

Preimplantation diagnosis

The preclinical years

Marilyn Monk

Institute of Child Health, University College London

Embryology / microsurgery

Oocytes, polar bodies, embryos and primordial germ cells.

Single cell diagnostic molecular biology

Gene mutation, modification, expression.

MOLECULAR BIOLOGY OF MICRORGANISMS

1959- 1969 Microbial Genetics

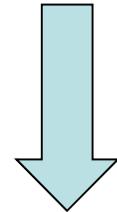
DNA replication and repair in *Escherichis coli*

1970 -1974 Development

Cell signalling and aggregation in *Dictyostelium discoideum*

MOLCULAR BIOLOGY of EMBRYOS

'Micronise' from millions of cells to one cell



1974 - Gene expression and its regulation in mammalian development

MOUSE

X Chromosome Inactivation / Imprinting / Methylation

Origin of the germ line

Deprogramming to stem cell

HUMAN

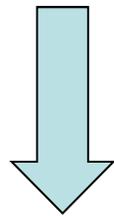
Preimplantation diagnosis

X Chromosome Inactivation/ imprinting / gene expression

Embryo and PGC cDNA libraries

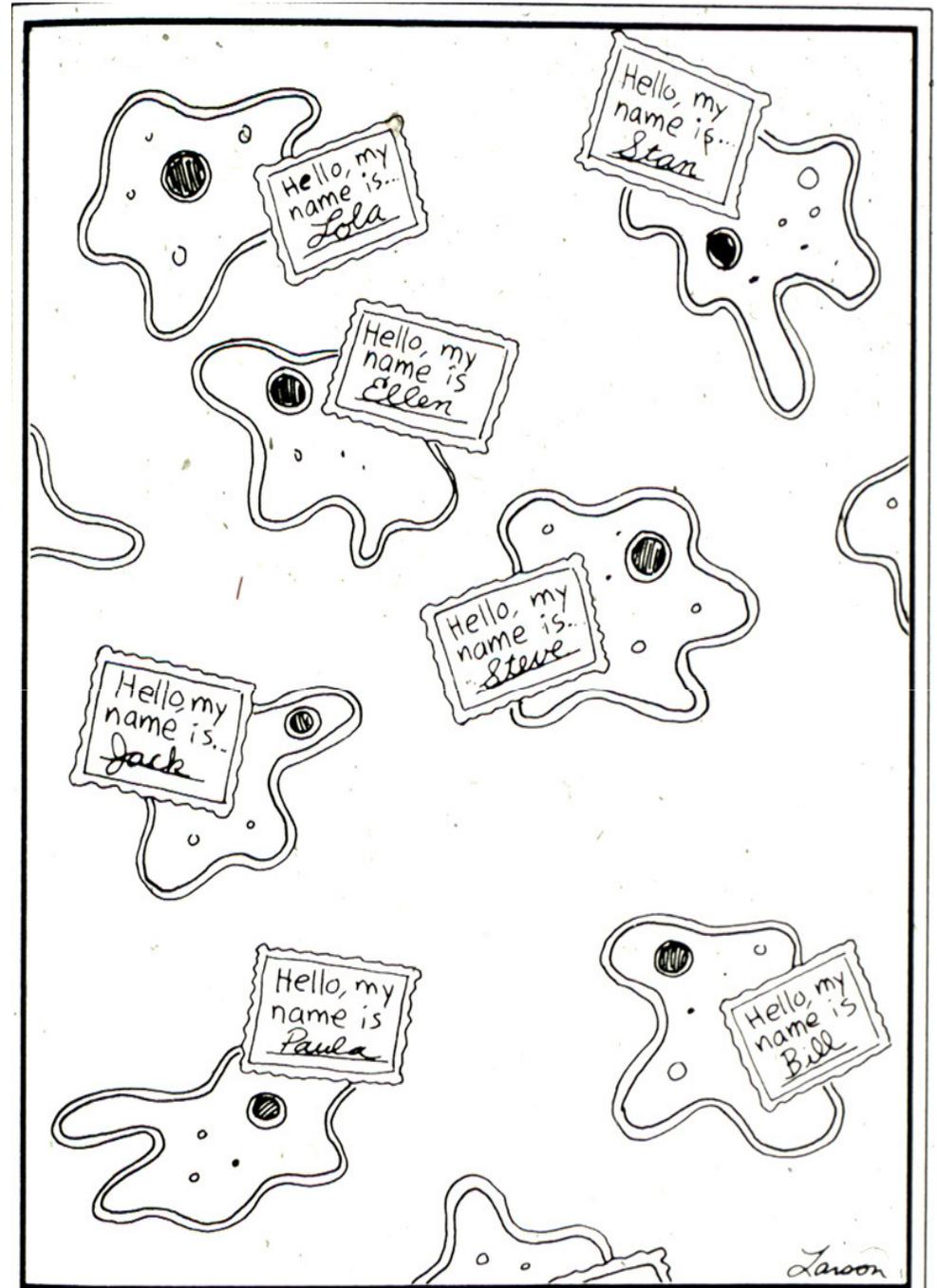
Embryo / cancer genes *ECSA*

Bored with bacteria



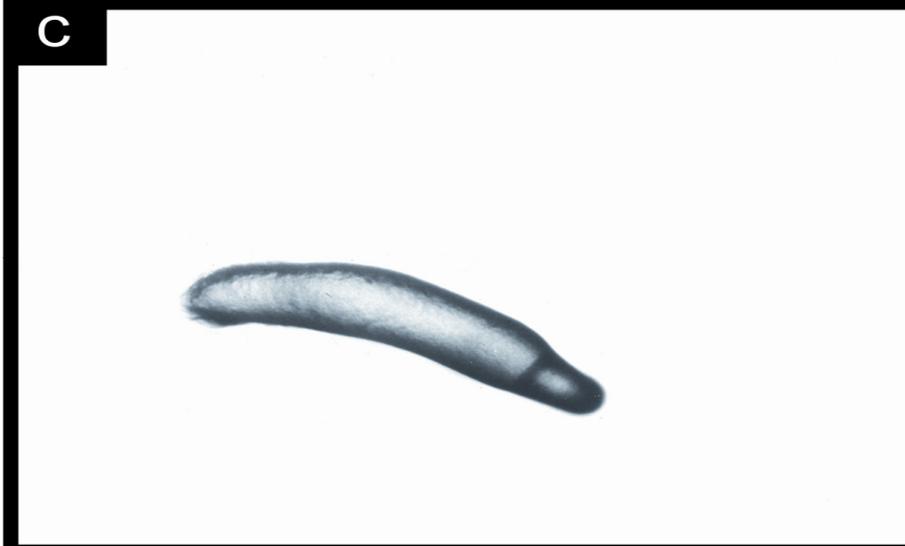
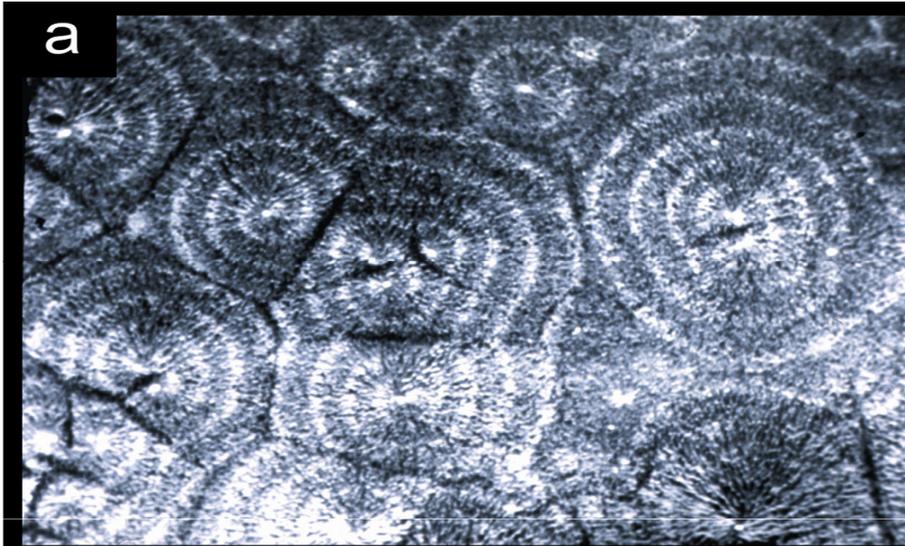
Slime mould amoebae
Dictyostelium discoideum

Cell communication
Single cells to multicellular



Amoeba conventions

Parameters of aggregation – cAMP signalling, signal periodicity, velocity - cell refractory period, movement duration & distance



1970 - 1974

Alcantara and Monk, 1974

Anne
McLaren

MRC
Mammalian
Development
Unit
1974 - 1992



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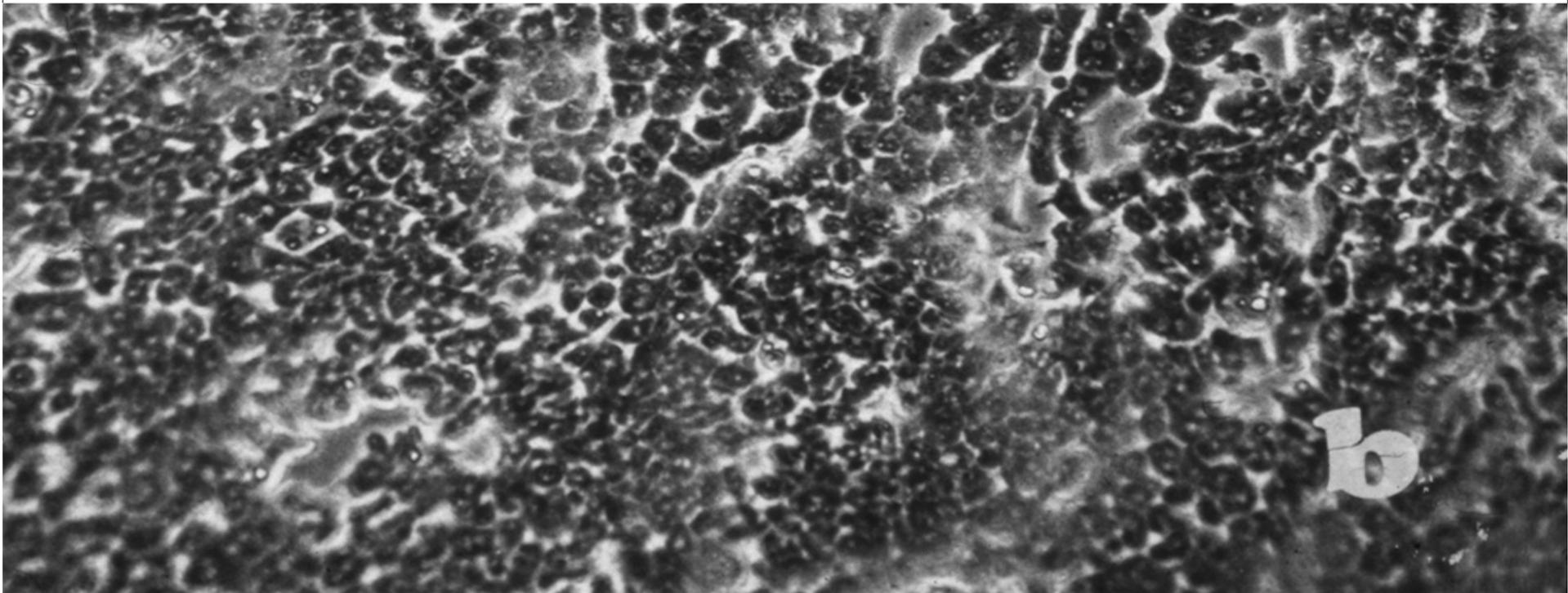
HUMAN

Preimplantation diagnosis

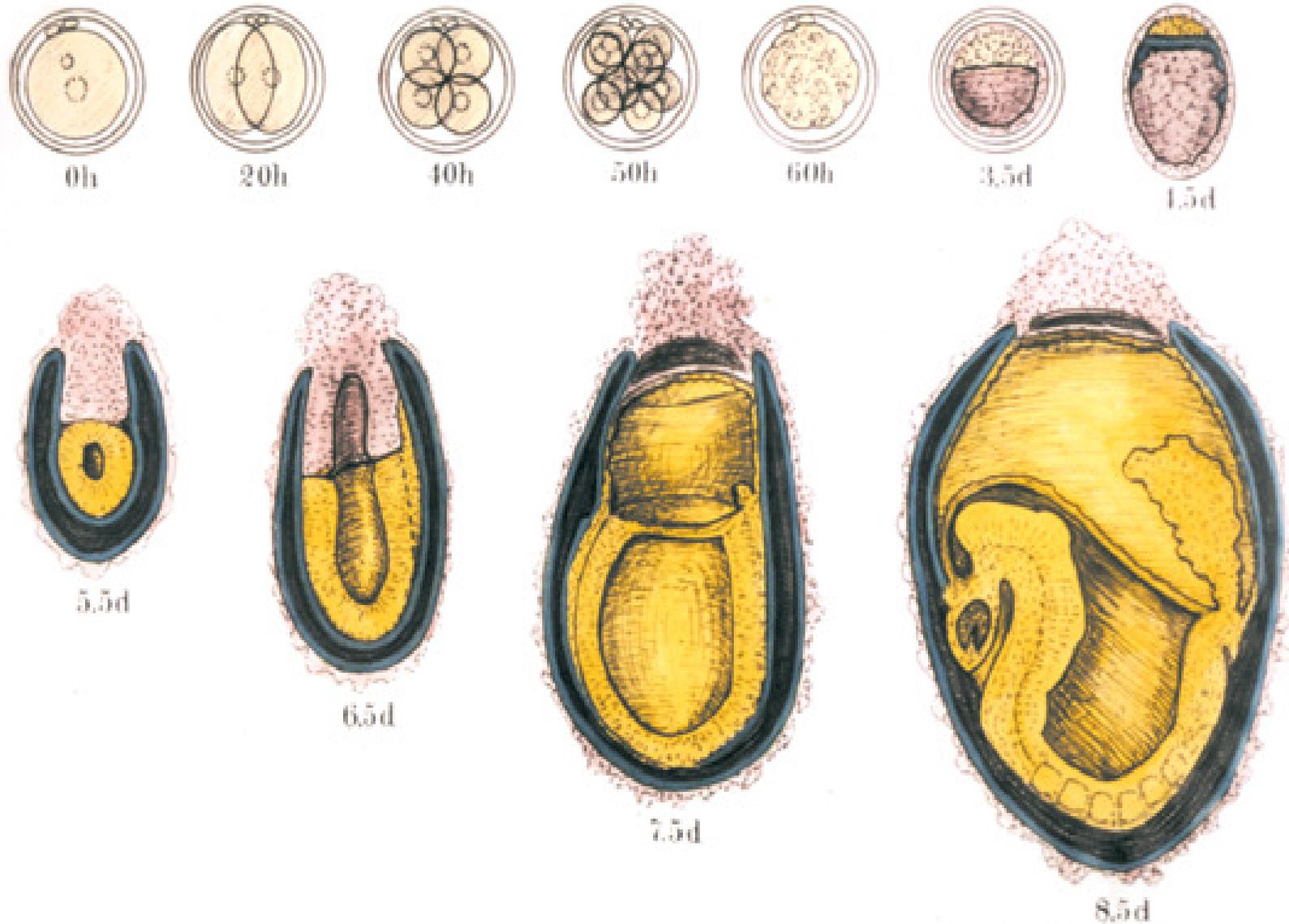
X Chromosome Inactivation/ imprinting / gene expression

