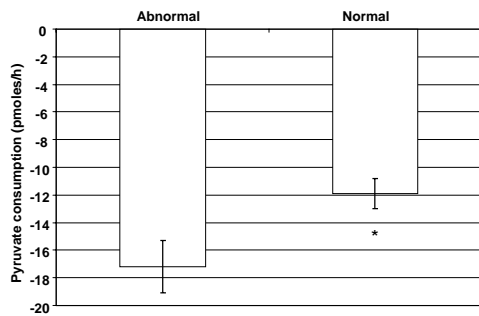
Oocyte karyotype assessed by WCP-mFISH



Chromosome abnormalities



Pyruvate consumption by oocyte with abnormal and normal karyotypes



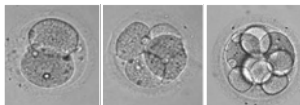
Markers?

- FF oxygen content (Van Blerkom et al., 1998).
- Follicular vascularity (Van Blerkom et al., 1998).
- Embryo chromosome abnormalities.

Metabolic profiling: identification of possible bio-markers of quality:

- Follicle: *in vitro* growth
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Embryo culture



Culture environment influences oocyte, cumulus and embryo metabolism and developmental capacity

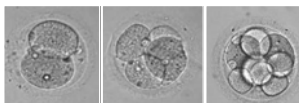
- Embryo culture medium nutrient composition affects:
 - Embryo metabolism: cell stress responses are alleviated by using more physiological culture medium (Gardner and Leese, 1990; Gardner and Sakkas, 1993).
 - Blastocyst formation (Rose-Hellekant et al., 1998).
 - Cell number (Van Soom et al., 1996).
 - Implantation rate (Barak et al., 1998).

Metabolic markers of embryo development

- Pyruvate metabolism associated with:
 - Human embryo implantation rate (Conaghan et al., 1993).
 - Human embryo development, polyspermy, parthenogenesis (Hardy et al., 1989).
- Mouse blastocyst glycolytic profile and glucose consumption associated with pregnancy outcome (Lane & Gardner, 1996).
- Mitochondria activity correlated with glutamine turnover (Harris et al., in prep.).
- Too much variation to be used reliably.

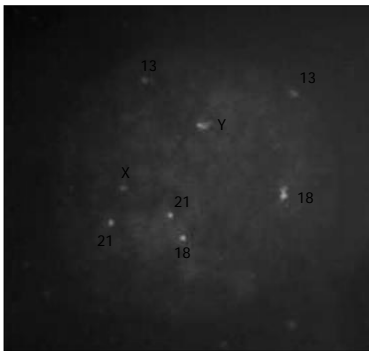
Biomarkers: Embryo Metabolism

- Net turnover of certain amino acids (Alanine, Arginine, Glutamine, Methionine, Asparagine & Leucine) by cleavage stage embryos (day 2-3) predicts blastocyst development *in vitro* (Foughton et al., 2002).
- Turnover of 3 key amino acids (Asparagine, Glycine & Leucine) on day 1-2 was significantly correlated with pregnancy and live birth (data from 52 patients) (Brison et al., 2004).



Embryo Chromosome Abnormalities

- Up to 50% of embryos produced *in vitro* have chromosomal abnormalities, including: aneuploidy and mosaicism.
- 13, 18, 21, X and Y are responsible for 65% of embryo chromosome abnormalities and >95% of chromosome aberrations in live births.
- Chromosome abnormalities cause approx. 50% of embryo wastage before implantation.
- Embryo chromosome errors can be introduced during:
 - *Oogenesis* (in cases of gonadal mosaicism)
 - *Meiosis* (segregation errors, non-disjunction, premature chromatid separation)
 - *Embryo cleavage divisions* (failed mitotic checkpoints)
 - *Maternal/Paternal balanced translocations* causing germ cell unbalanced translocations

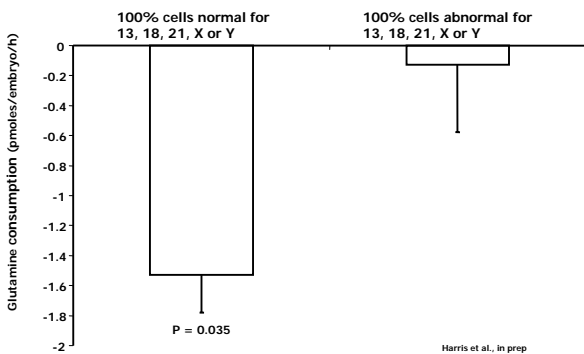


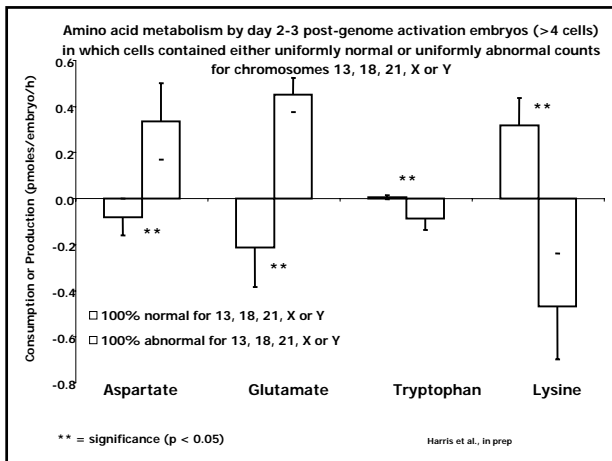
5 probe FISH on an embryo blastomere, assessing for the presence of chromosomes 13, 18, 21, X and Y.

- | |
|------------|
| Red = 13 |
| Aqua = 18 |
| Green = 21 |
| Blue = X |
| Gold = Y |

This blastomere nucleus was from a male embryo and had normal counts for the chromosomes analysed.

Glutamine consumption by day 2-3 pre-genome activation (<4 cells), non-arrested embryos at the start of 24 hours culture





- Plasma Lysine higher in children with Trisomy 21 (Lejuene *et al.*, 1992; HJ Heggarty *et al.*, 1996)
- Deficiencies in Glutathione, Serine and Tryptophan (Lejuene *et al.*, 1992)
- Increased Methionine requirement (K McLeod, 1996)
- Trisomy (chr16) in mice certain up-regulates genes involved in metabolism of Glutamate, Aspartate, Alanine, Arginine, Tyrosine, Phenylalanine & Tryptophan metabolism (Vaisman *et al.* 1981)

- ### Summary
- Metabolic and mitochondria activity may aid follicle/oocyte selection mechanisms during development *in vivo*.
 - Metabolic profiling *in vitro* may prove to be beneficial in the selection of oocytes and embryos in ART.
 - Patient groups? E.g. Diabetes, PCOS, advanced maternal age?
 - Metabolic profiling is a useful for optimisation of culture conditions *in vitro*.
 - Prospective, controlled studies: further advances?

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