



Reprogenetics

PGD for infertility

Santiago Munné

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Oxford, UK

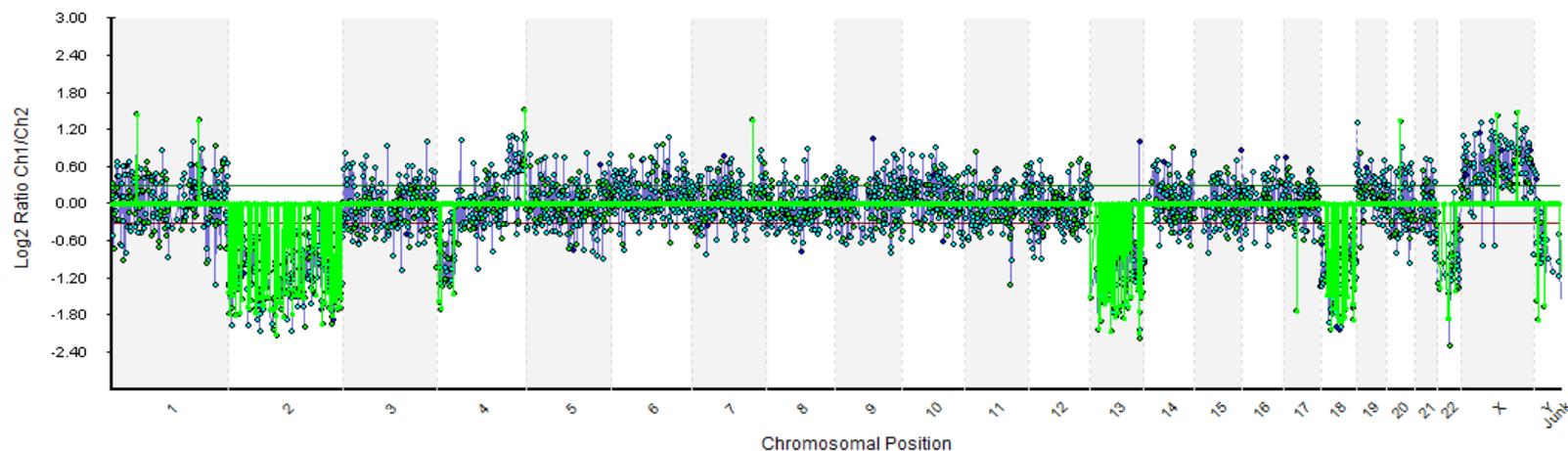
Hamburg, Germany

Asia:

Kobe, Japan

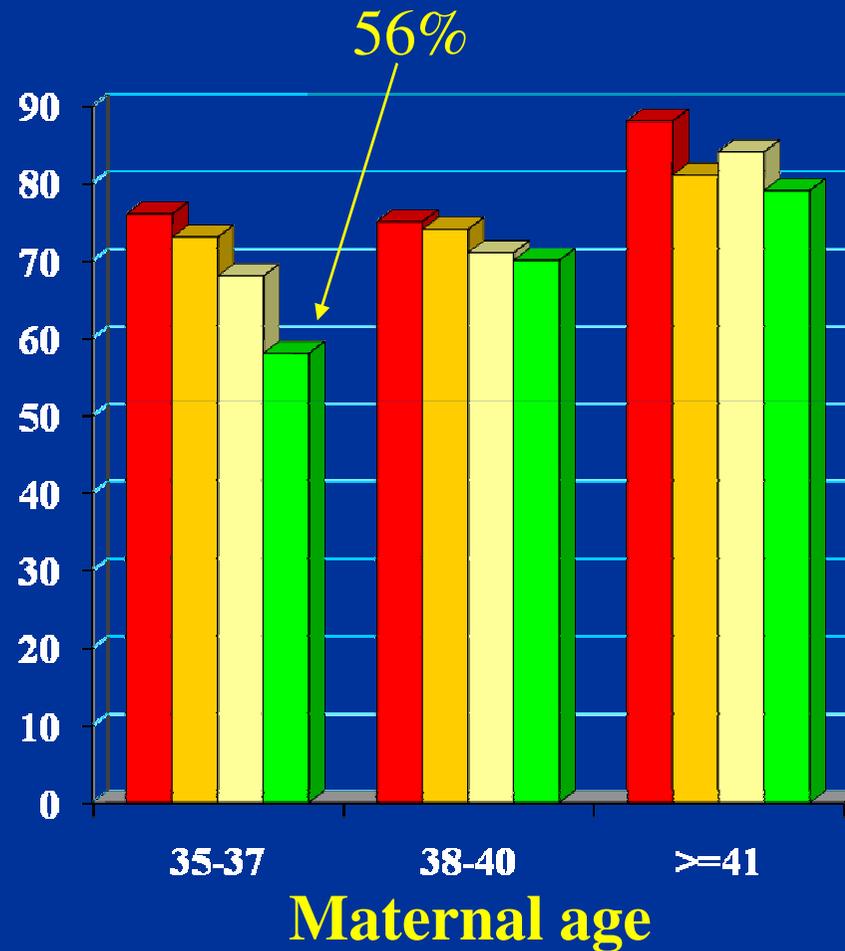
South America:

Lima, Peru



The majority of embryos with 'good' morphology are chromosomally abnormal

% chromosomally abnormal embryos



Morphology:

- arrested
- slow
- dysmorphic
- Good

embryos analyzed: 6054. Morphologically normal embryos: 3751. Source: Munné et al. 2007. Similar results also found by Munne et al 1995, Marquez et al. 2000, Magli et al. 2007.

PGD

Hypothesis

PGD may improve ART outcome in women of advanced maternal age Munné et al. (1993)

Despite large studies indicating the advantages of aneuploidy screening, the notion that PGS for infertility is beneficial is not shared uniformly.