Embryo and PGC cDNA libraries

Embryo / cancer genes *ECSA*



One to a few hundred cells – micro molecular biology

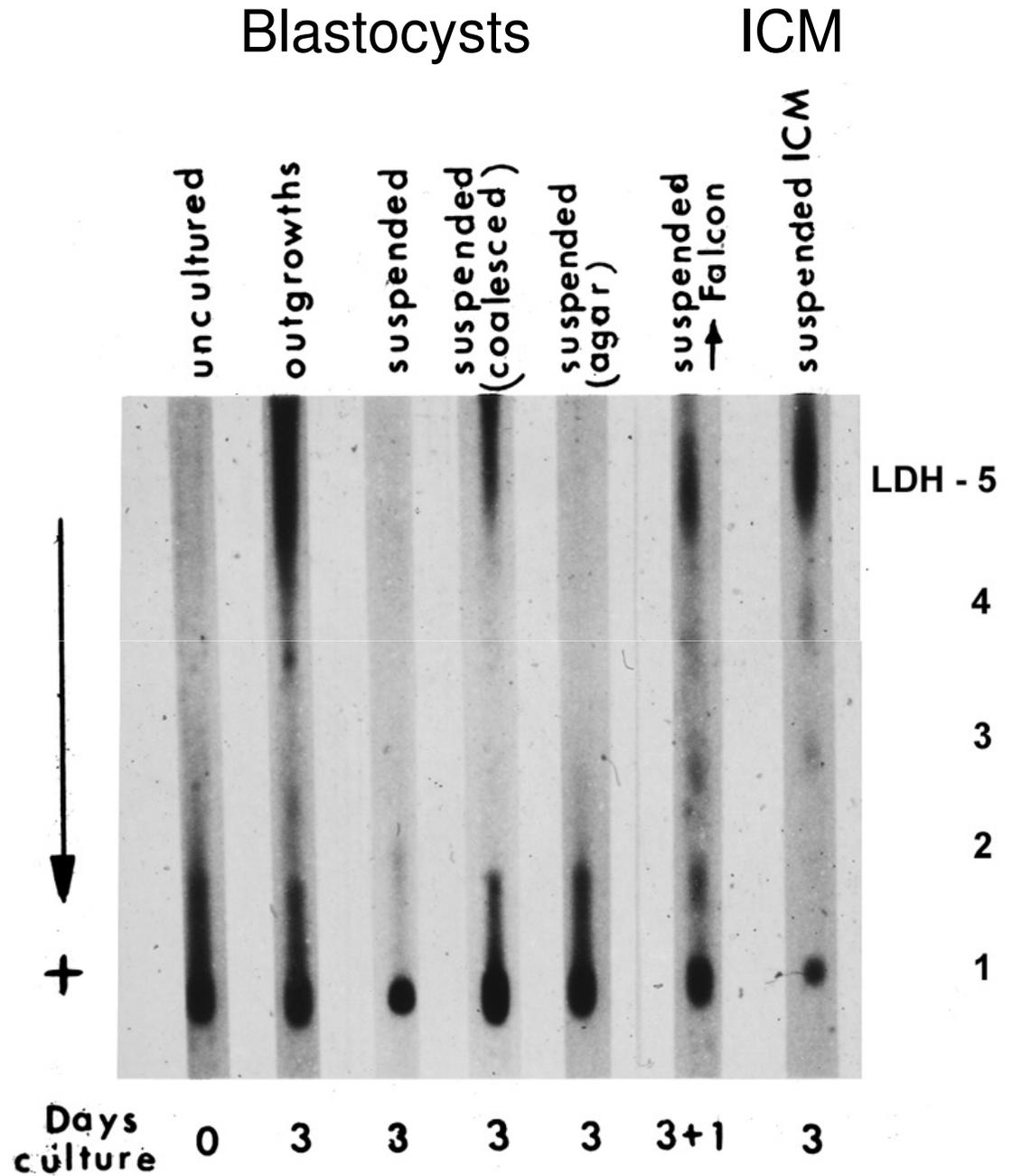


Slide of Rosa Beddington

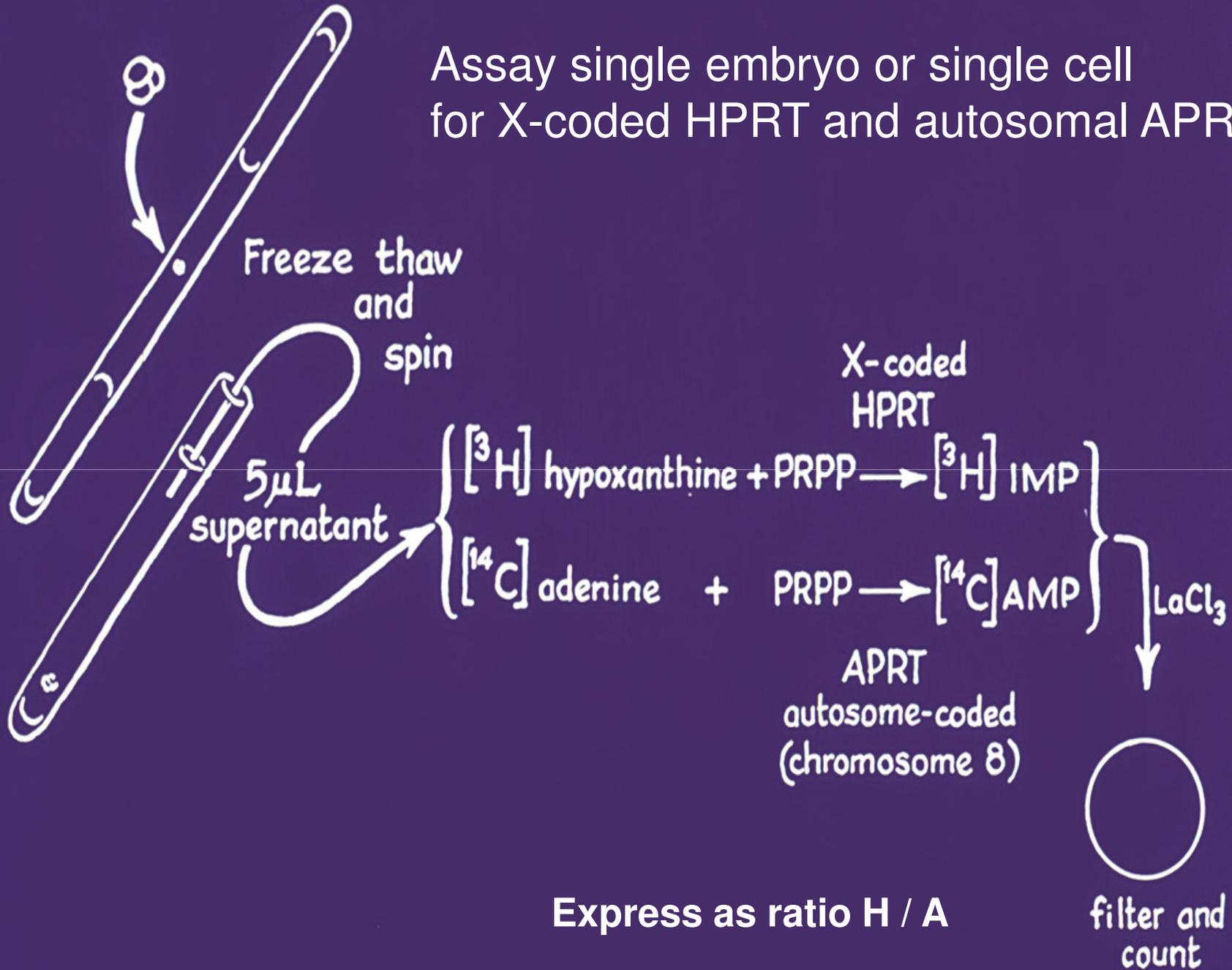
Electrophoresis
of LDH
in single embryos
in 2 uL minigels

Delayed
'implantation'
in vitro

Monk and Petzoldt
Nature 1977



Assay single embryo or single cell for X-coded HPRT and autosomal APRT



Express as ratio H / A

filter and count

Monk and Kathuria Nature 1977

Ratio $\frac{\text{HGPRT}}{\text{APRT}}$

$\frac{\text{HGPRT}}{\text{pM}} \frac{\text{hr}}{\text{egg}}$

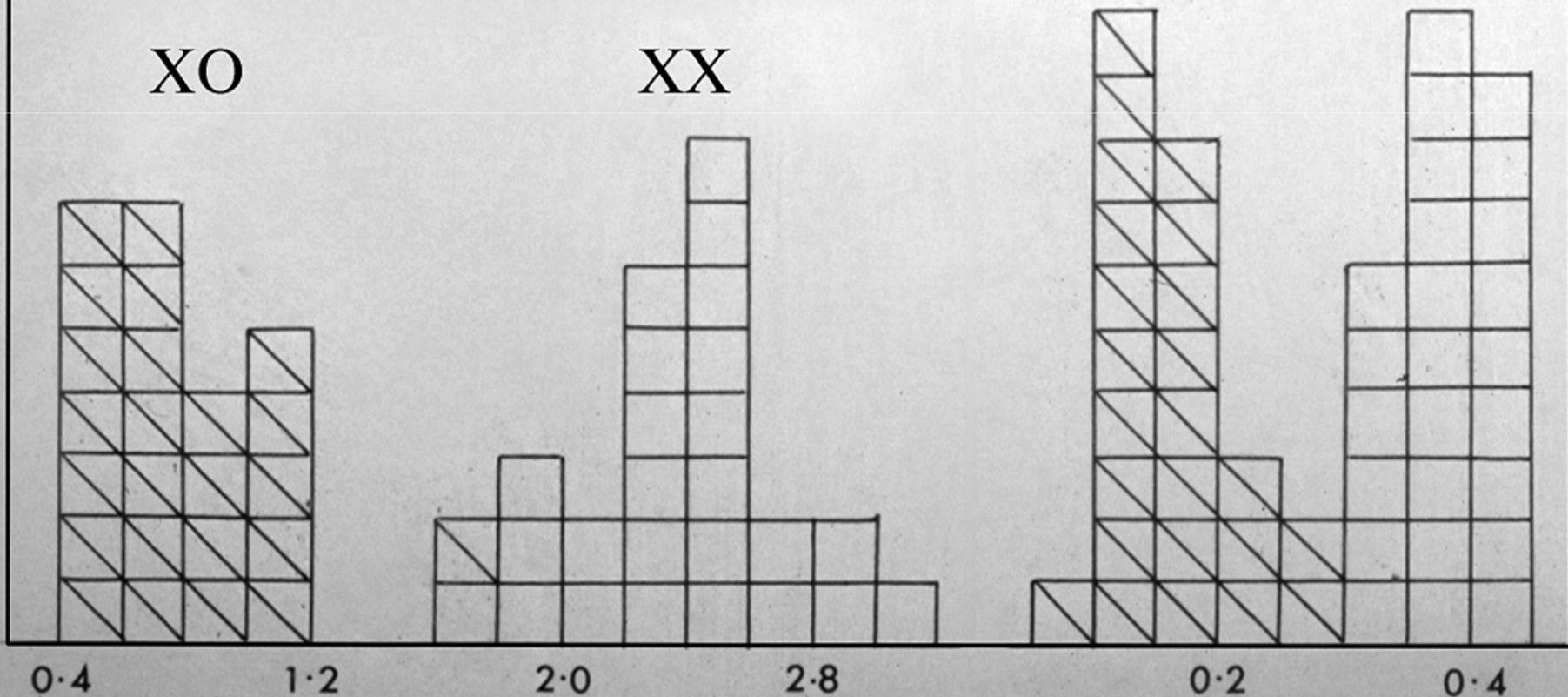
Two X chromosomes are active in single oocytes

