

## MAKING GAMETES AND HELPING EMBRYOS

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1.- Servei de Medicina de la Reproducció.  
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Barcelona, Spain

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- 1. Making gametes
  - Embryonic stem cells (ESC).
  - Derivation and differentiation
  - Gamete differentiation from ESC
    - Derivation of oocytes
    - Derivation of sperm
  - Derivation of oocytes from fetal stem cells
  - BM stem cells as a source of gametes?
- 2. Helping embryos
  - Assisted hatching
  - PGS

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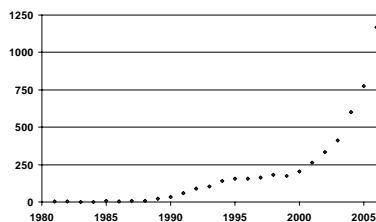
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REPORTS  
**Embryonic Stem Cell Lines Derived from Human Blastocysts**  
James A. Thomson\*, Joseph Itskovitz-Eldor, Sander S. Shapiro, Michelle A. Waknitz, Joseph J. Swiergiel, Victoria S. Marshall, Jeffrey H. Jones  
www.sciencemag.org SCIENCE VOL 282 6 NOVEMBER 1998 1145



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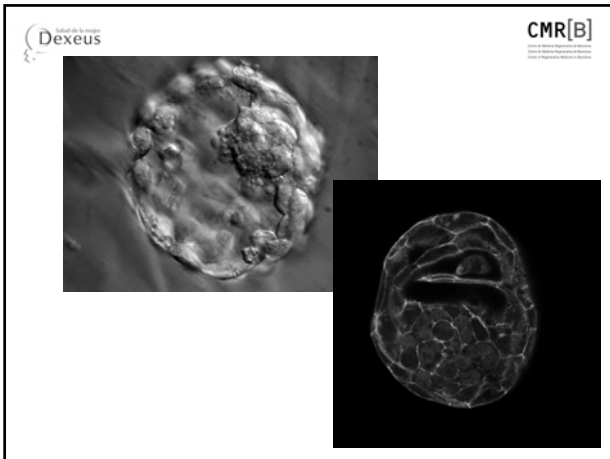
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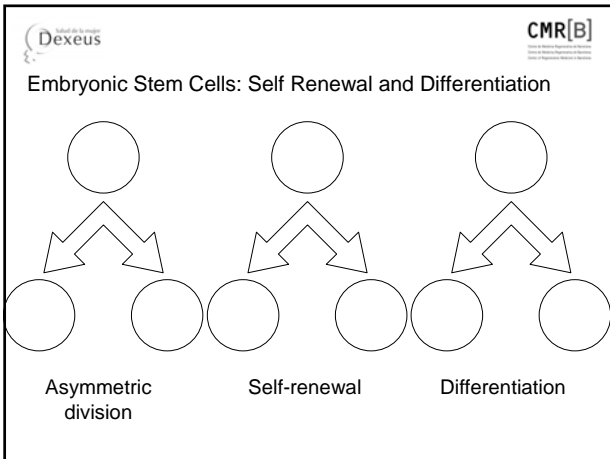
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<p><b>Differentiation</b></p> <p>ADIPOCITO Dani et al., 1997          ASTROCITO Fraichard et al., 1995          CARDIOMIOCITO Doetschman et al., 1985          Maltsev et al., 1993          CONDROCITO Kramer et al., 2000          HEMATOPOYÉTICAS Nakano et al., 1996          DEFINITIVAS Nishikawa et al., 1996          Wiles et al., 1991          CELULAS DENDRÍTICAS Fairchild et al., 2000          CELULAS ENDOTELIALES Risau et al., 1988          Yamashita et al., 2000          KERATINOCITOS Bagutti et al., 1996          Yamashita et al., 2000          PRECURSORES DE LINFOCITOS Potocnik et al., 1994</p>	<p>MASTOCITOS Tsai et al., 2000          NEURONAS Bain et al., 1995          Strubing et al., 1995          OLIGODENDROCITOS Brastle et al., 1999          Liu et al., 2000          OSTEOBLASTOS Buttery et al., 2001          ISLOTES PANCREÁTICOS Lumelski et al., 2001          HEMATOPOYÉTICAS Doetschman et al.,          PRIMITIVAS 1985          Nakano et al., 1996          MÚSCULO LISO Yamashita et al., 2000          MÚSCULO ESTRIADO Rohwedel et al., 1994          ENDODERMO DEL SACO Doetschman et al.,          VITELINO 1985          MESODERMO DEL SACO Doetschman et al.,          VITELINO 1985</p>
<p><b>GERM CELLS - GAMETES</b></p>	