Contradicting PGD results using day 3 biopsy and FISH

Positive effect

Gianaroli et al. 1999
Munne et al 1999
Gianaroli et al 2001a
Gianaroli et al. 2001b
Munne et al. 2003
Gianaroli et al. 2004
Munne et al. 2005
Munne et al 2006
Verlinsky et al. 2005
Colls et al. 2007
Garrisi et al. 2009
Rubio et al. 2009

No effect (small)

Werlin et al. 2003
Jansen et al. 2008
Mersereau et al. 2008
Scholcraft et al. 2009

No effect (Large)

Staessen et al. 2004
Platteau et al. 2005

Negative effect

Mastenbroek et al. 2007
Hardarson et al. 2008

Contradicting PGD results using day 3 biopsy and FISH

Proposed explanations:

- 1) Mosaicism and self-correction
- 2) Sub-optimal PGD and biopsy methods

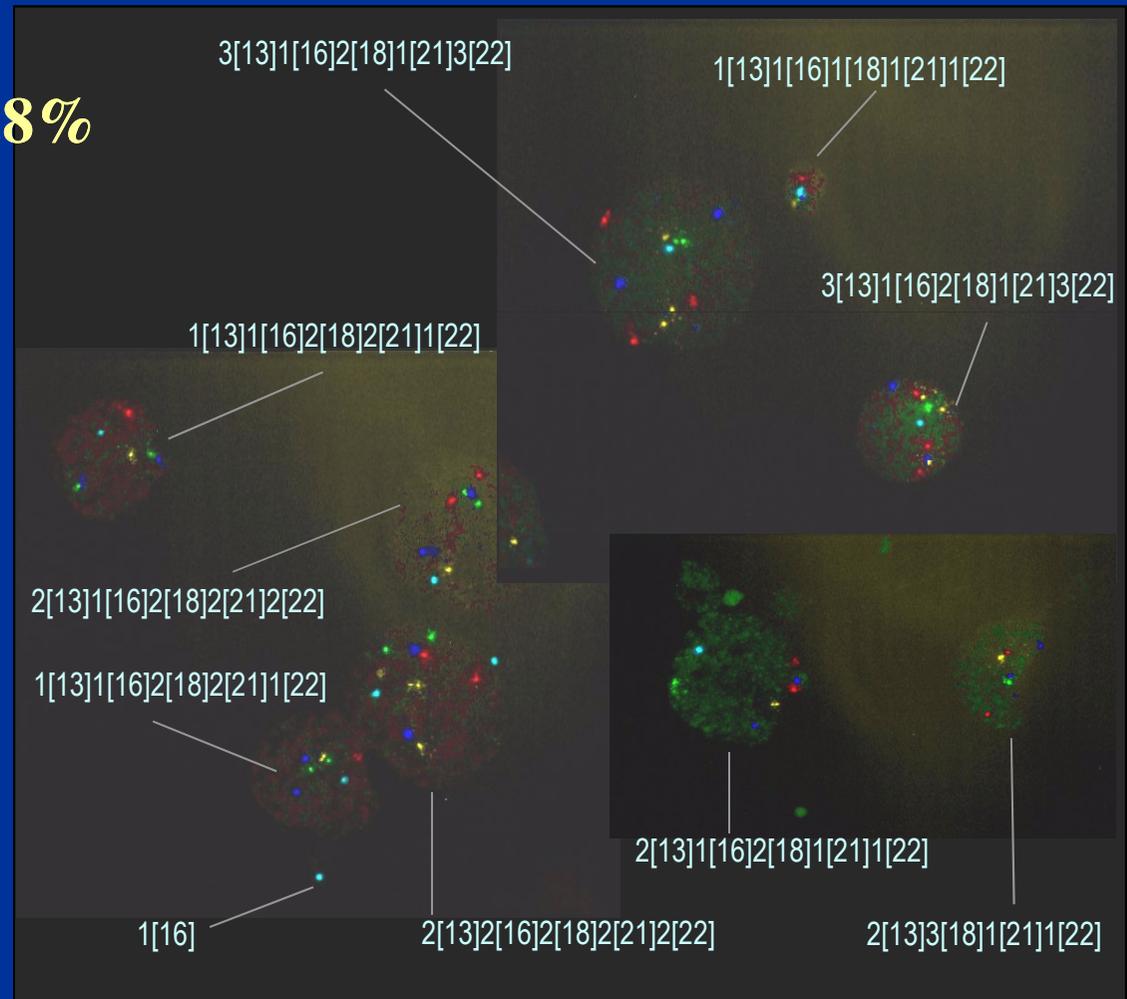
Mosaicism

Mosaicism produces <7% misdiagnosis

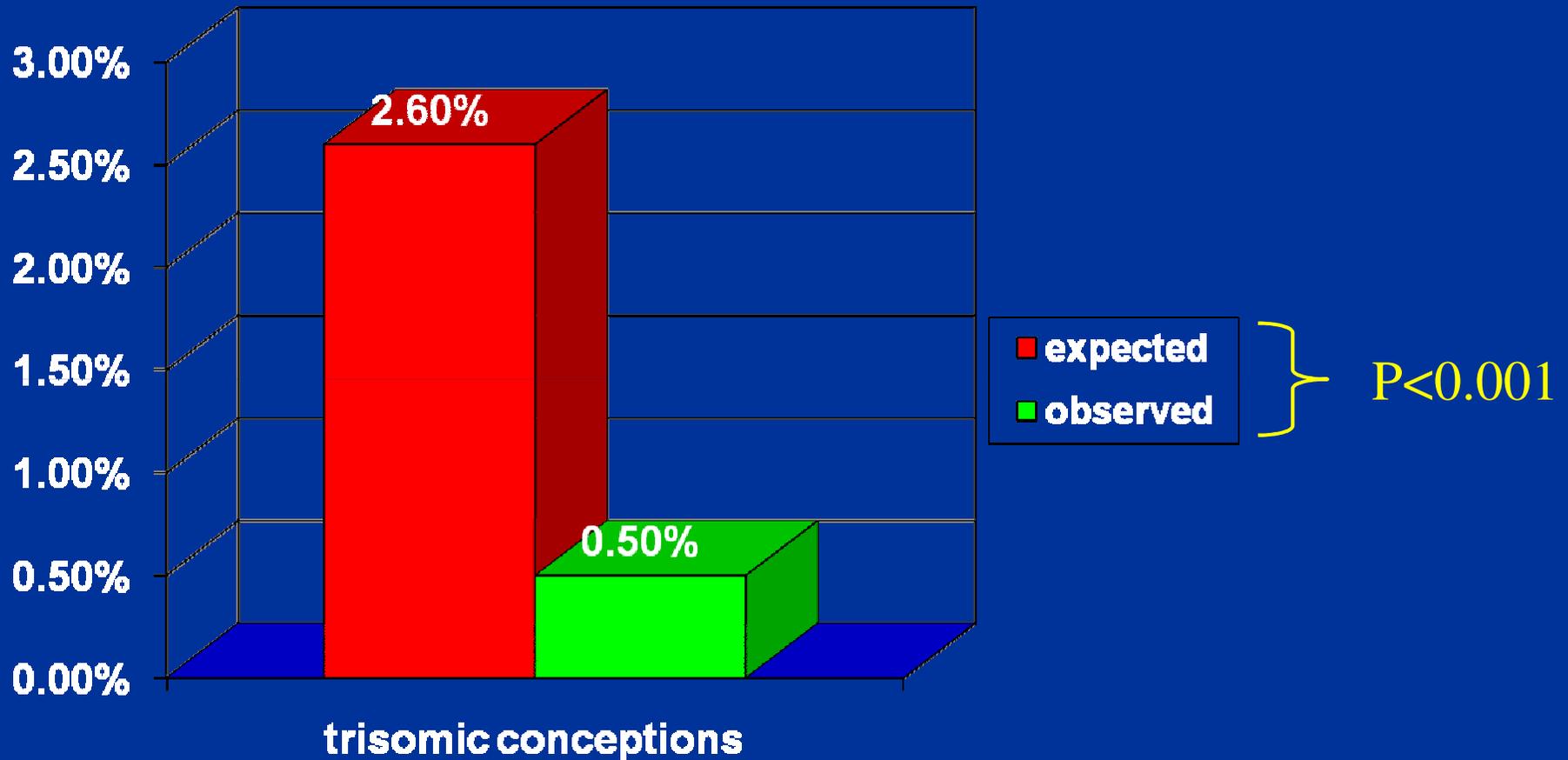
592 embryos found abnormal by PGD were reanalyzed and found to be:

normal	13	} 6.8%
mosaic <49% abnormal	27	
mosaic 50-99% abnormal	124	
mosaic 100% abnormal	297	
homogeneously abnormal	131	

Colls et al. (2007)



Negative predictive value



Aneuploidy rates for chromosomes X,Y,13,18,21. Munne et al. 2006 and Reprogenetics data up to 10/2007. Average age 37, Observed: Based on 2300 pregnancies after PGD, Expected: Eiben et al. 1994. Observed and expected adjusted by maternal ages

self-correction myth

Trisomy correction is rare: UPD evidence

UNIPARENTAL DISOMY:

Trisomy rescue: creates a zygote with 2 chromosomes from one parent and none from the other.

EXAMPLE TRISOMY 15:

Trisomy 15 in cleavage stage embryos: 1.874% a

UPD-15 in newborns: 0.001% b

Estimated correction of trisomy 15 to UDP: 1/3 c

Trisomy 15 day 3 embryos that self-corrected: $a \times b \times c = 0.56\%$

a: Munne et al. (2004), b: From: OMIM, c: 1/3 of corrections will produce UPD

Fetus seldom self correct: it's the placenta that becomes abnormal



ELSEVIER

DEVELOPMENTAL
BIOLOGY

Human cytotrophoblasts acquire aneuploidies as they differentiate to an
invasive phenotype

Jingly F. Weier^{a,b}, Heinz-Ulrich G. Weier^b, Christine J. Jung^c, Matthew Gormley^a, Yan Zhou^c,
Lisa W. Chu^b, Olga Genbacev^c, Alexi A. Wright^c, Susan J. Fisher^{a,c,d,*}

2005, Dev. Biol. 279, 420-432

This work questions the assumption that placental confined mosaicism is the result of fetal self-correction. At the contrary, it suggests that normal fetuses may develop abnormal placenta.

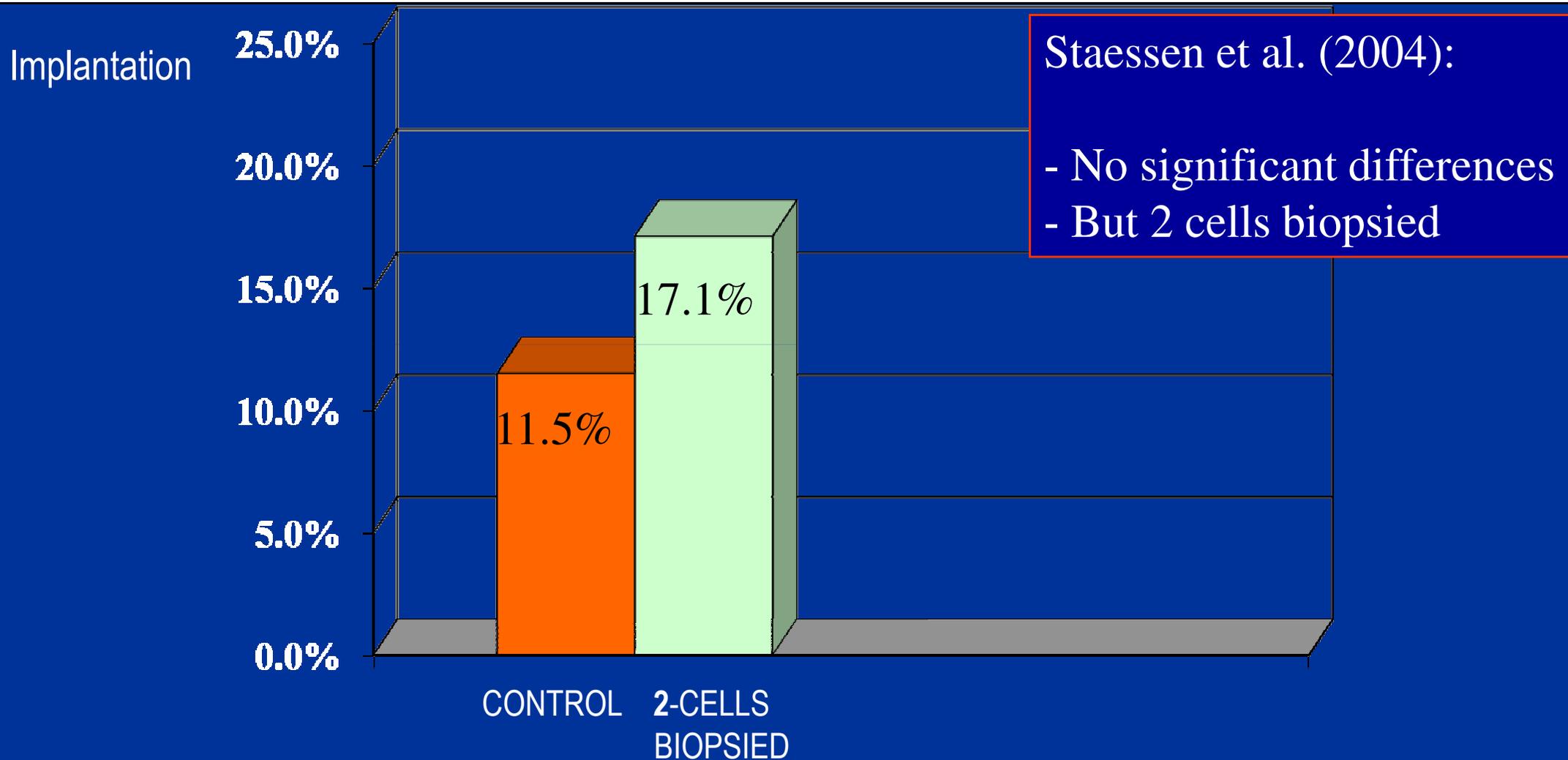
**Sub-optimizal methods
(FISH studies, day 3 biopsy)**