XO

XX

XO

XX



Postimplantation - 6d

Dosage of HPRT is **bimodal**

6d epiblast

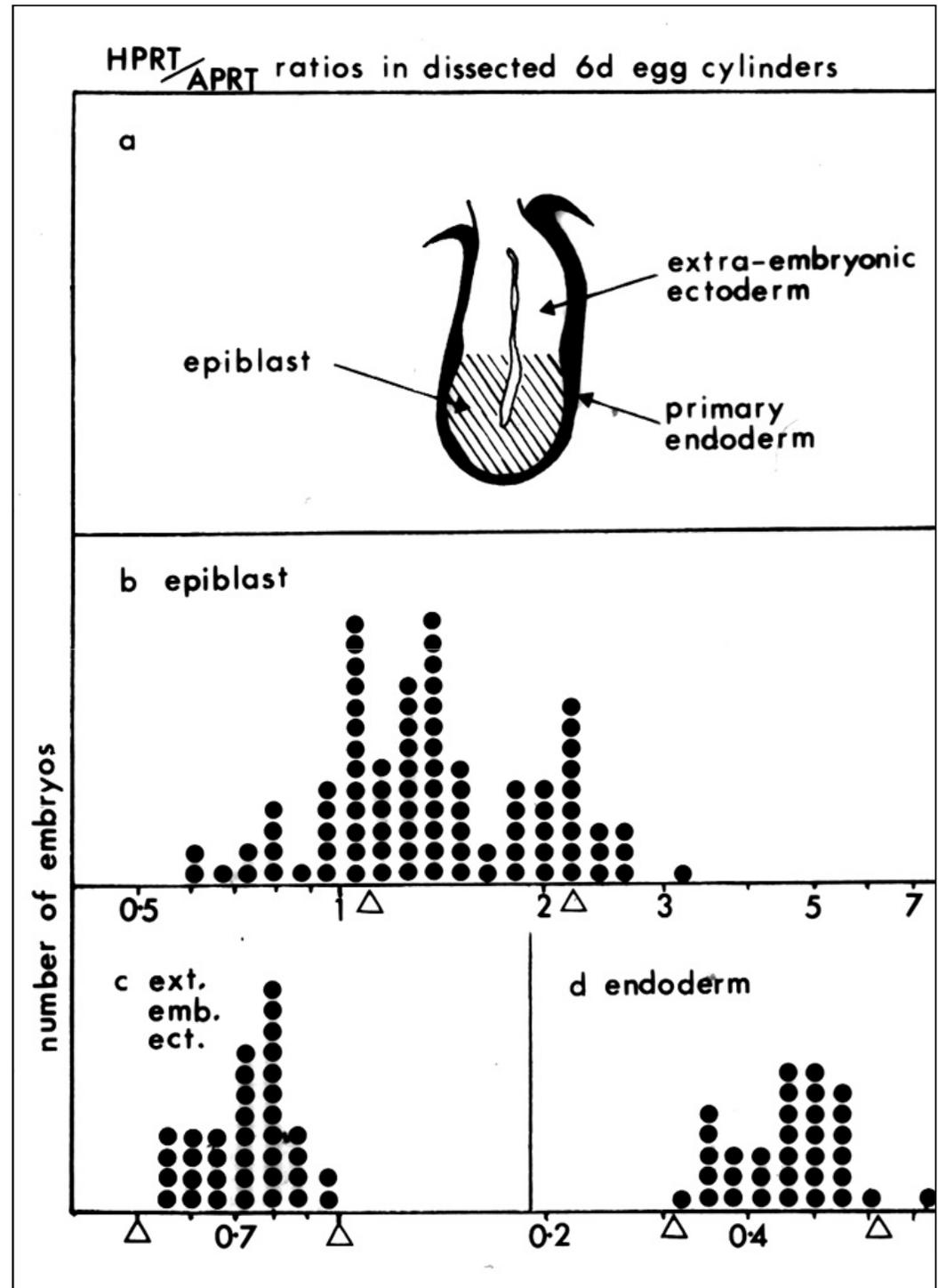
Two X chromosomes active

Dosage HPRT unimodal

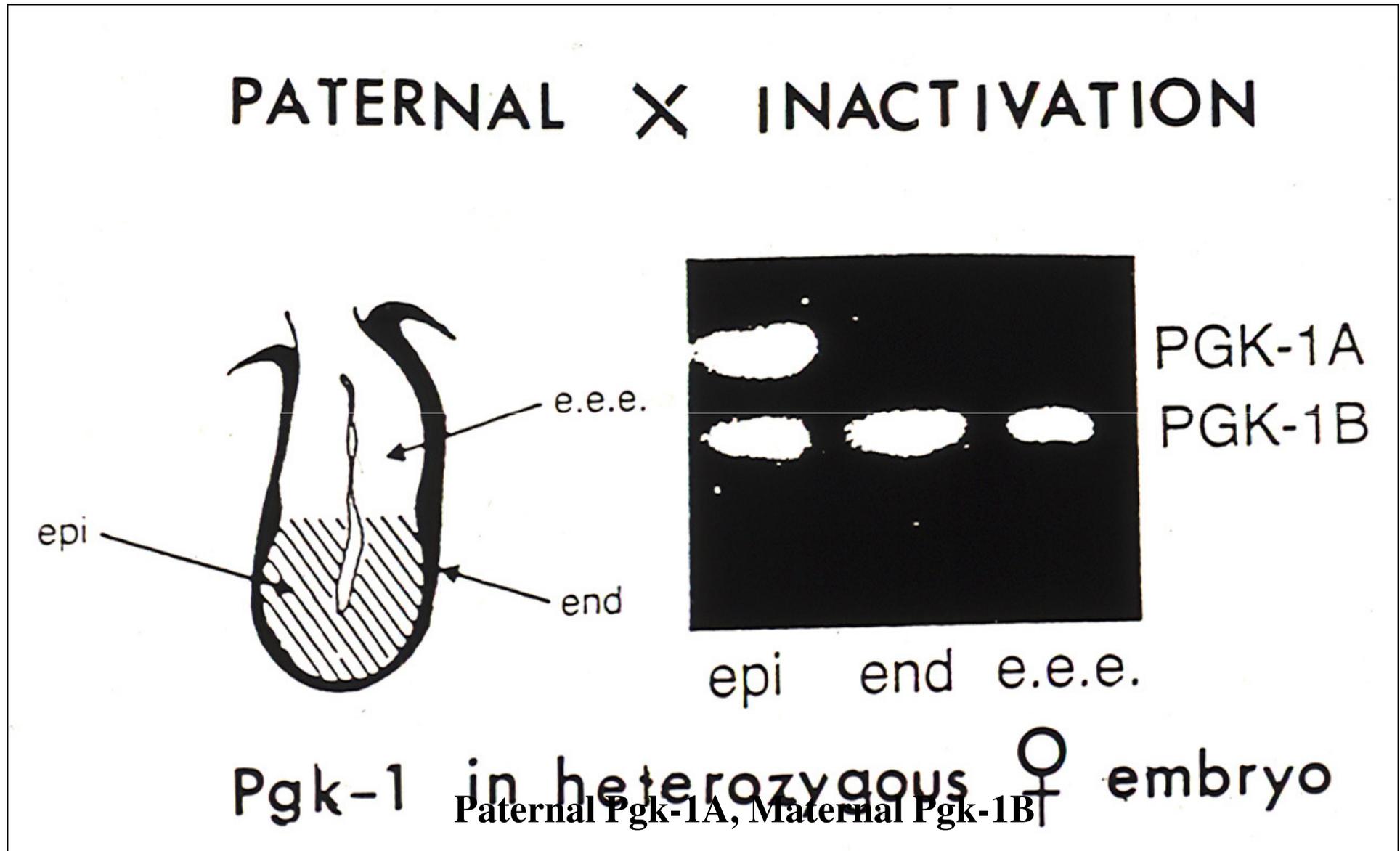
6d ex emb ect & ex emb end

One X active

Monk & Harper, Nature 1979

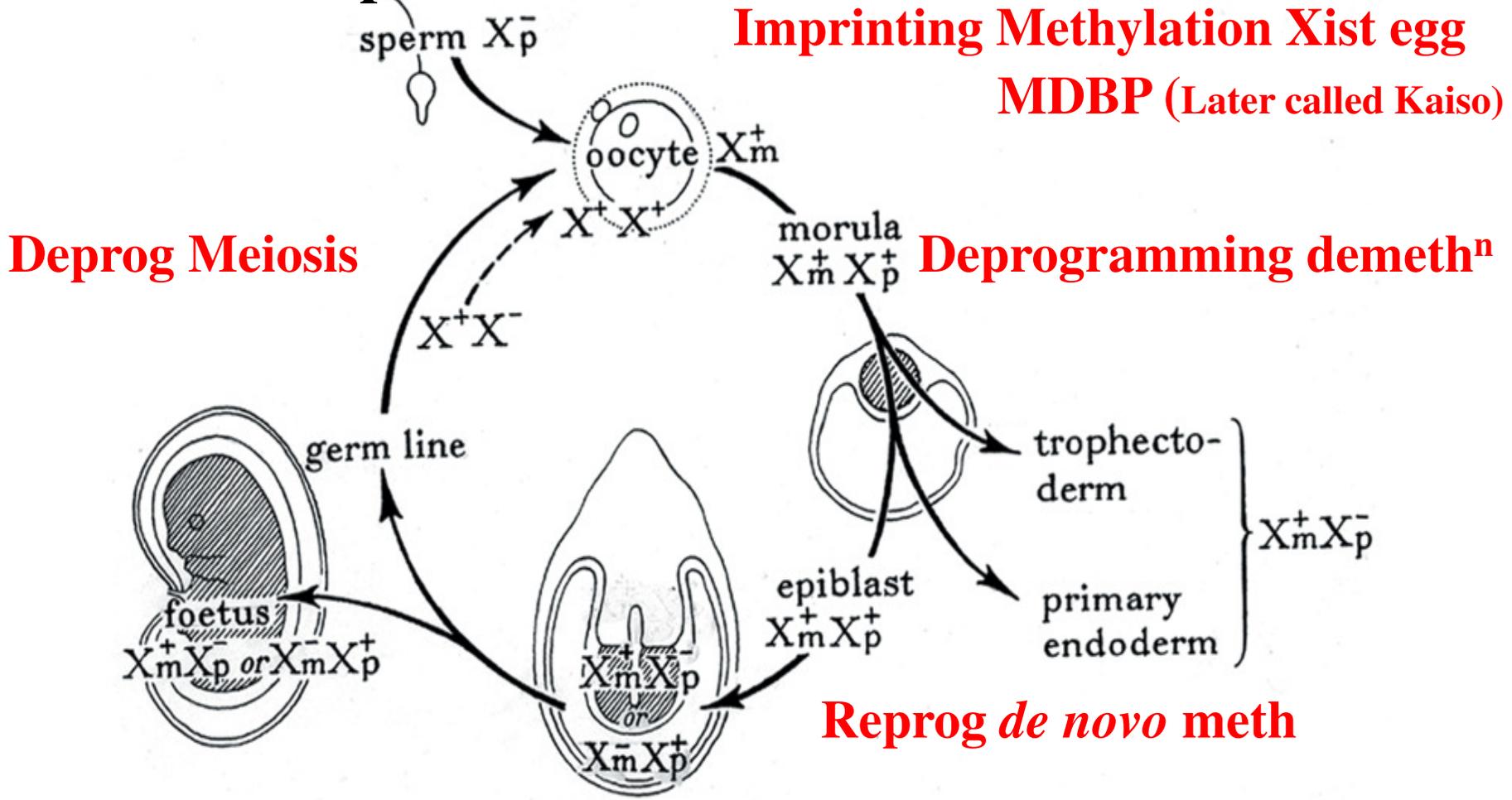


Primary non-random paternal X-inactivation



McMahon, Fosten & Monk, 1981; West & Pappaiannou. 1981

XCI in development

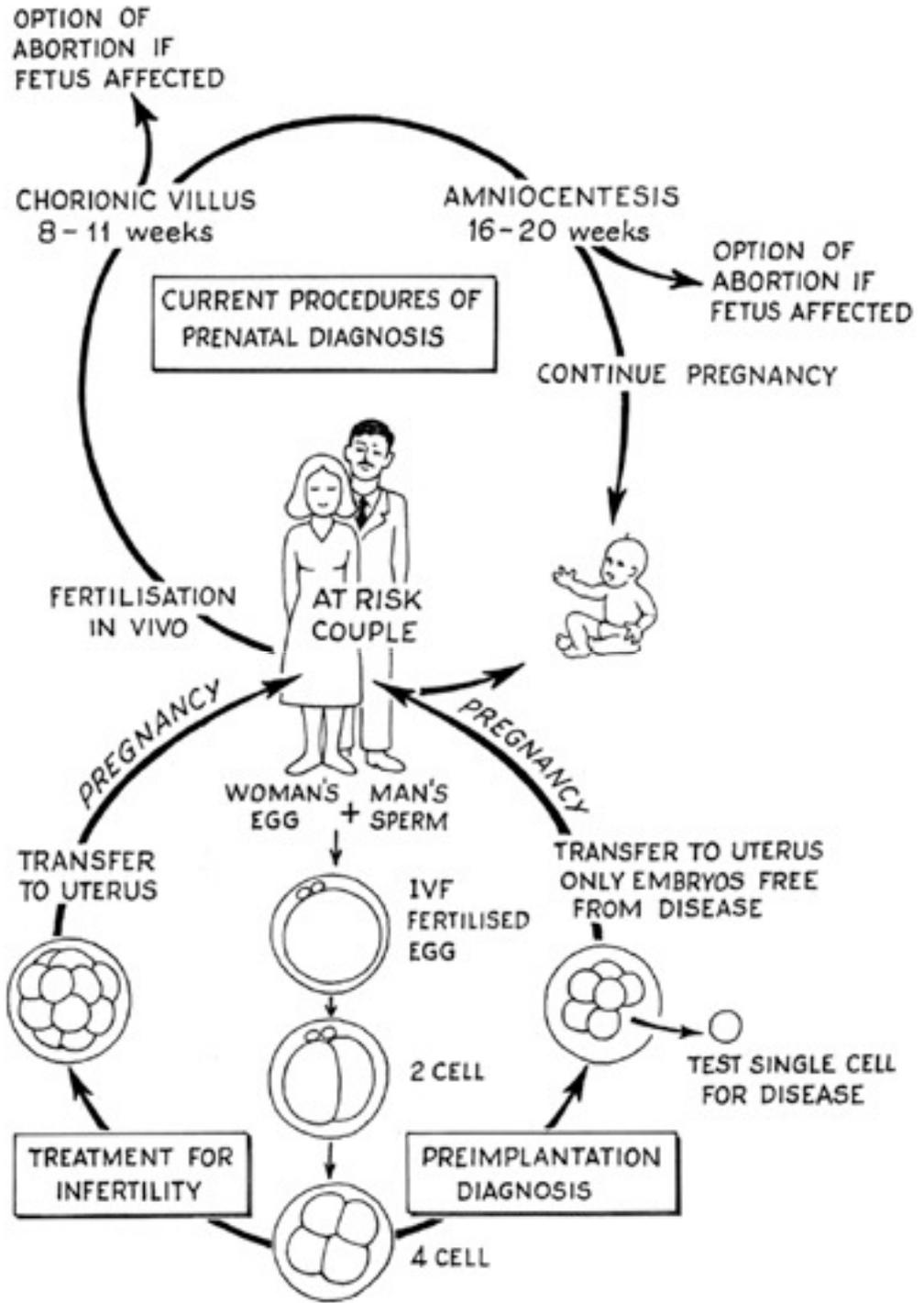
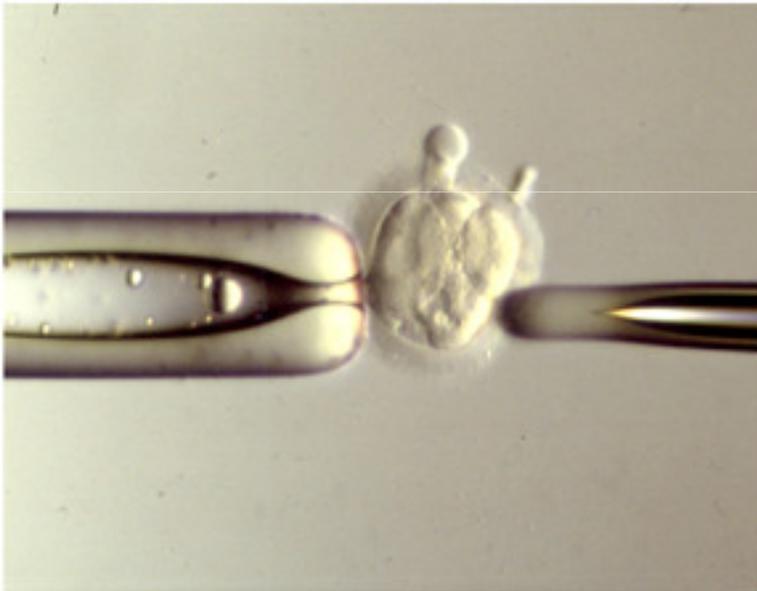


Late origin of the germ line

Monk et al 1977 - 1983

PID → PGD

Biopsy of a blastomere from 8-cell embryo (Tetsuya Goto)



IVF

HPRT deficiency → Lesch Nyhan

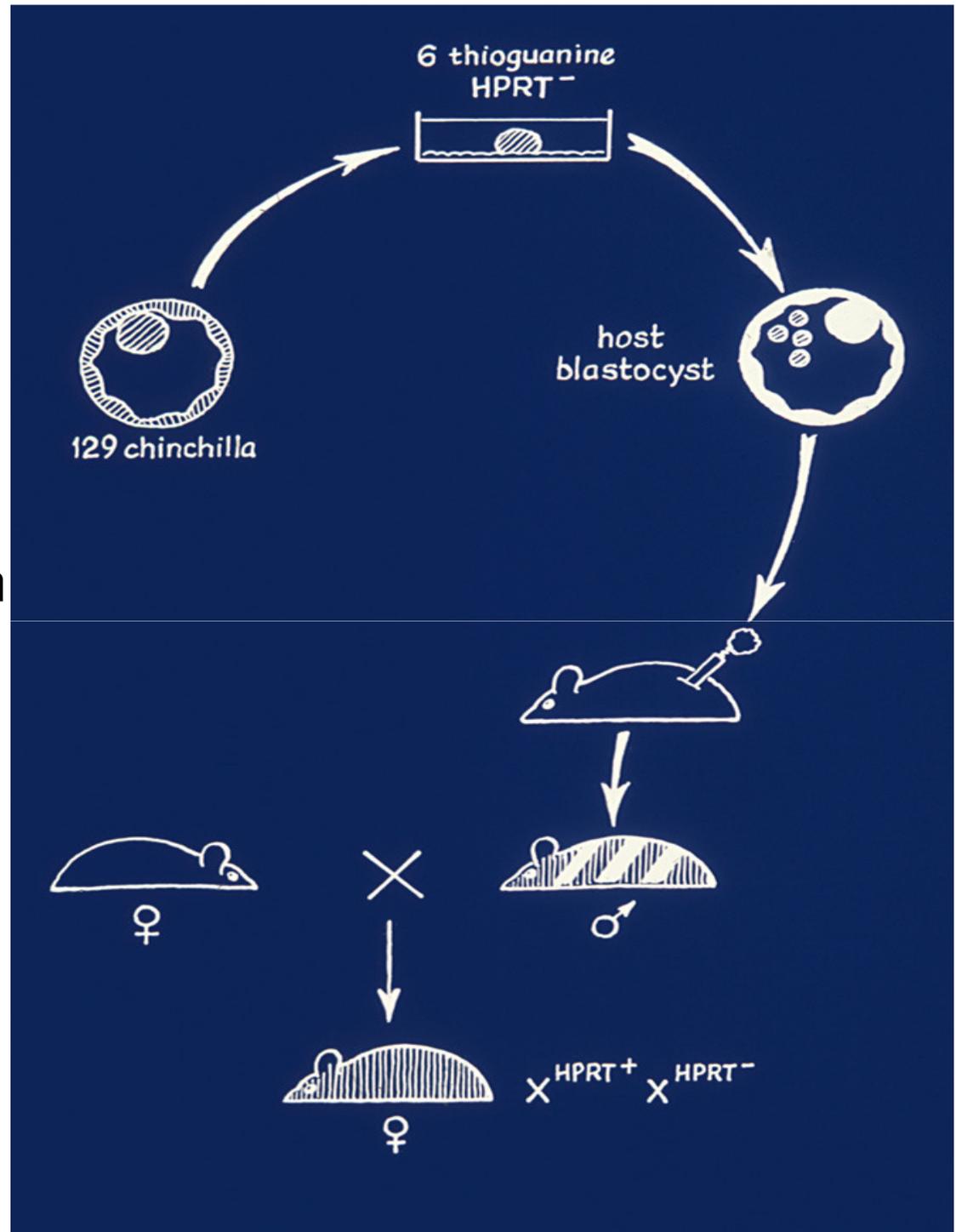
The first genetically engineered mouse

HPRT-deficient (Lesch-Nyhan) mouse embryos derived from germline colonization by cultured cells

Hooper M, Hardy K, Handyside A, Hunter S, and Monk M.

