

Biology of Reproduction Unit

## Biology of Oocyte Maturation

Carlos E. Plancha<sup>1,2</sup>

<sup>1</sup> *Unidade de Biologia da Reprodução, Inst. Histologia e Biologia do Desenvolvimento, Faculdade de Medicina de Lisboa, Portugal*  
<sup>2</sup> *CEMEARE – Centro Médico de Assistência à Reprodução, Lisboa, Portugal*

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## Oogenesis

**OOCYTE GROWTH**

Primary follicle, Secondary follicle, Antral follicle, Corpus luteum

**OOCYTE MATURATION**

M<sub>I</sub>, M<sub>II</sub>, GV

**OVULATION**

◆ **Growth Phase:**

- Oocyte diameter increases
- Organelle redistribution
- High transcriptional and translational activity
- Accumulation of RNA / proteins
- Incompetent → Competent ooc.

◆ **Oocyte Maturation:**

Complex series of nuclear and cytoplasmic events with resumption of the 1<sup>st</sup> meiotic division and **arrest at M<sub>II</sub>** shortly before ovulation

**Ovulation**

**FERTILIZATION**

Resumption of meiosis → PN formation  
Embryo development

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## Oogenesis *in vivo* (including oocyte maturation) takes place inside a morfo-functional unit: **The Ovarian Follicle**

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Factors involved in oogenesis and folliculogenesis

Igf-1,2,3

GDF-9

FSH, LH

Cellular interactions

Perifollicular matrix

Laminin

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Somatic-germinal cellular interactions

TZP (Trans-Zonal Process)

Adapted from Baker, Oogenesis and ovulation

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What are TZPs ?

Granulosa cell extensions that transverse the zona pellucida and terminate at the oocyte cell surface (Herlig and Adams, 1967; Anderson and Albertini, 1976)

Microfilaments: - F-actin → TZP-Act

Microtubules: - α+β tubulin → TZP-MT

TZPs have a heterogeneous cytoskeletal content

Gap junctions

Cumulus cells

TZPs

Oocyte surface

N Dekel et al. (1978) Gam Res 1:47.

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## The oocyte-granulosa cell interface

- 1 Gap/adherens jxns
- 2 Oocyte exocytosis
- 3 GC exocytosis

Anderson and Albertini, 1976

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## Antral ovarian follicle

In antral follicles, meiotic arrest is maintained by granulosa cell signaling

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## FSH modulate TZP phenotype in GCs

1. FSH-like priming reduces cumulus TZP-MT density

Genotype	Unprimed	Primed
FSH $\beta$ +/-	10.52 $\pm$ 0.75 (83) <sup>a</sup>	9.81 $\pm$ 0.61 (79) <sup>a</sup>
FSH $\beta$ -/-	24.3 $\pm$ 1.45 (74) <sup>b</sup>	8.94 $\pm$ 0.57 (83) <sup>a</sup>

CMH Combelles et al. (2004) Hormonal control of somatic cell oocyte interactions during ovarian follicle development. Mol Repr Devel 69: 347.

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## FSH modulate TZP phenotype in GCs

### 2. Absence of FSH enhances cumulus TZP-MT density

CMH Combelles et al. (2004) Hormonal control of somatic cell oocyte interactions during ovarian follicle development. *Mol Repr Devel* 69: 347.

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## Meiotic cell cycle progression

### Ovulation

Prophase I

Maturation

Ovulation

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## Lessons from the hamster model

LH (in vivo)

and/or

FSH

Step I

Step II

Step III

CE Plancha, DF Albertini (1994) Hormonal regulation of meiotic maturation in the hamster oocyte involves a cytoskeleton mediated process. *Biol Reprod* 51:952-964