Optimal PGS methods

	Optimal PGS	Questionable PGS
Biopsy media	with aminoacids	simple media
Biopsy time / embryo	1 min	> 5 min
# cells biopsied	one	two
Fixation method	Carnoy's	Tween 20
# chromosomes tested	≥ 8	≤ 6
# analysts / case	2	1
Use of NRR*	yes	no
Large experience	yes	no
Error rate	<10%	10-50%
Number of zygotes	>5	≤ 5

**NRR: No result rescue, or re-testing of dubious chromosome with different probes.*

PGD for AMA: randomized studies



Staessen et al (2004)

Two cell biopsy is detrimental

Hum. Reprod. Advance Access published September 21, 2009

human
reproduction

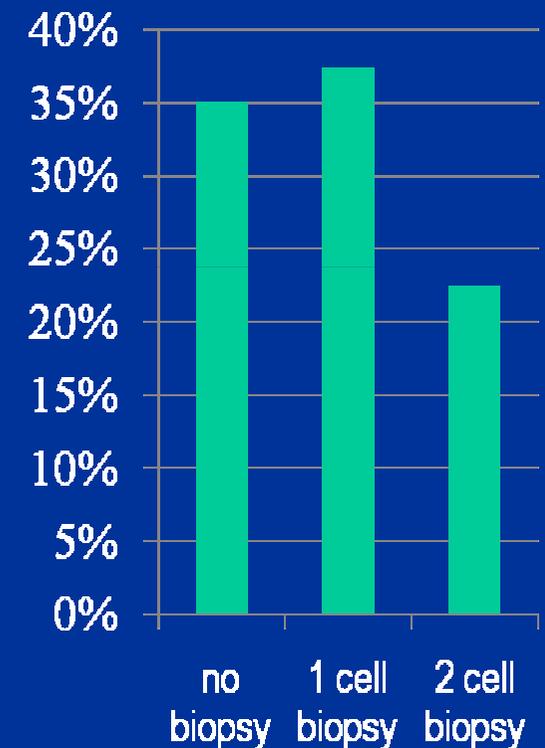
ORIGINAL ARTICLE *Embryology*

Impact of cleavage-stage embryo biopsy in view of PGD on human blastocyst implantation: a prospective cohort of single embryo transfers

A. De Vos^{1,4}, C. Staessen², M. De Rycke², W. Verpoest¹,
P. Haentjens³, P. Devroey¹, I. Liebaers², and H. Van de Velde¹

“The data presented here clearly indicates that two cell biopsy significantly impacts clinical outcome. Our previous report providing no arguments in favour of PGS (Staessen et al., 2004) was criticised by others arguing that PGS might have been beneficial if only one cell had been removed (Cohen et al., 2007). In respect to the present findings, this criticism seems justified”.

ongoing pregnancy



$P < 0.001$

PGD for AMA: randomized studies

Mastenbroek et al. (2007)

- 1) 20% of cycles undiagnosed (literature: 1-3% of embryos *)
- 2) 59% implantation reduction due to biopsy:

implantation

Control	14.7%	} 59% reduction
Biopsied, no PGD	6.0%	
Biopsied and PGD	16.8%	

- 3) Average number of embryos analyzed was only 5
- 4) Chromosomes 15 and 22 (21% abnormalities) not analyzed

* 1% Gianaroli et al. (2004), 3.1% Colls et al. (2007)

Number of chromosomes analyzed

At least 9 chromosomes should be tested:

# chromosomes analyzed	% abnormal fetuses detectable
5	28%
6	47%
9	70%
12	80%
24	100%

Minimum number of embryos to do PGD

# 2pn's	Av. Age	Pregnancies
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1-5	38	22%
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1-5	38	22%
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1-5	38	22%
-----	----	-----

6-9	38	36%
-----	----	-----

6-9	38	36%
-----	----	-----

6-9	38	36%
-----	----	-----

10-13	38	40%
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10-13	38	40%
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10-13	38	40%
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≥ 14	38	48%
------	----	-----

≥ 14	38	48%
------	----	-----

≥ 14	38	48%
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Data: Last 300 cases, to 7/2007, Saint Barnabas Medical center, unpublished

Error rate should be <10%

Analysis of remaining cells of embryos previously analyzed by PGD:

study	technique	error rate
Baart et al 2004	FISH	50.0%
Li et al. 2005	FISH	40.0%
Gleicher et al. 2009	FISH	15-20%
Munne et al. 2002	FISH-9	7.2%
Colls et al., 2007	FISH-9	4.7%
Magli et al. 2007	FISH-9	3.7%
Munne et al. 2010	array CGH	1.8%

PGD for Recurrent Pregnancy Loss (RPL)

All controlled PGD studies on idiopathic RPL show a decrease in miscarriages

Idiopathic RPL :

Werlin L, et al. (2003) Preimplantation genetic diagnosis (PGD) as both a therapeutic and diagnostic tool in assisted reproductive technology. *Fertil Steril*, 80:467

Munné et al. (2005) Preimplantation genetic diagnosis reduces pregnancy loss in women 35 and older with a history of recurrent miscarriages. *Fertil Steril* 84:331