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Dexeus CMR[B]

## Derivation of Oocytes from Mouse Embryonic Stem Cells

Karin Hübner,<sup>1</sup> Guy Fuhrmann,<sup>3</sup> Lane K. Christenson,<sup>4</sup>  
 James Kehler,<sup>1</sup> Roland Reinbold,<sup>1</sup> Rabindranath De La Fuente,<sup>2</sup>  
 Jennifer Wood,<sup>4</sup> Jerome F. Strauss III,<sup>4</sup> Michele Boiani,<sup>1</sup>  
 Hans R. Schöler<sup>1\*</sup>

www.sciencemag.org SCIENCE VOL 300 23 MAY 2003

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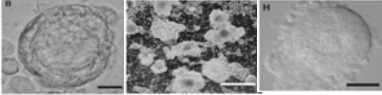
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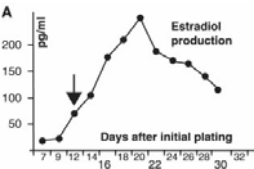
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Dexeus CMR[B]

### Formation of follicular structures



After 2 weeks in culture, structures similar to advanced follicle-like structures appear. Most of them degenerate but around 20% give rise to "oocytes" > 40 µm.



Days after initial plating

• The functional activity of the follicles is evidenced by estradiol production

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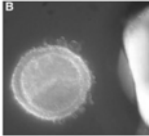
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Dexeus CMR[B]

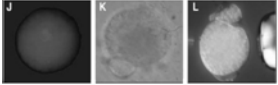
### Oocyte characterisation

- At D26 of culture, oocyte-like cells ( 50-70 µm) are released from surrounding somatic cells and float freely in the supernatant.



- All ZP markers are + except ZP1 (accounting for fragile ZP structure)

Addition of gonadotrophins to isolated follicles results in extrusion of oocytes and formation of a polar body-like structure



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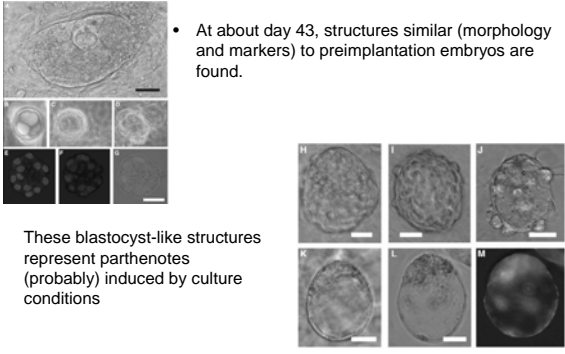
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Dexeus CMR[B]

### Blastocyst-like structures derived from ES cells

- At about day 43, structures similar (morphology and markers) to preimplantation embryos are found.