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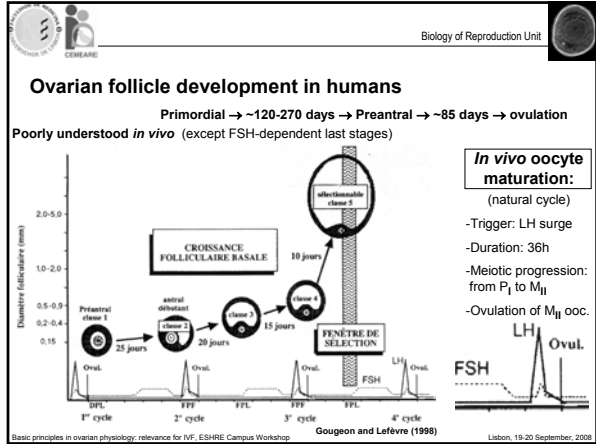
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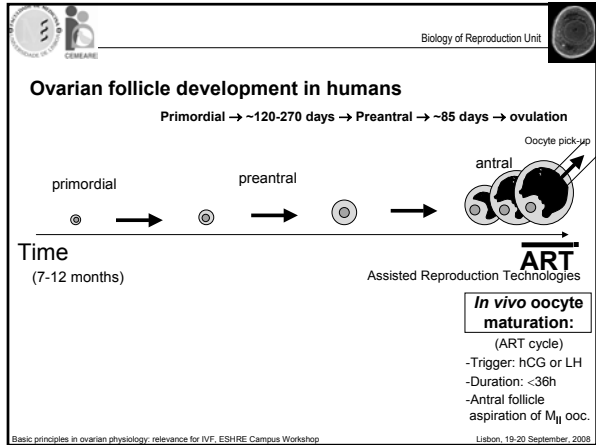
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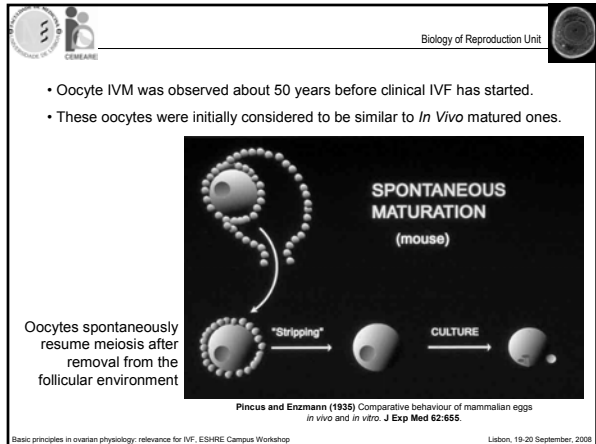
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Mouse

However,

Efficiency of meiotic cell cycle progression is lower in IVM oocytes

Maturation Rates	
<b>IVM</b>	40%
<b>IVO</b>	99%

A Sanfins et al. (2003) Biol Reprod 69:2059-2067.

This means,

Signs of meiotic competence loss in IVM oocytes

Partial loss in oocyte quality in IVM

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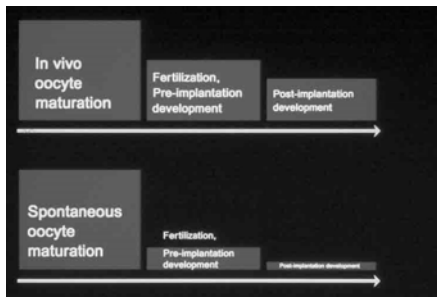
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However, accumulating evidence shows that IVM oocytes exhibit less fertilization and developmental competence than *In Vivo* matured ones



Schroeder and Eppig 1988; Eppig and O'Brien 1998; Cha and Chian 1996; Merrillod et al. 1999; Moor et al. 2001; Trounson et al. 2001

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Our group has been interested in finding both experimental and morphological evidence to explain functional differences between IVM and IVO oocytes, and to improve culture conditions

**Morphological signs of oocyte polarity loss may explain the observed decrease in oocyte quality in IVM oocytes**



**Oocyte polarity may be essential for oocyte function**

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**Oocyte polarity** Evidence in Mammals

Immature oocytes exhibit polarity  
Eccentric GV specifies animal pole

**Human** primary oocyte in primordial follicle     **Rabbit** primary oocyte in primordial follicle     **Hamster** and **Mouse** full grown primary oocytes from antral follicles

TE Schroeder (1985) *Develop Growth Diff* 27:311.