Nature **326**, 292-5 (1987)

Hprt⁺ / Hprt⁻ female →



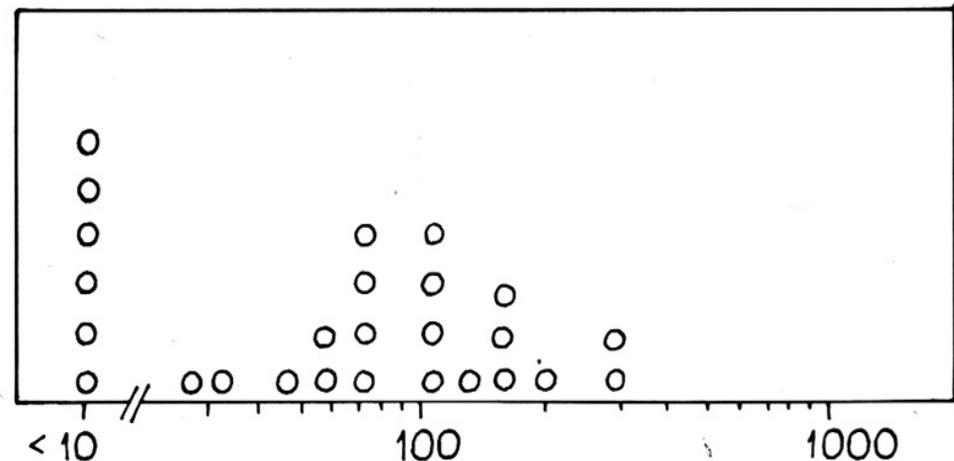
Embryos from Hprt+/Hprt- mother
'Lesch Nyhan' mouse model

Diagnosis
of Hprt deficient embryos
by HPRT enzyme activity
in single blastomere
from 8-cell embryo

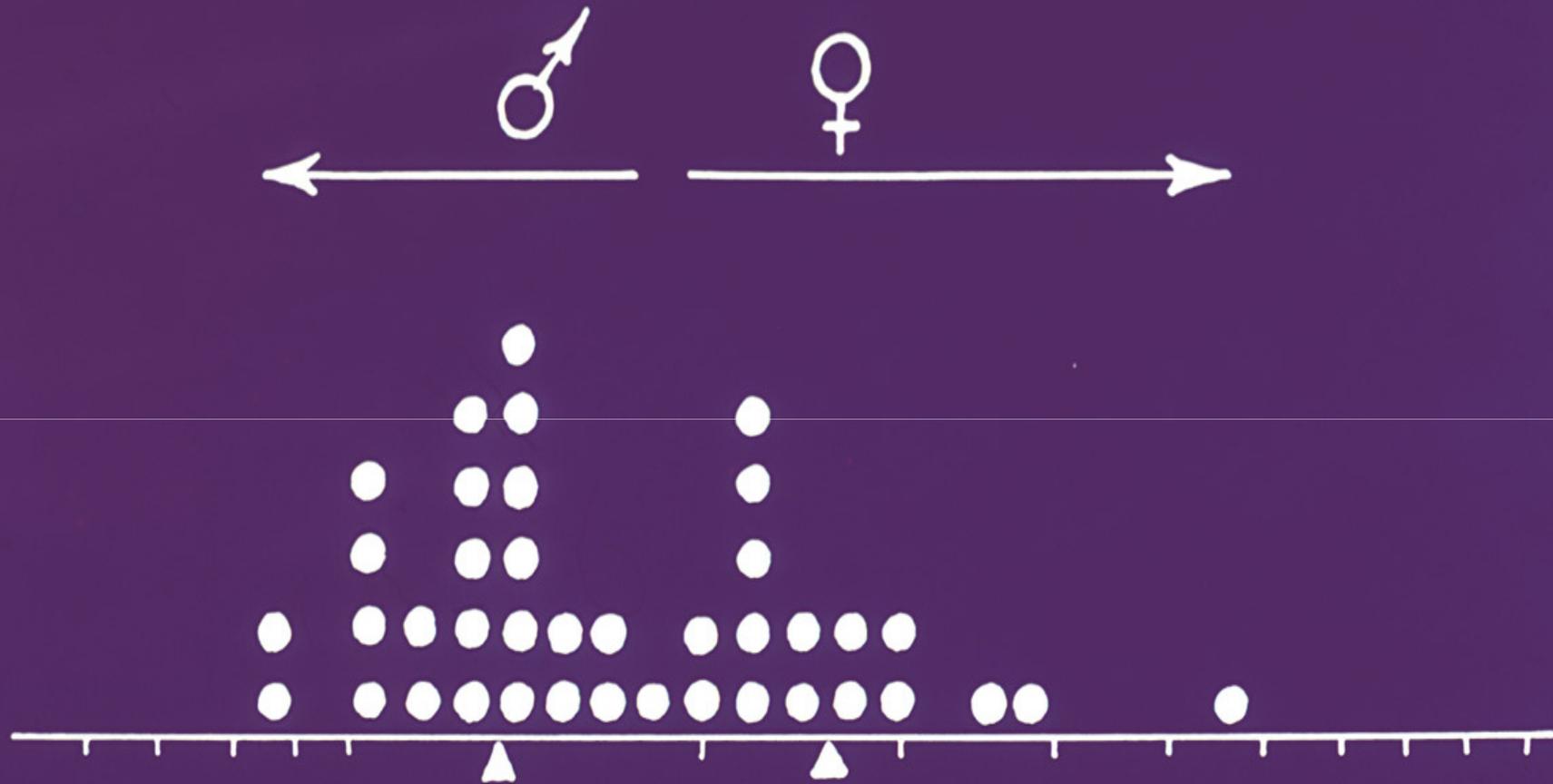
Confirm diagnosis in
transferred biopsied embryos
(A Handyside)

Monk et al, Lancet 1987

HPRT in single blastomeres from 8-cell embryos



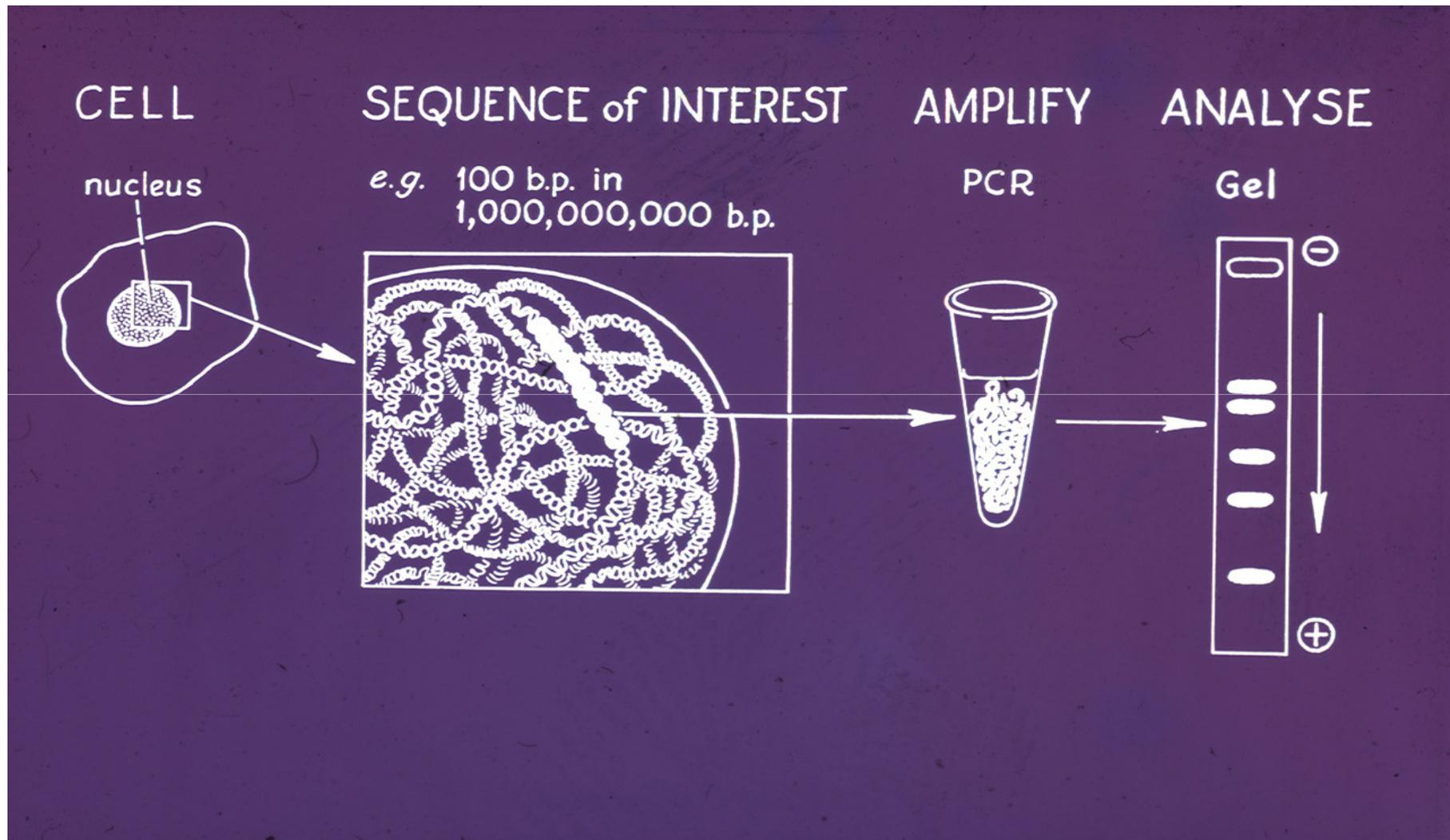
Sexing by Hprt dosage



HPRT : APRT ratio

Monk & Handyside 1988

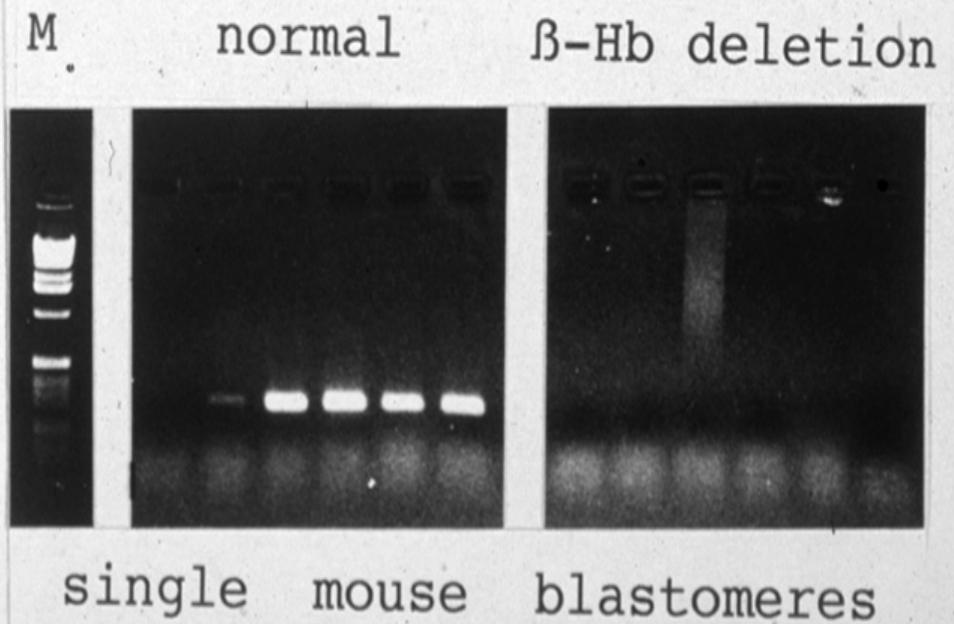
Single Cell PCR



Moving racks of tubes between water baths! – Holding and Monk



Holding and Monk, Lancet 1989



Harper
Fosten
McMahon
Lindsay
Holding
Ao
Grant
Kenealy
Zuccotti
Daniels
Kontogianni
Thornhill
Goto
Adjaye
Lorenzo
Huntriss
Salpekar
Hitchens

Monogenic disease genes

SINGLE CELL ANALYSIS

X - Linked

Duchenne Muscular Dystrophy*
Lesch - Nyhan Disease*
Severe Combined Immunodeficiency*
Fragile X Syndrome*
Kennedy's disease*
X-inactivation specific transcript*
Mono-amine oxidase A*
Sex chromosome-linked repeat sequences*
Haemophilia

Autosomal

Sickle Cell Disease*
Myotonic Dystrophy*
Small nuclear riboprotein polypeptide N*
Antitrypsin Deficiency
Thalassaemia
Cystic Fibrosis
Tay Sachs Disease

***At Molecular Embryology Unit, ICH**

INHERITED GENETIC DISEASE

OOCYTE DIAGNOSIS

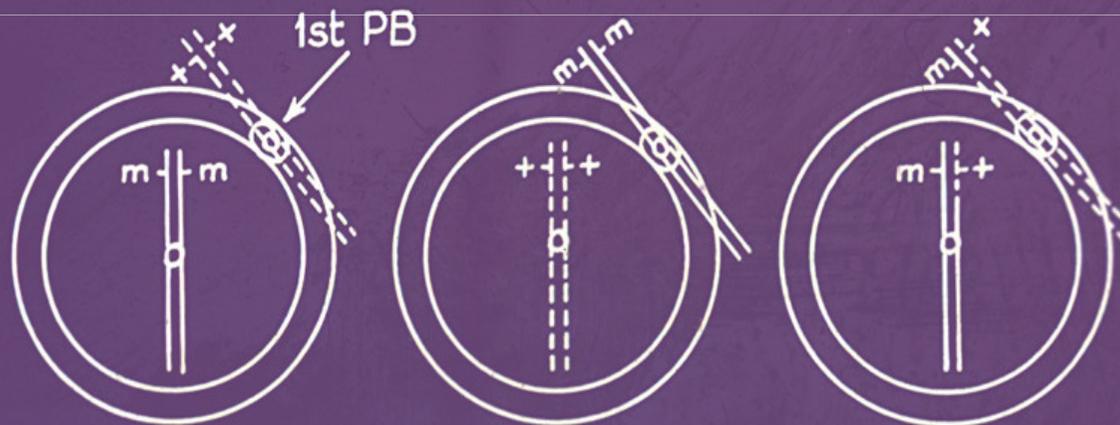
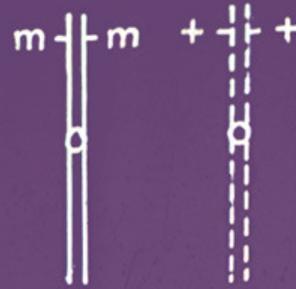
Biopsy of first polar body

Fertilise only eggs without defect

Avoids interference with embryos

Diagnosis of egg by examination of polar body

Carrier mother



homozygous
egg will be
mutant

homozygous
egg will be
normal

heterozygous
egg may be
mutant or normal

Diagnosis of sickle cell in human polar body

Separate polar body and egg (Peter Braude's lab)