Garrisi et al. (2009) Effect of infertility, maternal age, and number of previous miscarriages on the outcome of preimplantation genetic diagnosis for idiopathic recurrent pregnancy loss. *Fertil. Steril* 92: 288

Rubio et al. (in press) Prognosis factors for Preimplantation Genetic Screening in repeated pregnancy loss. *Reprod Biomed Online*, in press

Reduction in miscarriages in RPL after PGD

PGD results according to previous number of miscarriages

# previous miscarriages	cycles	% loss expected	% loss after PGD	
2	90	29%	19%	N.S.
>2	190	38%	9%	p<0.00.1

Reduction in miscarriages in RPL after PGD

PGD results according to age when previous number of miscarriages is 3-5

maternal age	cycles	% loss expected	% loss after PGD	
<35	78	26%	10%	p<0.025
≥35	202	39%	13%	p<0.001

Garrisi et al. (2009), Reprogenetics, unpublished results

Reduction in miscarriages in RPL after PGD

PGD results according to fertility

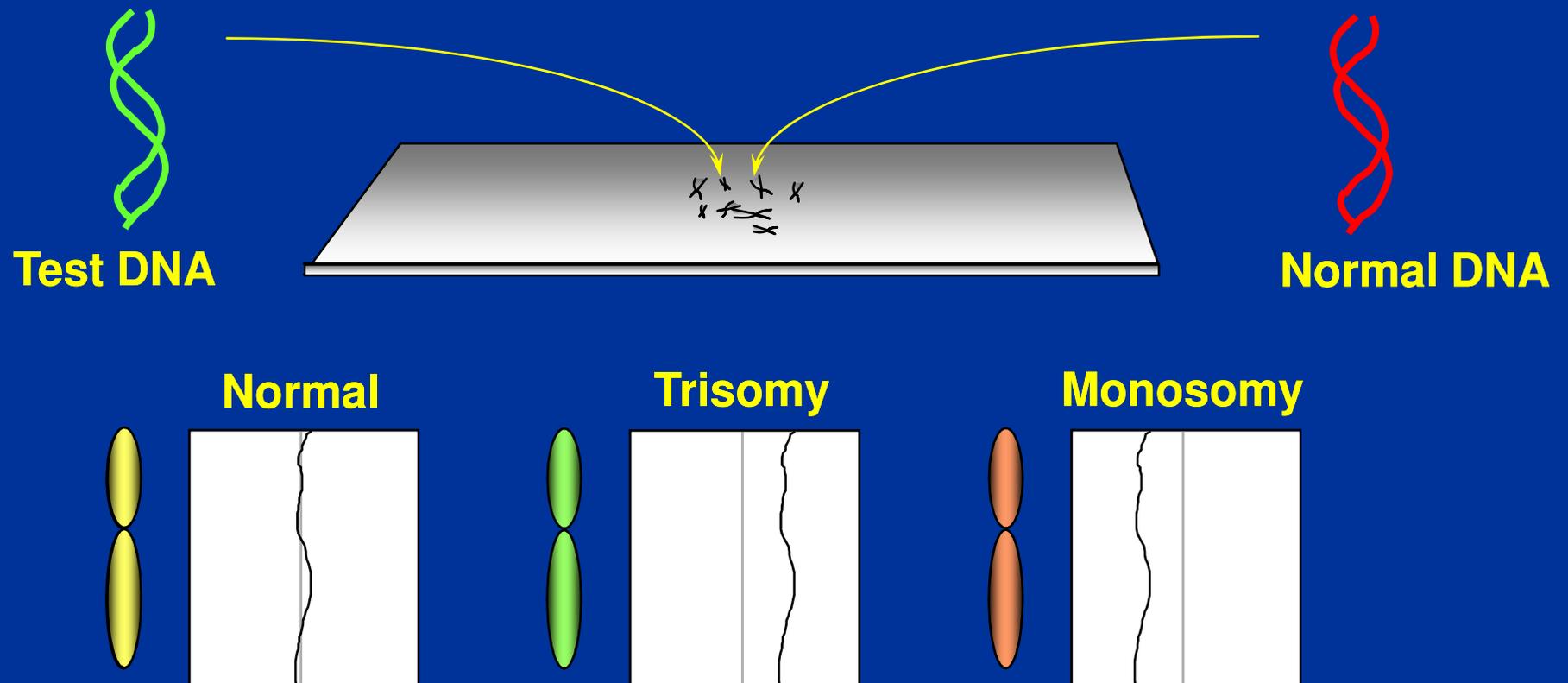
method	cycles	% loss expected	% loss after PGD	p	% to term
IVF	115	35%	14%	p<0.01	34%
natural	124	41%	15%	p<0.005	37%

Average maternal age: 37.5
Garrisi et al. (2009)

New approach to PGD:

- **24 chromosome analysis by arrays**
- **Blastocyst biopsy and vitrification**

Comparative Genome Hybridization (CGH)



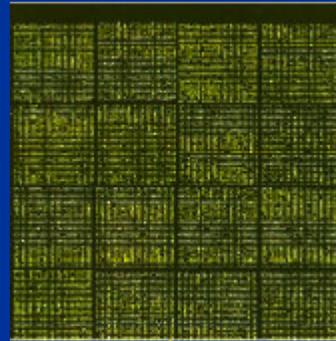
Kallioniemi et al. (1992), applied to single cells by Wells et al. (1999)