**“All animal eggs have a polar structure”** (Raven, 1961)

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**Oocyte polarity** Evidence in Mammals

Asymmetric organization in oocytes intensifies during maturation

Oocyte Growth → Oocyte Maturation

Prophase I     Metaphase II

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**Oocyte polarity** Evidence in Mammals

Asymmetric organization in oocytes intensifies during maturation

mouse     hamster

OF Albertini, SL Barrett (2004) The developmental origins of mammalian oocyte polarity. *Sem Cell Dev Biol* 15:999.

Mafalda Rato, CE Plancha (2005) Personal results

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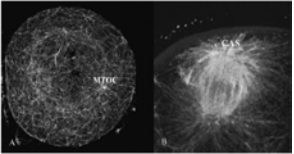
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**Oocyte polarity** **Evidence in Mammals**

Cortical anchoring of GV and meiotic spindle



DF Albertini, SL Barrett (2004) The developmental origins of mammalian oocyte polarity. *Sem Cell Dev Biol* 15:599.



A Sanfins, DF Albertini, CE Plancha (2005) unpublished results.

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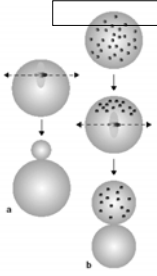
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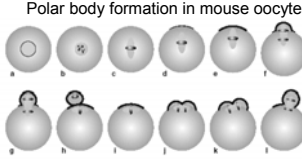
**Oocyte polarity**

**Eccentric meiotic spindle determines polar body emission**



Mechanisms of asymmetric cell division

- . a) different volumes
- . b) different components



Polar body formation in mouse oocytes

B Maro, M-H Verlhac (2002) Polar body formation: new rules for asymmetric divisions. *Nature Cell Biol* 4:E281.

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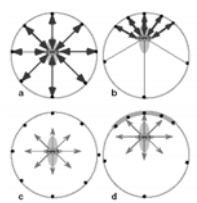
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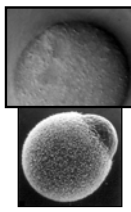
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**Oocyte polarity**

**Eccentric positioning of the meiotic spindle requires formin-2**



Putative role of formin-2 in polar body formation



B Maro, M-H Verlhac (2002) Polar body formation: new rules for asymmetric divisions. *Nature Cell Biol* 4:E281.

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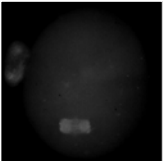


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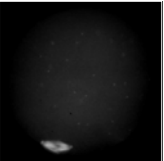
**Oocyte polarity**  
**Signs of oocyte polarity loss in IVM oocytes**

a. GV and spindle anchoring at the cortex is partially lost during IVM

**IVM**



**IVO**



A Sanfins, DF Albertini, CE Plancha (2004) unpublished results.

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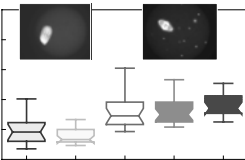
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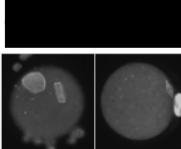
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**Oocyte polarity**  
**Signs of oocyte polarity loss in IVM oocytes**

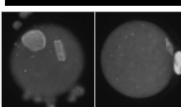
b. Number of cytoplasmic MTOCs partially decreases during IVM



**IVM**



**IVO**



A Sanfins et al. (2003) Distinctions in meiotic spindle structure and assembly during in vitro and in vivo maturation of mouse oocytes. *Biol Reprod* 69: 2059.

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**Oocyte polarity**

**Number of cytoplasmic centrosomes**

**IVM**

IVO: ++ MTOCs

IVM: -- MTOCs

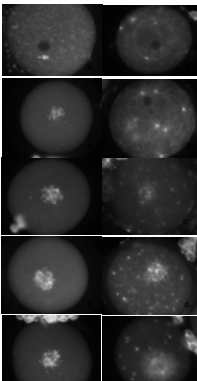
1H

2H

3H

4H

5H



**IVO**

Tubulin  
Centrosomes  
DNA

A Sanfins et al. (2004) Meiotic spindle morphogenesis in in vivo and in vitro matured mouse oocytes: insights into the relationship between nuclear and cytoplasmic quality. *Hum Reprod* 19: 2899-2906.