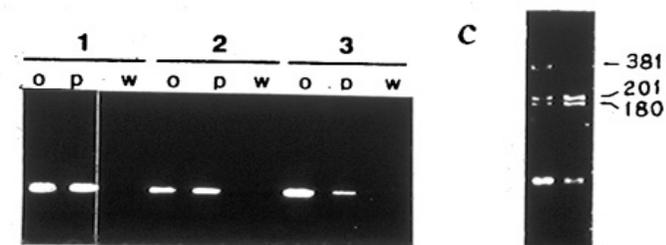
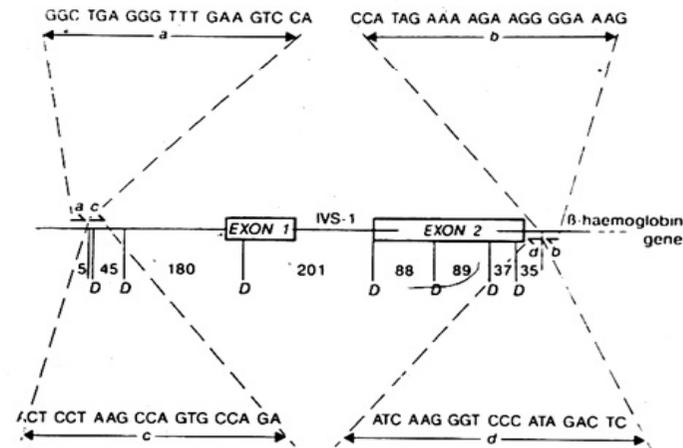
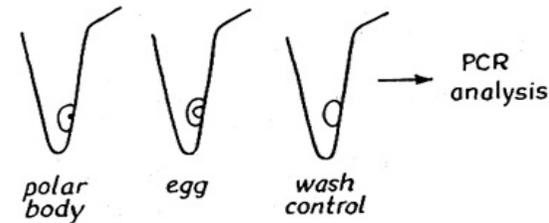
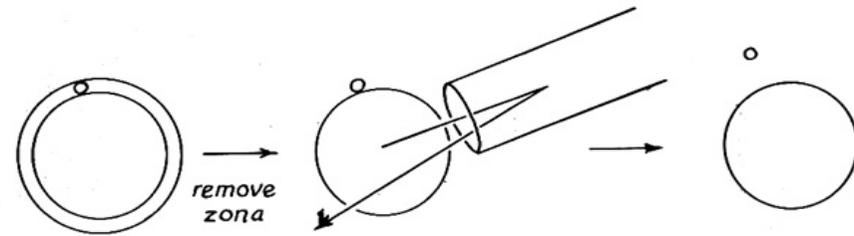
Nested PCR amplification of beta haemoglobin sequence over sickle cell mutation

in

Individual human oocytes and polar bodies.

Monk M and Holding C.
Lancet **335**, 985-8 (1990)

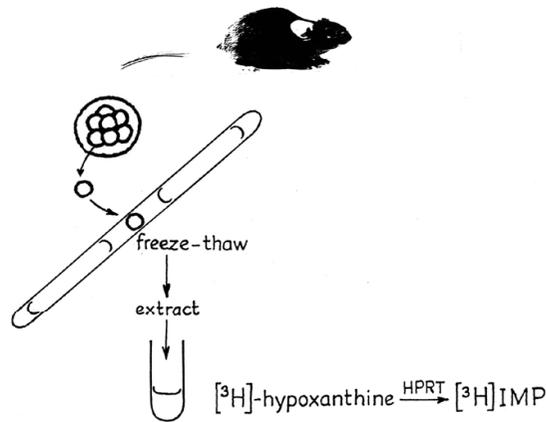
OOCYTE DIAGNOSIS



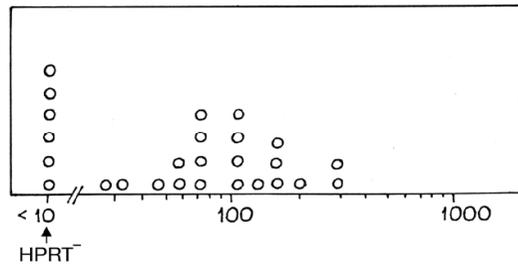
Early Models for Preimplantation Diagnosis

Summary

a. Lesch-Nyhan $X^{HPRT+} X^{HPRT-}$

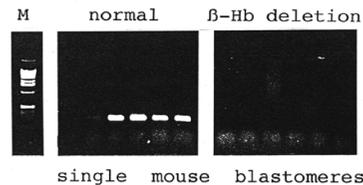
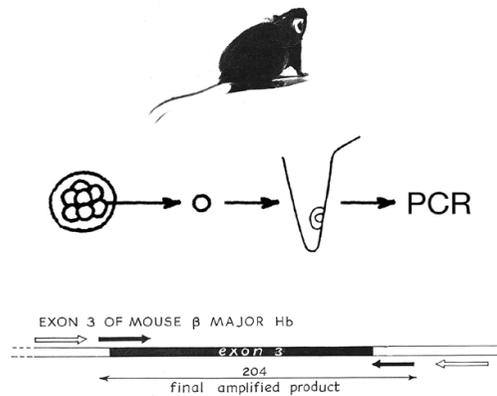


HPRT in single blastomeres from 8-cell embryos



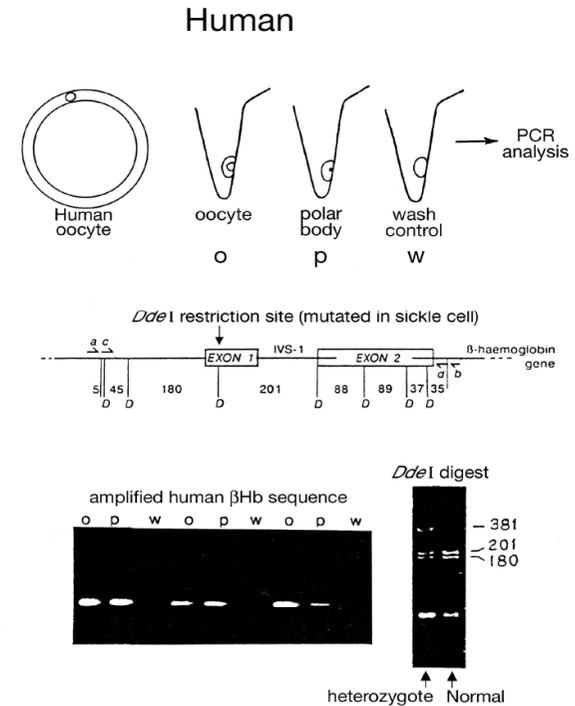
Monk et al, 1987

b. Thalassaemia $\beta Hb^+ / \beta Hb^\Delta$



Holding & Monk, 1989

c. Sickle cell anaemia



Monk & Holding, 1990

QUALITY CONTROL

Efficiency of amplification in single cell

Efficiency of detection of both alleles

Extent of contamination

"Fingerprint" origin of cell

Monk, Kenealy and Mohadjerani 1993

Cell Recycling

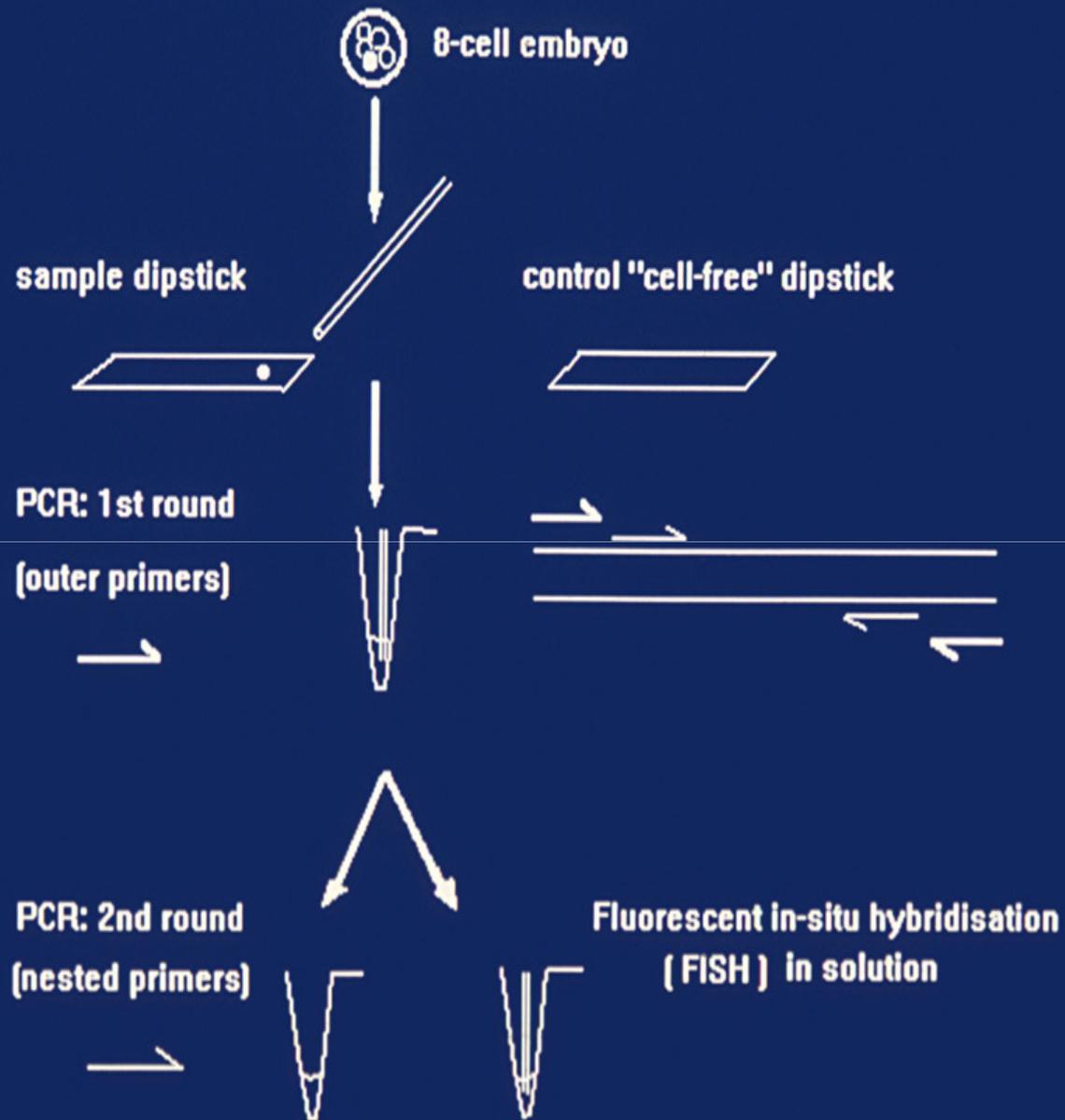
**A method to analyse both chromosomes
and the DNA of specific gene sequences**

in the same single cell fixed to glass.