Array CGH

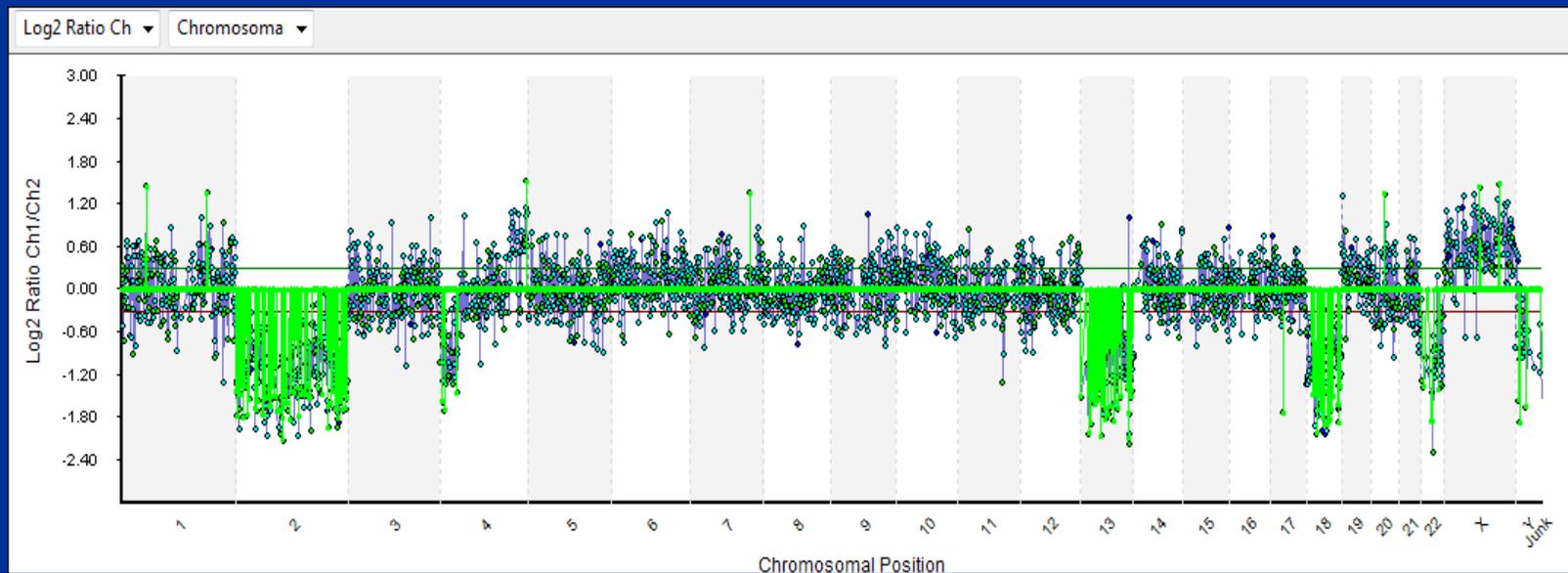
Test
DNA



Normal
DNA

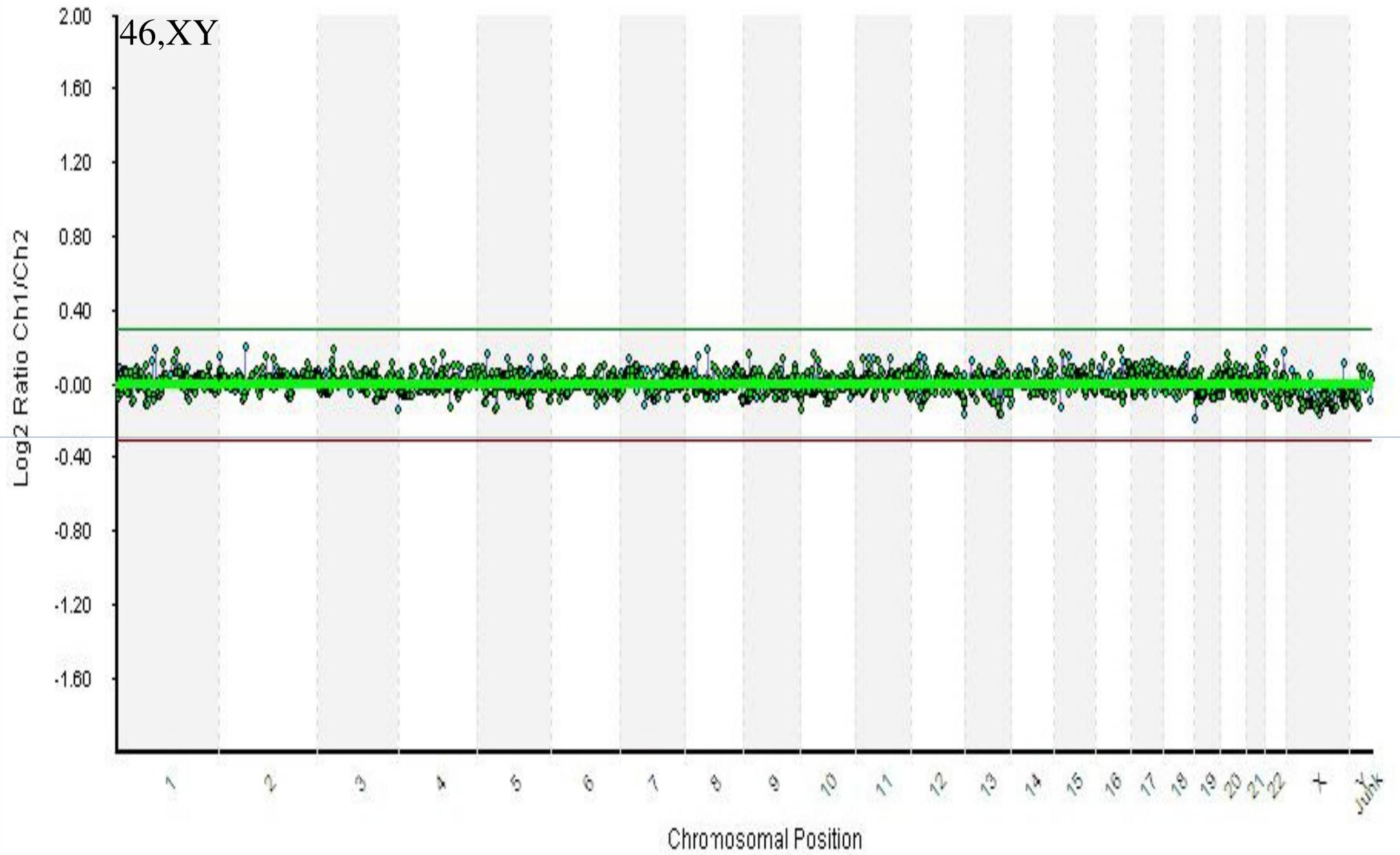


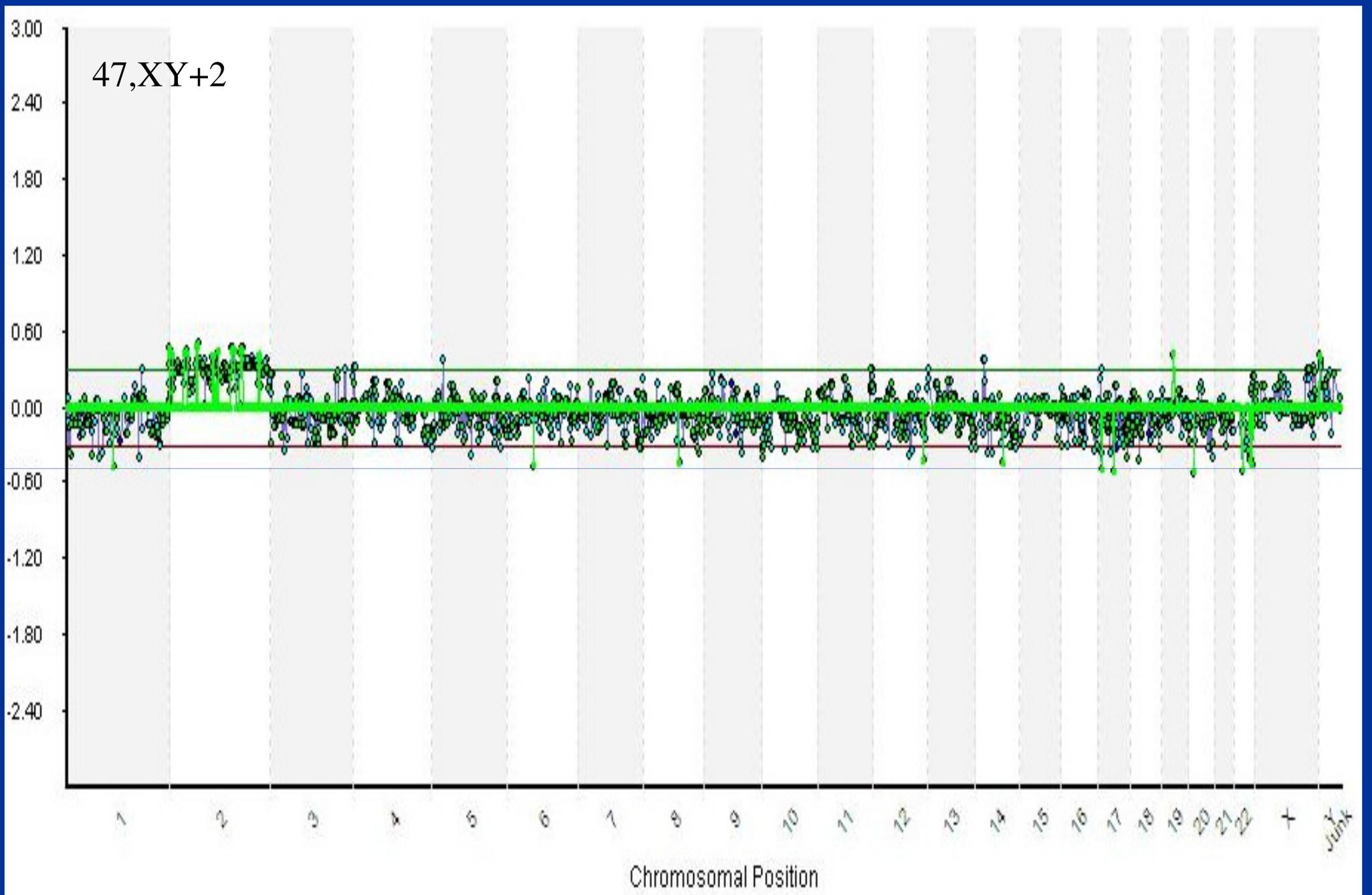
2700 probes
Same band resolution as karyotype



aCGH advantages

- All **24 chromosome** type of aneuploidies detected
- Results in **24** hours; allows for PB or day 3 biopsy
- Parental DNA **not** required: ad hoc decisions possible
- Used in **>15,000** patients with mental retardation





aCGH validation: no results

Embryos undiagnosed:

biopsy on day 3: 2% (16/724)

biopsy on day 5: \approx 0% (0/64)

Gutierrez-Mateo et al. (in press)

aCGH validation: error rate

- Validation method 1: single cells from cell lines analyzed*

Error rate in euploid cell lines: 0/9