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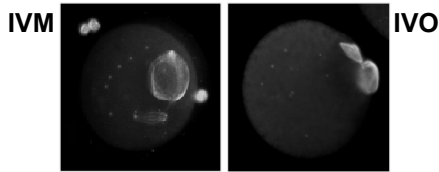
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## Oocyte polarity

Signs of oocyte polarity loss in IVM oocytes

c. Retention of MTOCs and oocyte volume is partially lost during IVM



Polar Bodies formed *in vitro* contain MTOCs

Ibanez et al. (2005) Genetic strain variations in the metaphase-II phenotype of mouse oocytes matured *in vivo* or *in vitro*. *Reproduction* 130:845-55.

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## Oocyte polarity

SL Barrett, DF Albertini (2007) Allocation of gamma-tubulin between oocyte cortex and meiotic spindle influences asymmetric cytokinesis in the mouse oocyte. *Biol Reprod* 76:949-57.

Polar Bodies formed *in vitro* contain MTOCs

Polar Body Status	% of Total	% Int Mts	% W/ Spindles	% W/ MTOCs
<b>IVO</b>				
Undetectable/Degenerate	82.6	0	0	0
Present/ Not Dividing	13.5	13.5	0	0
Dividing/ Post Division	3.8	0	1.9	1.9
<b>IVM Basal</b>				
Undetectable/ Degenerate	0	0	0	0
Present/ Not Dividing	53	52.9	0	29.4
Dividing/ Post Division	47	11.8	35.3	17.6
<b>IVM Hyped</b>				
Undetectable/ Degenerate	16.6	0	0	2.3
Present/ Not Dividing	57.1	47.7	0	33.3
Dividing/ Post Division	28.6	2.3	23.8	23.8

IVM conditions do not promote stable MTOCs that bind to the oocyte cortex

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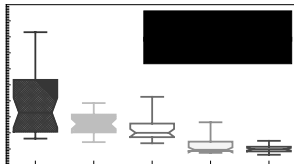
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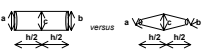
## Oocyte polarity

Why is oocyte polarity important in ARTs?

d. Area of MII meiotic spindle increases during IVM



Quantification of Spindle Size and Shape



$$\text{Area of spindle} = \left( \frac{h/2(\pi r c)}{2} \right) + \left( \frac{h/2(\pi r c)}{2} \right)$$

A Sanfins et al. (2003) Distinctions in meiotic spindle structure and assembly during *in vitro* and *in vivo* maturation of mouse oocytes. *Biol Reprod* 69: 2058.

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## Oocyte polarity

Why is oocyte polarity important in ARTs?

Oocyte polarity and efficiency of asymmetric meiotic division are partially lost during *in vitro* maturation

- GV and spindle anchoring at the cortex
- Number of cytoplasmic MTOCs
- Retention of MTOCs and cell volume by oocyte
- Area of meiotic spindle
- Meiotic spindle pole width
- functional differences?** (meiotic, fertilization and developmental competence)

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## *In vitro* maturation of human oocytes (human IVM)

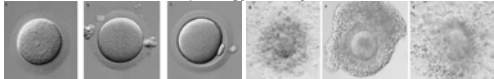
First successful IVM birth:

- from immature oocytes collected at cesarean section for oocyte donation.

Cha et al. (1991) Pregnancy after *in vitro* fertilization of human follicular oocytes collected from nonstimulated cycles, their culture *in vitro* and their transfer in a donor oocyte program. *Fertil Steril* 55:109-113.

IVM treatment have gained promise in ART since it diminishes the need of administrating fertility drugs to induce superovulation.

Nuclear and cellular morphology of oocyte maturation and COCs



Picton (2002) Oocyte maturation *in vitro*. *Curr Opin Obst Gynec* 14:295-302  
Chian, Lim and Tan (2004) State of the art in *in-vitro* oocyte maturation. *Curr Opin Obst Gynec* 16:211-219

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## *In vitro* maturation of human oocytes (human IVM)

Although there are situations were IVM treatment is already recommended

Although there are situations were IVM treatment may turn out to be useful

IVM results need to be improved

### Future directions

Clear need of basic Cell Biology studies on oocyte maturation in animal models and in humans, in order to integrate data into a coherent and scientific knowledge

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