FISH plus PCR

Holding, Thornhill & Monk 1994, 1996

RECYCLING THE SINGLE CELL

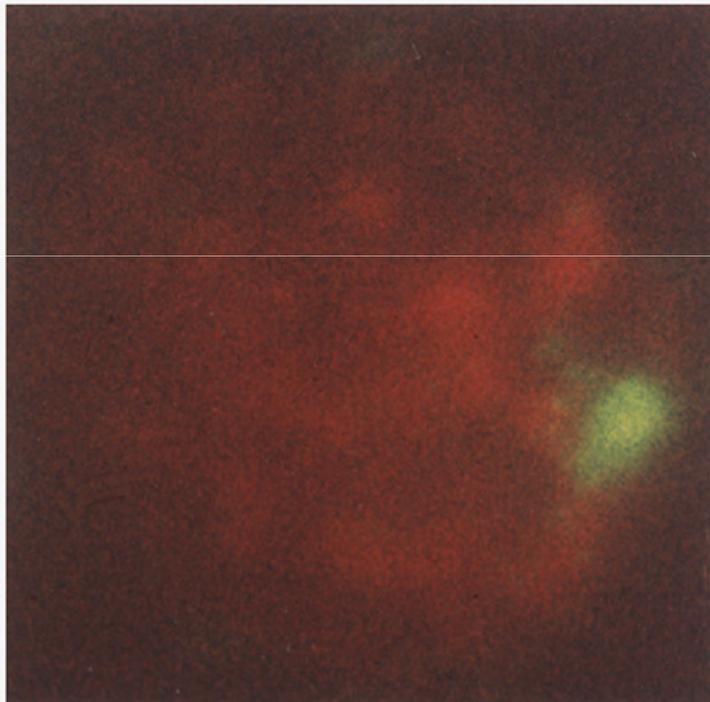


CELL RECYCLING

chromosomes and genes in a single cell

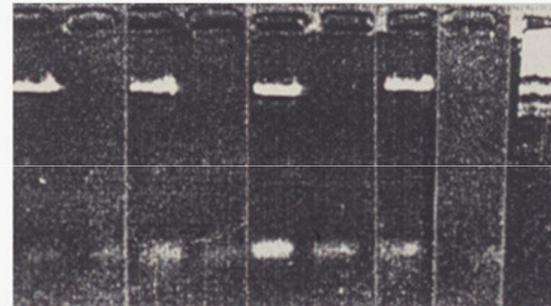
SINGLE BLASTOMERE

Y-CHROMOSOME BY FISH



B-GLOBIN SEQUENCE BY PCR

BL C BL C BL C BL C M



Results for 4 individual
blastomeres fixed to glass

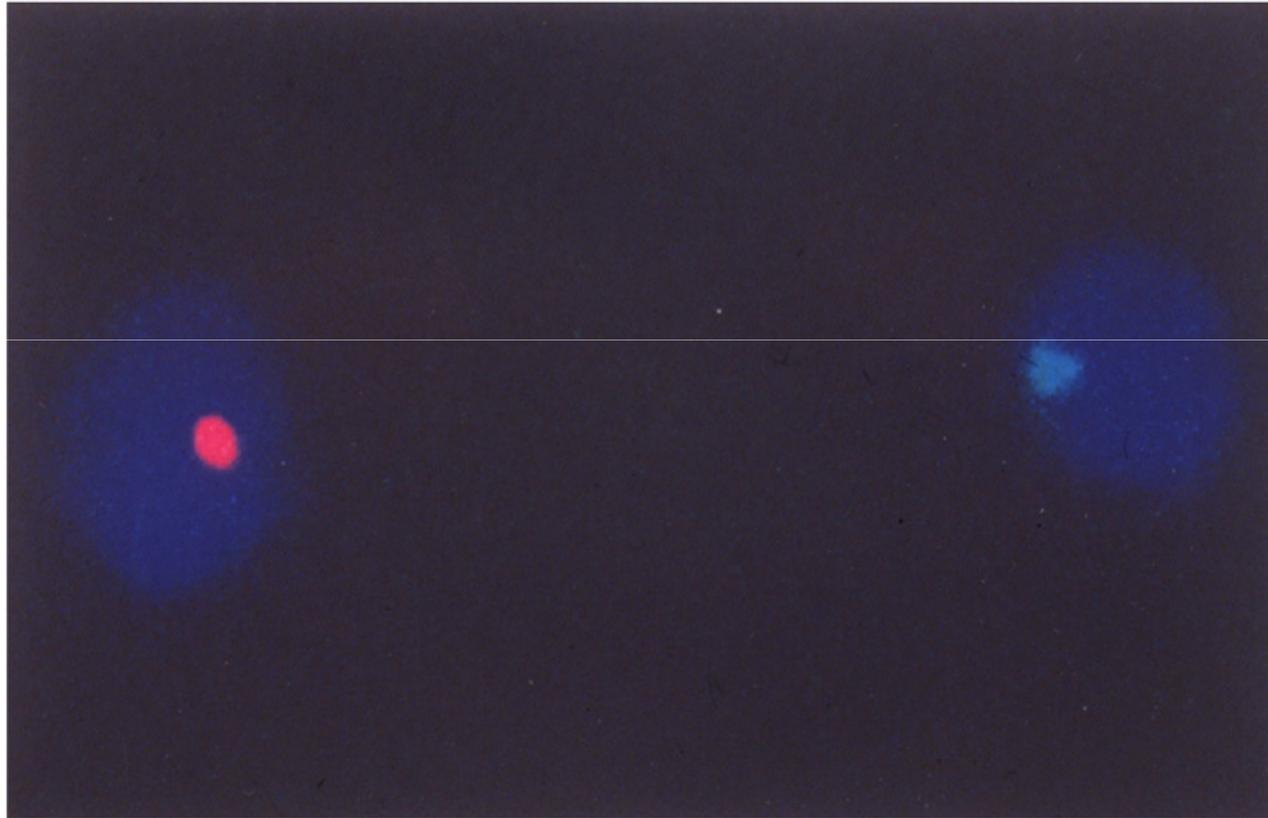
Thornhill et al, 1993

Also sexing and DMD, Thornhill and Monk 1996

X OR Y CHROMOSOME IN SINGLE HUMAN SPERM

49.0% X

49.2% Y



Thornhill, 1993

Gene expression and its regulation in embryos and germ cells

- **Direct analysis (one or two genes per embryo)**

Expression of known genes

Expression of imprinted genes

Epigenetic mechanisms of regulation of expression – CpG methylation

- **Embryo and PGC amplified cDNA (unlimited analysis)**

Comparison of expression of many genes within and between embryos

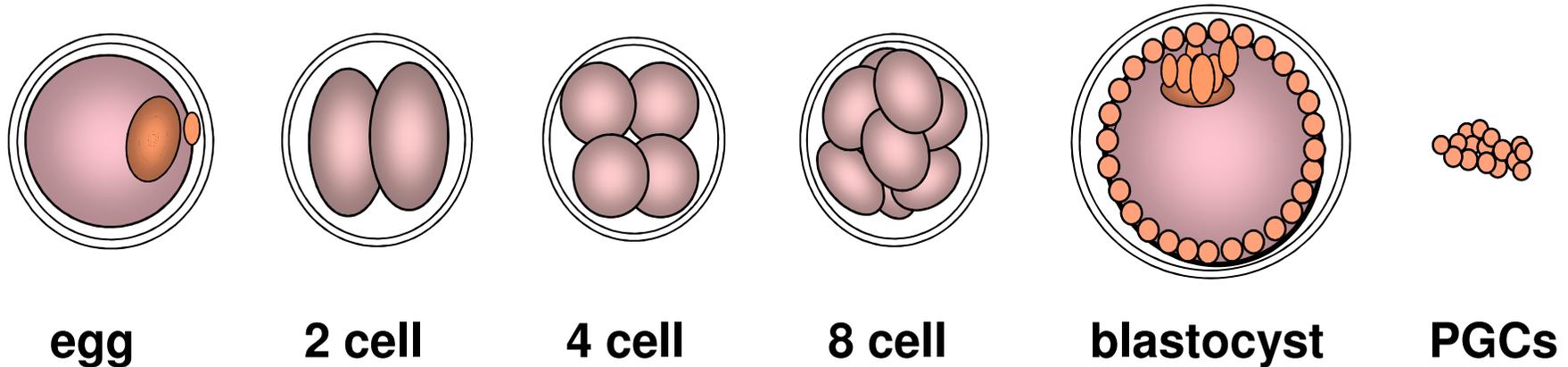
Identification of stage specific and treatment specific genes

(GV, MI, MII, embryos and PGCs +/- S/O +/- IVM Marker genes)

Isolation of novel genes - oocyte-, embryo-, PGC-specific expressed genes

Isolation of embryo / cancer genes

cDNA libraries from single embryos **a limitless resource of all the expressed genes in**



Human eggs and embryos, surplus to IVF requirement and donated for research.

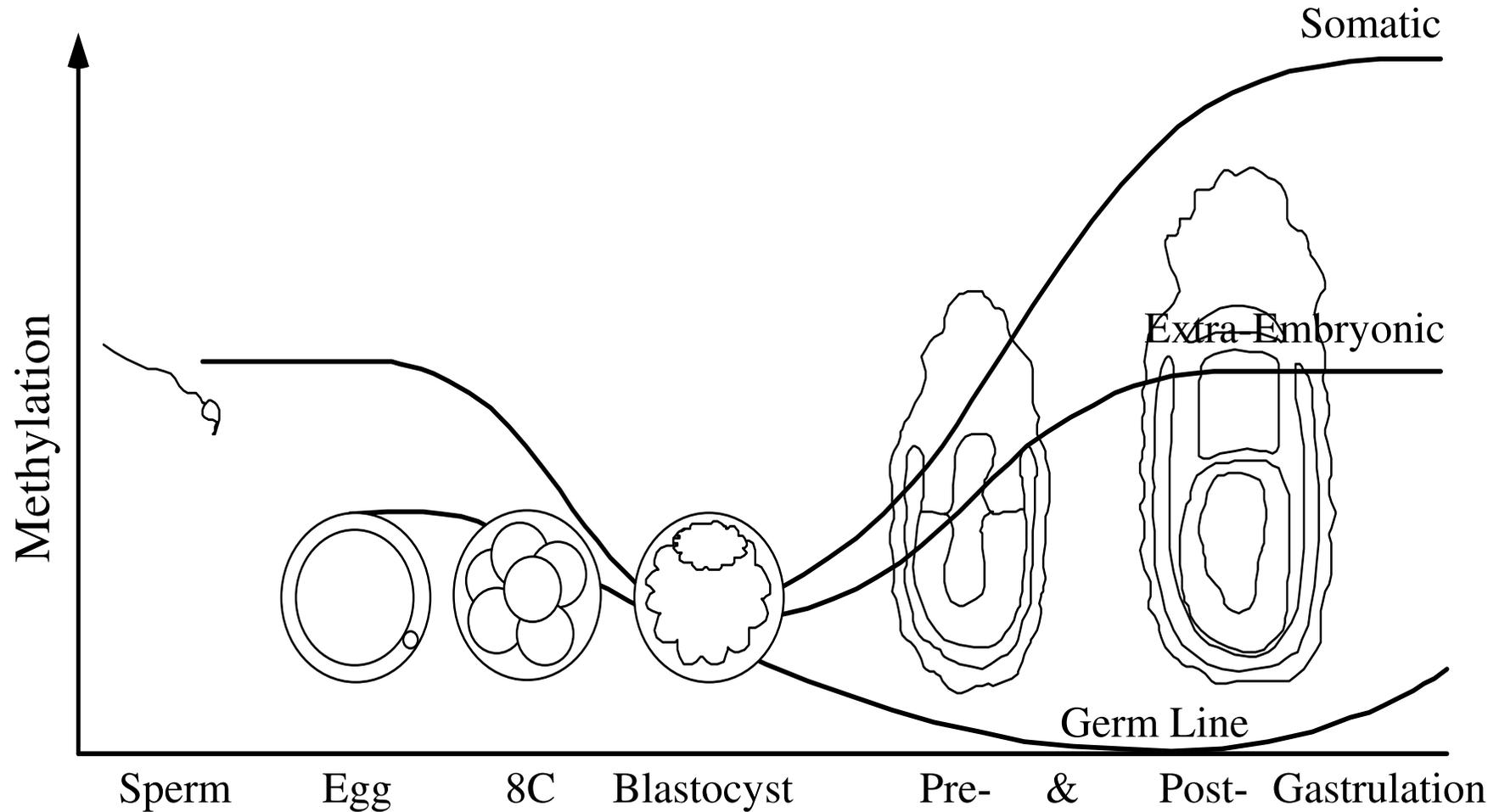
Human fetal primordial germ cells, EC and ES cells

Lysed to release mRNA which is copied and amplified as cDNA

cDNA is cloned into vector for libraries

cDNA screened for known and novel embryonic genes

Demethylation deprogramming and reprogramming in development



Monk, Boubelik & Lehnert 1987

Genes specifically expressed in human embryonic cells?

Initiation de-programming to proliferative stem cell

De-programming erasure gametic epigenetic programmes

Maintenance undifferentiated (archetypal) stem cell state

Immortality Embryo stem cells removed from

developmental constraints grow indefinitely

ICM \rightarrow ES PGC \rightarrow EG

Invasiveness e.g., trophectoderm, PGC migration (*OLF-R?*)

Expression expected to be specific to embryos and germ cells

Human embryonic stem cells

Globally demethylated,

Self renewing and grow indefinitely *in vitro*

Undifferentiated and pluripotent

Invasive

Cancer cells share these properties

Isolate human embryonic and primordial germ cell genes which are not expressed in somatic cells.

Test their expression on cancer cells

New cancer genes? New cancer vaccines?

Isolation and identification of oocyte-, embryo- and PGC-specific expressed genes also expressed in cancers

Differential display

DNA sequence Design primers

Confirm stage specific expression (or treatment specific)

Confirm not expressed in range of somatic tissues

Test expression on range of cancer cDNAs

Database analysis Map Identify pseudogenes

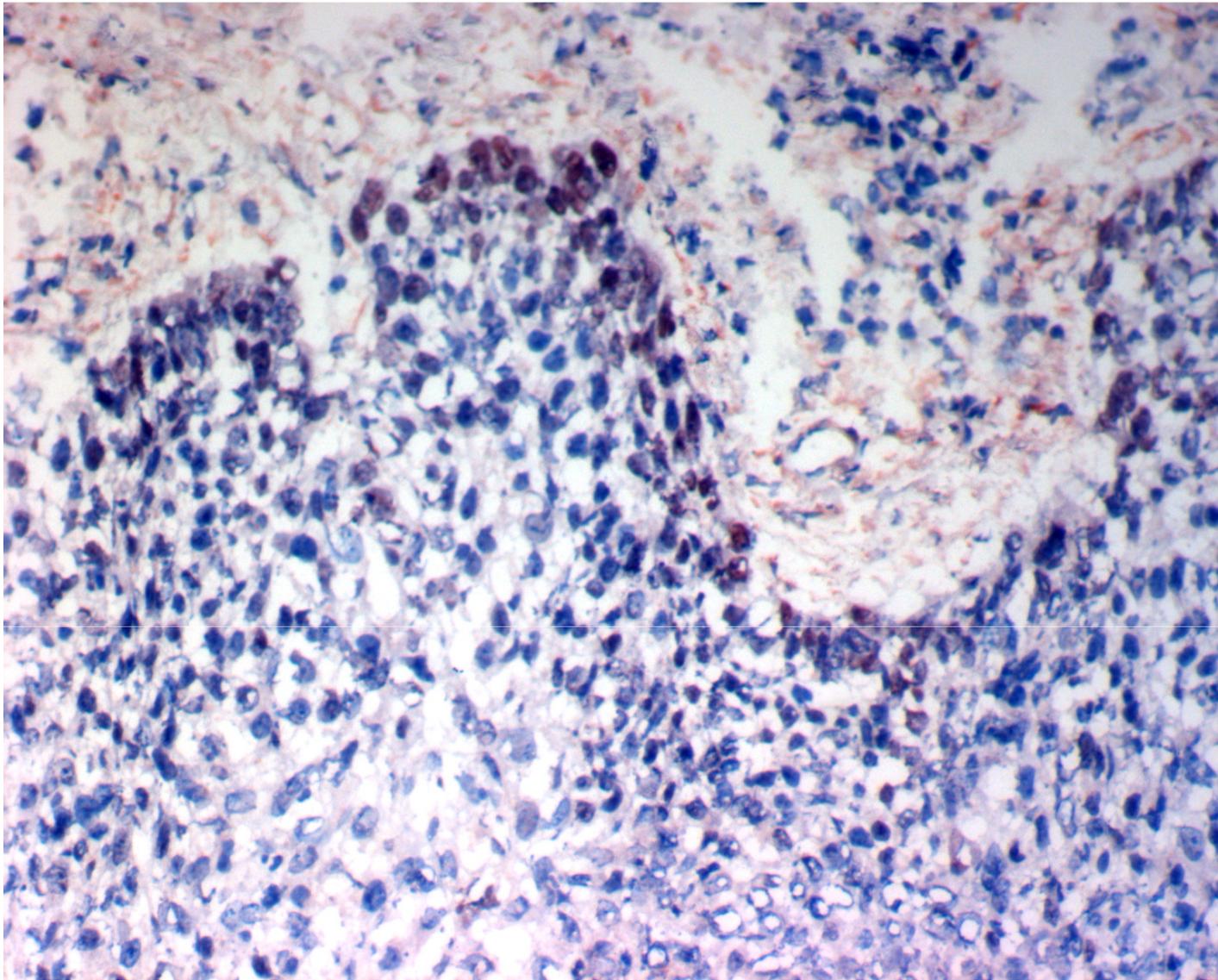
Extend sequences, identify gene exon/intron structure

Northern blot for protein

Immuno-histochemistry on cancer sections

Co-expression with other CT genes

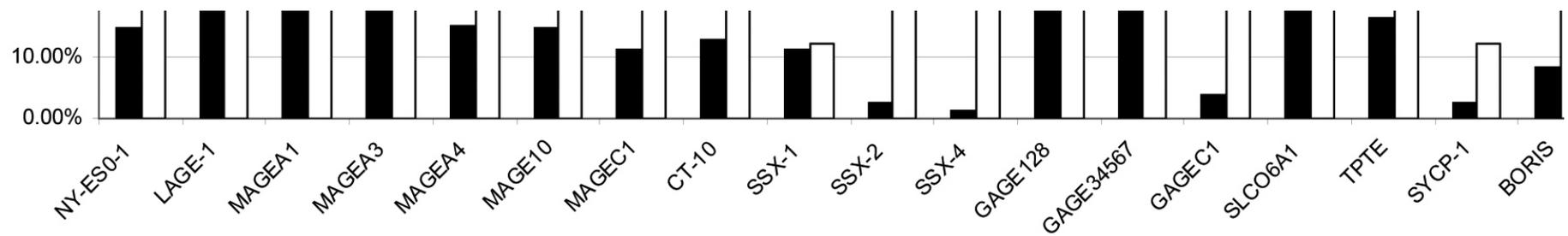
Presence of antibody in patient serum



**ECSA stains basally located subpopulation of cells
in squamous cell NSCLC ...putative cancer stem cells?**

Monk & Holding, 1991

John, Fortunato, Cebon and Monk



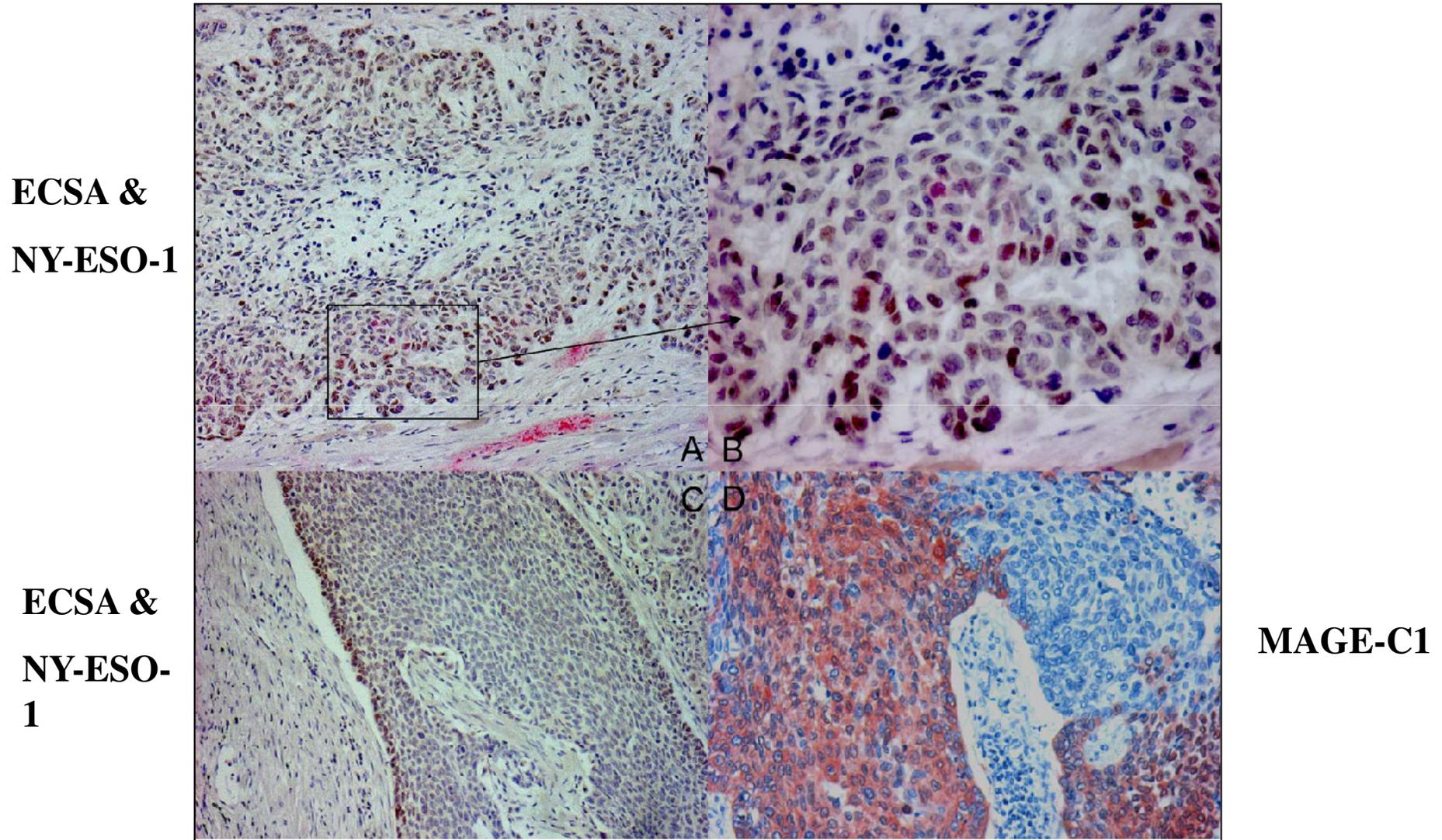
*P<0.001, χ^2 test.