Error rate in aneuploid cell lines: 0/42

- Validation method 2: Reanalysis of the rest of the embryo by FISH with 19 chromosomes probes**

Error rate from day 3 biopsies: 1.8% (1/56)

* Mamas et al (submitted), ** Gutierrez et al. (in press)

Day 3 biopsy, day 5 transfer and array CGH

Cycles performed: 219

Maternal age (av.) 37.5

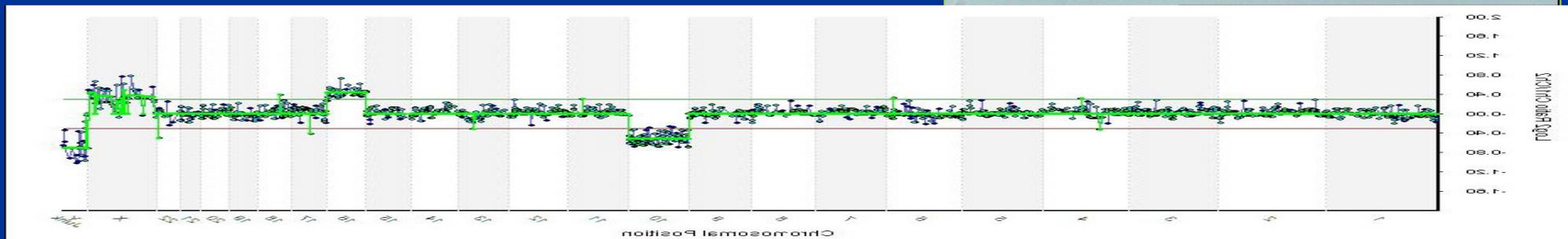
	<i>Pregnancy Rate</i>		<i>Ongoing Pregnancy Rate</i>	
	<u>Per Cycle</u>	<u>Per ET</u>	<u>Per Cycle</u>	<u>Per ET</u>
Control	37%	37%	31%	31%
PGD	46%	60%	42%	55%
	NS	< 0.001	NS	< 0.001

* Expected from each center SART data, controlled by age
Data from 24 centers. Munné et al. (2010) ESHRE, and unpublished data

array CGH on blastocyst biopsies: Why?

Advantages:

- 1) More DNA: More robust diagnosis
- 2) Eliminates some mosaic embryos
- 3) Reduces error rate
- 4) Reduced impact of embryo biopsy
- 5) Less embryos to process
- 6) Facilitates single embryo transfer
- 7) Uterine environment optimized after thaw



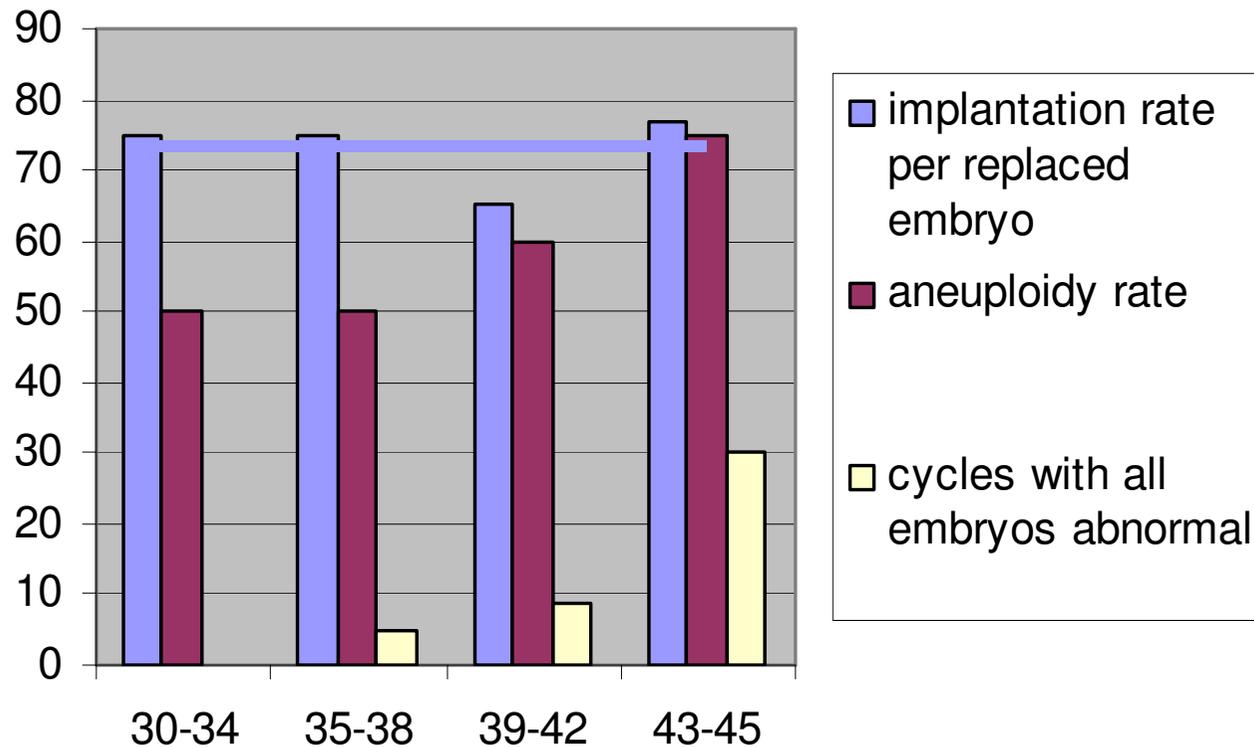
CGH on blastocyst biopsies: clinical results

	Cycles	Mat. age	Prev. failed	embryos replaced cycles	implant. (+ sac)	ongoing preg.
CGH :	45	37.7	2.4	2.0	67%	79%
control :	113	37.1	1.2	2.7	28%	60%

p=0.0003

Schoolcraft et al. (in press)

CGH on blastocyst biopsies: Implantation is independent of age



Patients <43 who are eligible for blastocyst transfer have a >95% change of having normal embryos available for transfer

Results of aCGH in PB and day 3 biopsies

ESHRE study: PB data

Average age	40
Cycles	42
Embryo replaced	n/a
Implantation rate	n/a
Pregnancy rate	19%
Error rate	11%

Reprogenetics: data day 3 biopsy

Average age	40
Cycles	107
Av. Embryos replaced	1.0
Implantation rate	31%
Pregnancy rate	26%
Error rate	2% *

** Gutierrez-Mateo et al., Fertil Steril, accepted*

**SNP and CNP arrays:
For diagnosis of aneuploidy**

aCGH vs. SNP arrays: Genome coverage

	# of probes	probe size	genome covered
aCGH	4,000	x 150,000 kb	= 600.0 Mb (25%)
SNPs	300,000	x 50 kb	= 1.5 Mb (>0.1%)

SNP arrays: Treff ' team validation

Comparison of implantation rates for those cases with mixed transfers:

- 33 transfers with a mix of SNP array normal and abnormal embryos
- 17 ongoing / delivered pregnancies
- 86 total embryos transferred:
 - 42 normal
 - 44 abnormal

	Embryo delivers	Failed Ongoing development
PGD normal	18	24
PGD abnormal	0	44

P<0.01

SNP arrays: Treff ' team

Blastocyst biopsy, Cryopreservation, SNP array, transfer in thawed cycle

- N=368
- Two centers: RMANJ, CCRM
- Age = 38.2 years
- Number of prior attempts = 2.4
- Blastocysts transferred = 1.6

- Pregnancy rates:
 - clinical: 80%
 - ongoing past 1st trimester: 76%
 - sustained implantation rate: 60%
 - rates equivalent at the two centers (differ by < 1%)

CONCLUSIONS

Conclusions: chromosome abnormalities

- Age and morphology are poor indicators of aneuploidy
- Less than 50% of good morphology day 3 embryos and less than 60% of blastocysts are normal in patients >35
- Selecting for euploid embryos should improve ART outcome

Conclusions: FISH studies

Studies with improved results differ from those that show no improvement in that:

- 1) Reduce biopsy damage (1 cell, experience, blast?)
- 2) Low error rate (fixation, NRR, 2 analyzers)
- 3) Analyze 16,15,21,22 chromosomes + ≥ 4 more
- 4) Extensive experience
- 5) Appropriate population (≥ 5 embryos, ≥ 35 y. old)

Conclusions: array CGH

- Blastocyst biopsy + CGH, SNP arrays + vitrification shows very high implantation rates (72%, av. Age 38).
- Array CGH and day 5 biopsy will produce same results
- Array CGH and day 3 biopsy improves results when normal embryos are available.
- Additional vitrification step may still be advantageous



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