CTA

Enrichment for CTAs in *ECSA* positive tumours

Caballero, Monk, Cebon,

ECSA is co-expressed with CT antigens in stem cells in squamous cell NSCLC



NY-ESO-1 and MAGE-C1 stain many more cells in the tumour

John et al, 2008

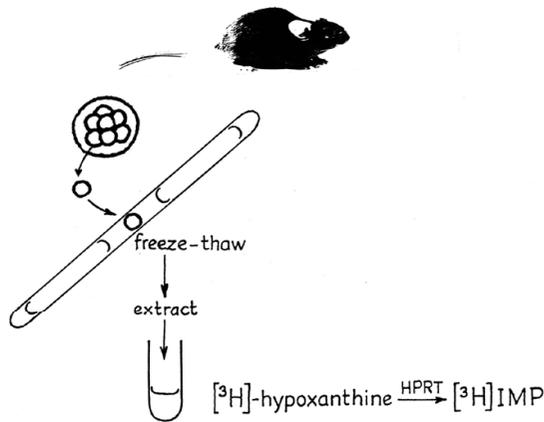
***ECSA* is an immunogenic cancer stem cell gene**

- Minority of cells, basally located, in lung cancer.
- Wide range of cancers - lung, colon, lymphoma, melanoma
- Many CT genes are co-expressed with *ECSA* but they also extend into other derivative cells (*NY-ESO-1*, *MAGE-C1*)
- **Caution** - *In vitro* may not reflect *in vivo*!
Passaged cancer cell lines and ES cells accumulate changes in gene expression e.g., *OCT4* in Clontech passaged cancers & cancer cell lines and *BORIS* in ES cells
(Monk & Holding, 2001; Monk, Hitchens & Hawes, 2008).

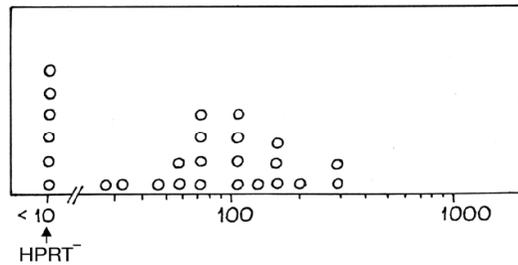
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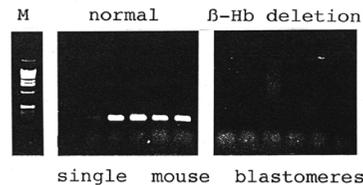
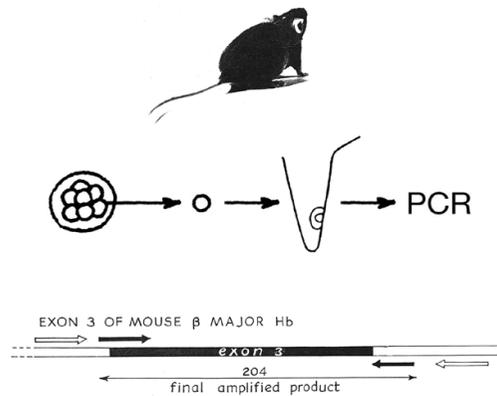


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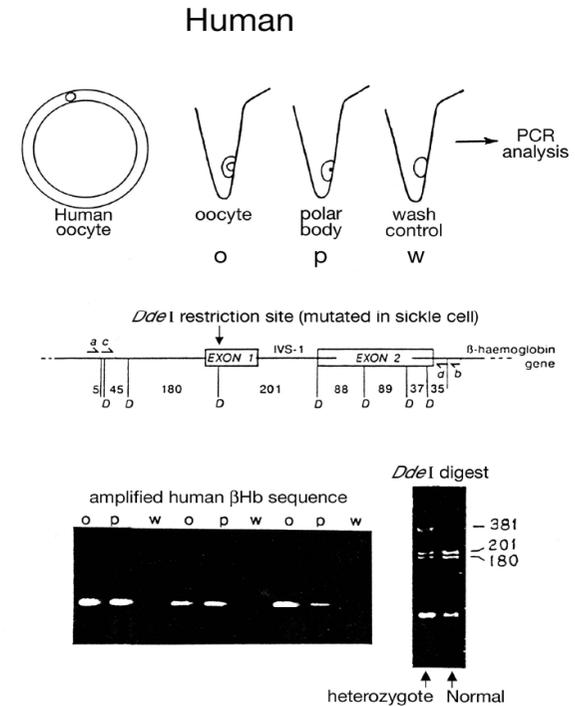
Monk et al, 1987

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Monk & Holding, 1990

ACKNOWLEDGEMENTS

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Kings College Hospital

Virginia Bolton

Hammersmith Hospital

Alan Handyside

Institute Child Health London

Roland Levinsky
Marcus Pembrey
Rob Daniels
Alan Thornhill
Elena Kontogianni
John Huntriss
Tetsuya Goto
James Adjaye
Roberto Lorenzi

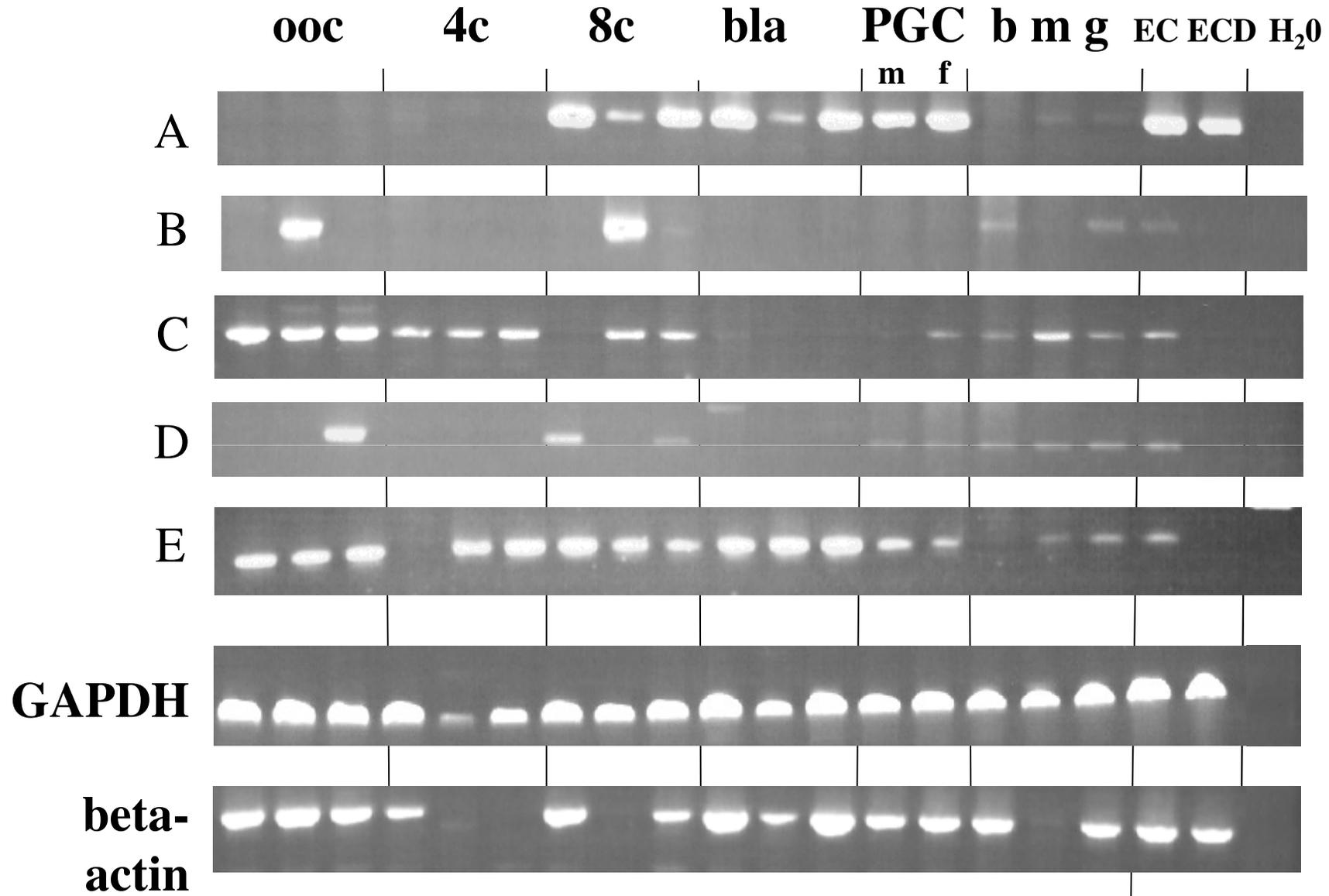
Maurizio Zuccotti
Cathy Holding
Megan Hitchins

Ludwig Institute of Cancer Research

Melbourne

Tom John
Suzanne Svobodova
Jonathan Cebon

Confirmation of specificity of expression of human embryonic genes



Profiles of expression of *ECSA*, *OCT4*, *BORIS* and *NANOG* in embryos and cultured ES cells

- ***ECSA***

Embryo-specific gene, putative cancer stem cell gene, expressed in high proportion of lung, liver and colon cancers.

Monk and Holding, 2001

- ***BORIS***

Brother of regulator of imprinted sites (brother of *CTCF*)

Expressed only in primary spermatocytes in testis (when *CTCF* off)

Candidate gene for deprogramming in spermatogenesis

CT gene

Loukinov et al., 2002

- ***OCT4* and *NANOG***

Standard well-known embryonic genes

Different developmental profiles of expression of *OCT4*, *BORIS*, *ECS-A* in human -

Gonads – testis and ovary

Oocytes

Preimplantation embryos

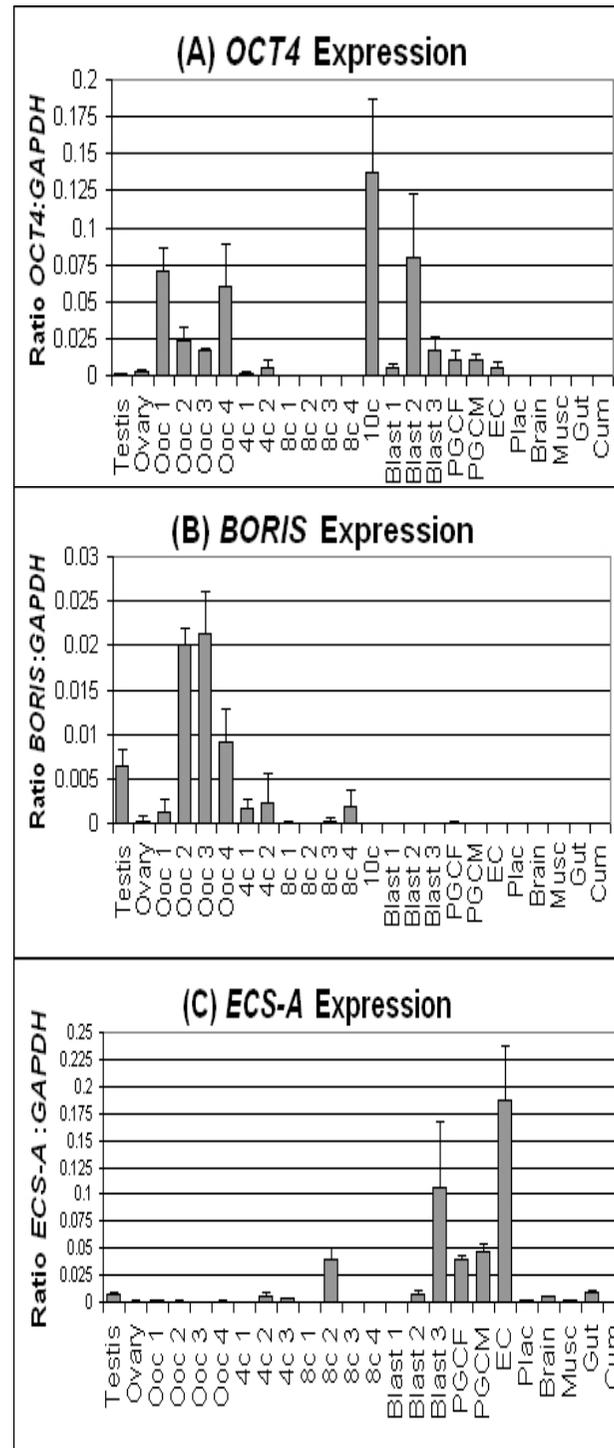
Primordial germ cells

Placenta

Fetal brain muscle and gut

Cumulus cells

Monk& Hitchins, 2006

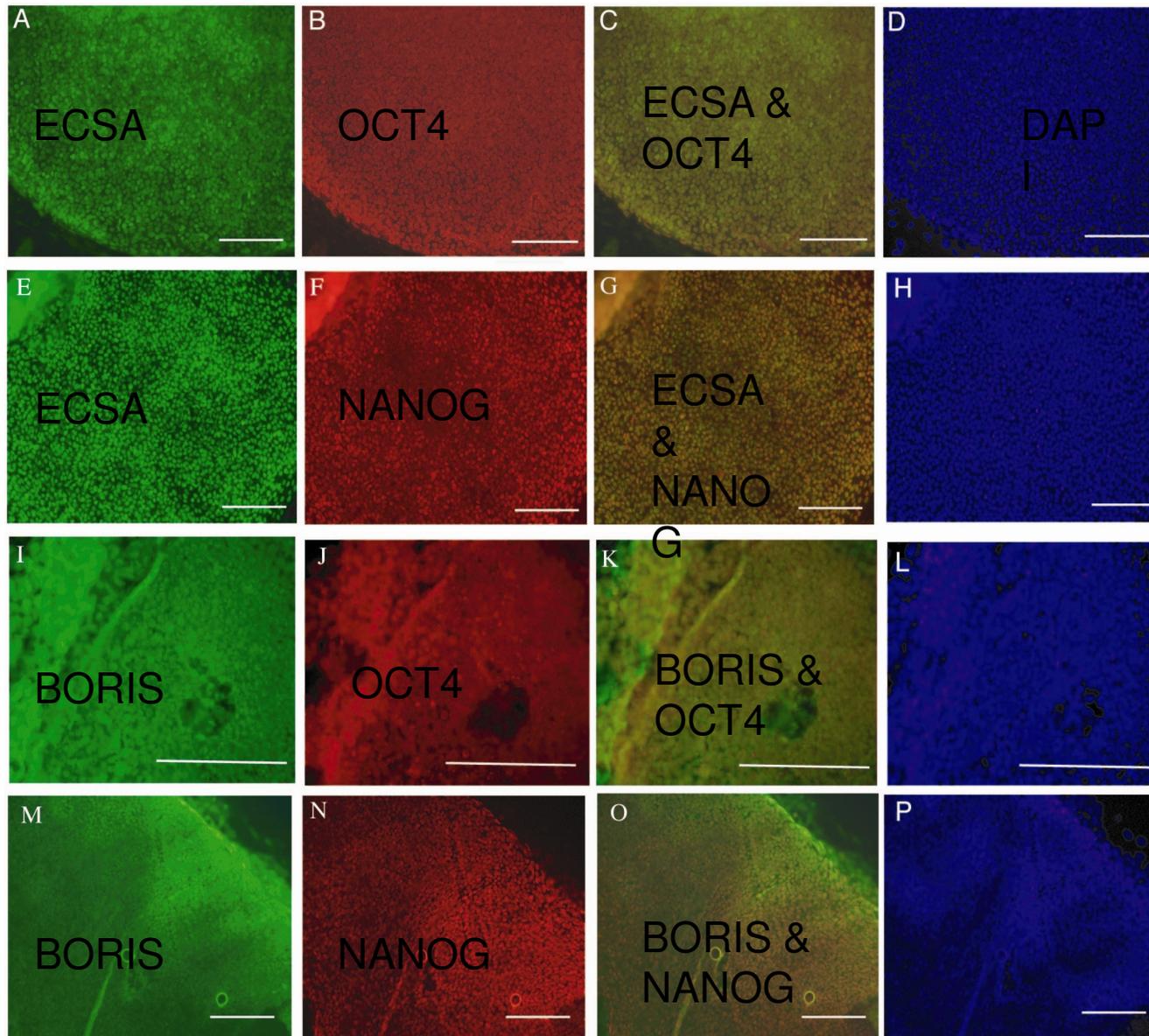


***OCT4*/GAPDH**

***BORIS*/GAPDH**

***ECS-A*/GAPDH**

Co-expression ECSA, BORIS, OCT4 and NANOG in human ES cells



Monk, Hitchins & Hawes, 2008

INHERITED GENETIC DISEASE

For couples at risk: 1 in 4 offspring affected

PRENATAL DIAGNOSIS

Amniocentesis ~16 } weeks pregnancy
Chorionic villus ~11 }

If fetus affected – abortion offered

INHERITED GENETIC DISEASE

PRE-IMPLANTATION DIAGNOSIS

Biopsy of single blastomere } <1 week after
Biopsy of 5 trophectoderm cells } fertilisation

Replace only embryo without defect to mother. Abortion is therefore avoided.

PRE-IMPLANTATION BIOPSY AND DIAGNOSIS

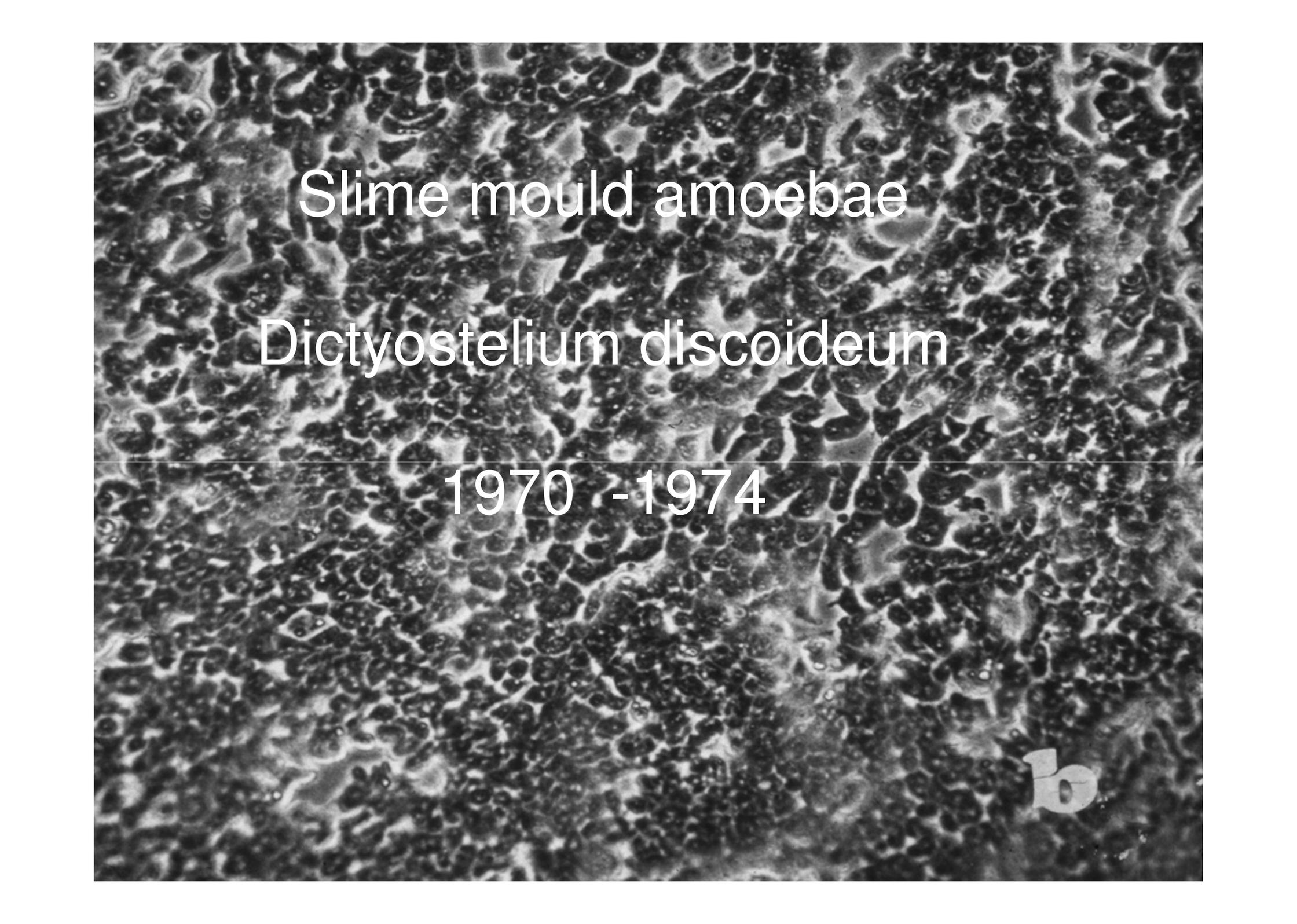
Sexing rabbit embryos by sex chromatin in trophoblast sample.

(Gardner & Edwards, 1968)

Embryo-splitting at 2-cell and 4-cell stage to produce monozygotic twins or quadruplets in cows and sheep.

(Fehilly & Willadsen, 1986)

Diagnosis of sex or genetic disease in a single cell from the 8-cell stage or a few trophoblast cells from the blastocyst.

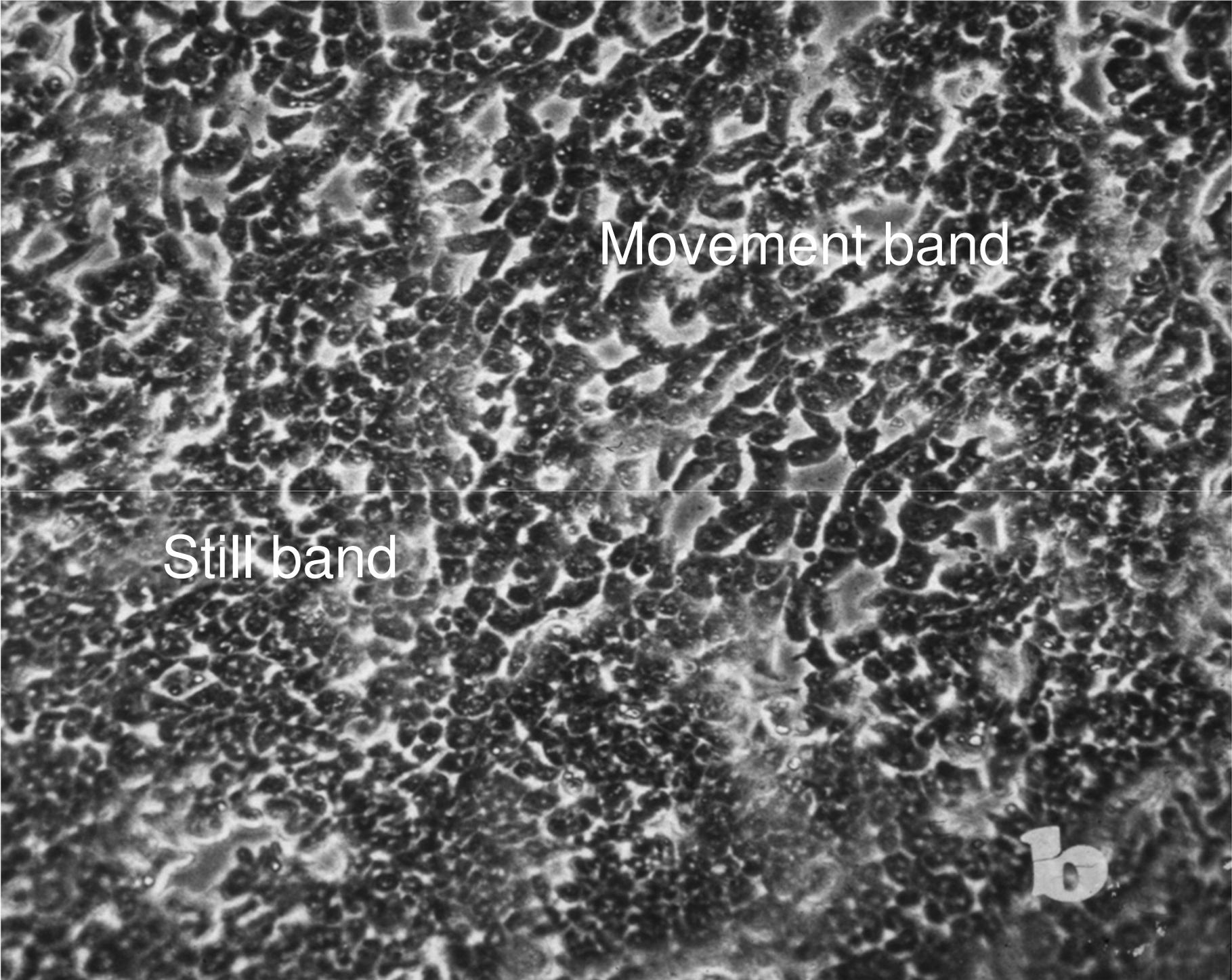
A black and white micrograph showing a dense population of Dictyostelium discoideum cells. The cells are small, roughly circular, and arranged in a confluent layer. They exhibit a granular internal structure and are separated by thin, light-colored channels. The overall appearance is that of a multi-cellular aggregate.

Slime mould amoebae

Dictyostelium discoideum

1970 -1974

b

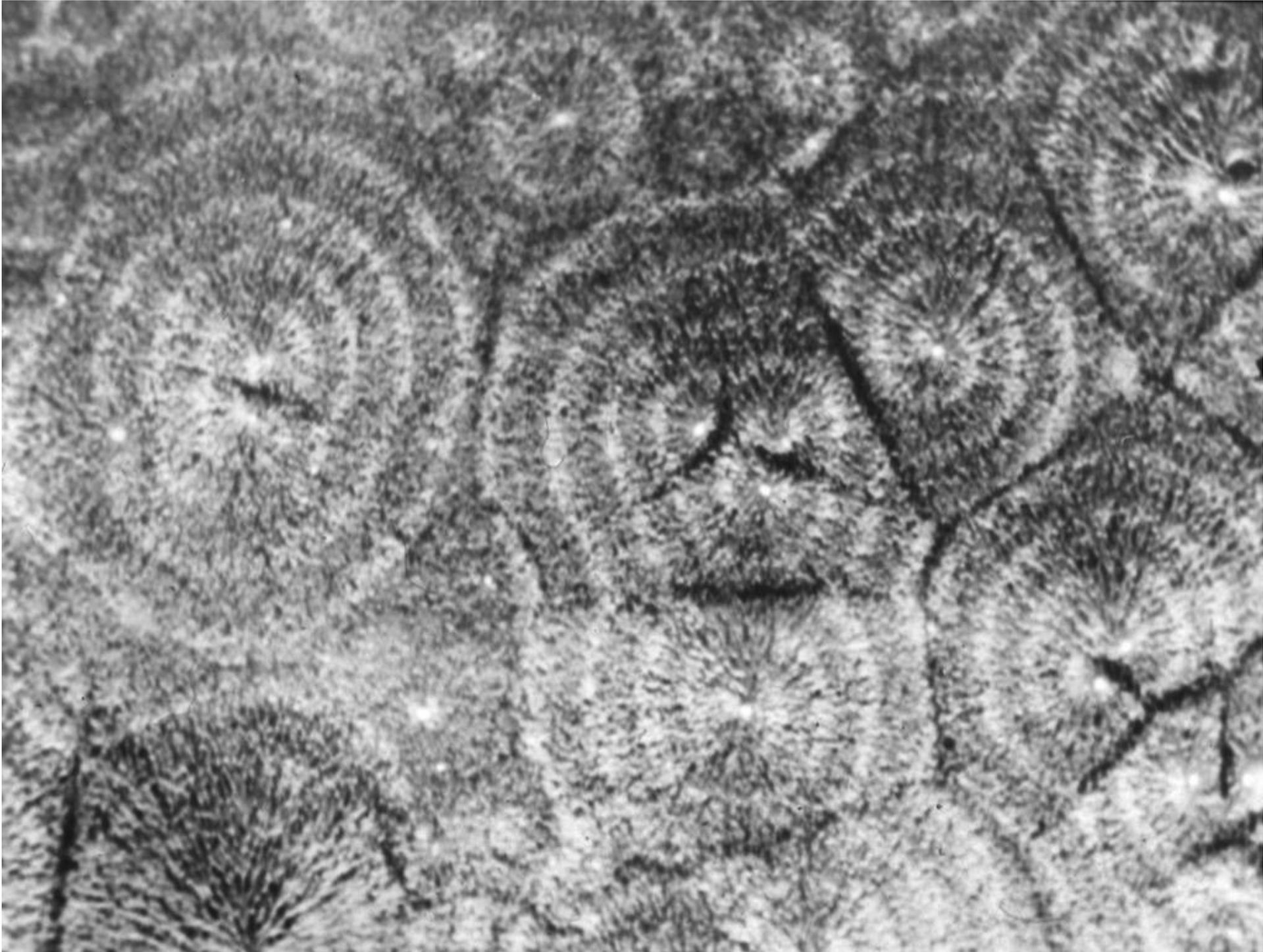


Movement band

The image shows a dense monolayer of cells, likely fibroblasts, in a culture dish. A horizontal band of cells is highlighted, showing a distinct change in cell morphology and density. The cells in the upper band appear more elongated and organized, while the cells in the lower band are more rounded and densely packed. A white horizontal line is drawn across the middle of the image, separating the two bands. The text 'Movement band' is written in white in the upper right quadrant, and 'Still band' is written in white in the lower left quadrant. A small white letter 'b' is located in the bottom right corner of the image.

Still band

b





Differential display identifies human embryo-specific expressed genes