These blastocyst-like structures represent parthenotes (probably) induced by culture conditions

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Dexeus CMR[B]

Differentiation (2007) 75:902-911 DOI: 10.1111/j.14724648.2007.00181.x © 2007, Copyright the Authors  
Journal compilation © 2007, International Society of Differentiation

**ORIGINAL ARTICLE**

Tingting Qiang · Yan Shi · Han Qin · Xin Ye · Wei Wei · Haisong Liu · Mingxiao Ding · Hongkui Deng

### Induction of oocyte-like cells from mouse embryonic stem cells by co-culture with ovarian granulosa cells

Our results demonstrate that granulosa cells were effective in inducing the differentiation of ES cell-derived PGCs into oocyte-like cells through direct cell-to-cell contacts. Our method offers a novel *in vitro* system for studying oogenesis; in particular, for studying the interactions between PGCs and granulosa cells.

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Dexeus CMR[B]

### Derivation of embryonic germ cells and male gametes from embryonic stem cells

Niels Geijsen<sup>1,2</sup>, Melissa Horoschak<sup>1,3</sup>, Kitai Kim<sup>1,3</sup>, Joost Gröbnau<sup>1</sup>, Kevin Eggan<sup>4</sup> & George Q. Daley<sup>1,3</sup>

<sup>1</sup>Whitehead Institute for Biomedical Research, 9 Cambridge Center, Cambridge, Massachusetts 02142, USA  
<sup>2</sup>Center for Regenerative Medicine and Technology, Massachusetts General Hospital, Boston, Massachusetts 02114, USA  
<sup>3</sup>Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, and Division of Pediatric Hematology/Oncology, The Children's Hospital and Dana Farber Cancer Institute, Boston, Massachusetts 02115, USA  
<sup>4</sup>Department of Molecular and Cellular Biology, Harvard University, 7 Divinity Avenue, Cambridge, Massachusetts 02138, USA

NATURE | VOL 427 | 8 JANUARY 2004 | www.nature.com/nature

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**Dexeus** **CMR[B]**

- Culture conditions: Embryoid body differentiation
- Marker expression pluripotency (Oct4).
- GC development markers (*Stella*, *Fragilis*)
- Genes exclusively expressed in GC line (*Dazl*, *Piwil2*, *Rnf17*, *Rnh2*, *Tdrd1* y *Tex14*)

During embryoid body formation pluripotency markers expression decreased with some expression of GC specific genes

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**Dexeus** **CMR[B]**

### PGC differentiation to functional gametes

- Upregulation of genes associated with male germ cell development (Sry, Acrosin, Haprin, markers of Sertoli cells)
- No expression of Zp proteins.
- Within the context of EB differentiation, the default program of female gametogenesis is suppressed.

### Meiosis

EB microenvironment is permissive for male germ cell development and meiotic maturation, even though highly inefficient

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**Dexeus** **CMR[B]**

#### Sperm morphology

EB derived cells have a similar morphology to testis derived haploid cells. Round spermatids?

#### Biological functionality

- ICSI with haploid cells into recipient oocytes
- 50% of the embryos cleaved to 2 cell stage
- 20% reach the blastocyst stage.
- FISH: normal for sex chromosomes

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Dexeus CMR[B]

**Embryonic stem cells can form germ cells *in vitro***

Yayoi Toyooka, Naoki Tsunekawa, Ryuko Akasu, and Toshiaki Noze<sup>a</sup>

<sup>a</sup>Mitsubishi Kagaku Institute of Life Sciences, 11 Minamioya Machi-4-chi, Tokyo 134-8511, Japan

PNAS | September 30, 2003 | vol. 100 | no. 20 | 11459

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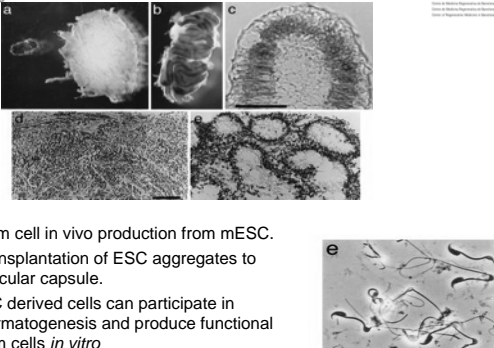
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Dexeus CMR[B]



- Germ cell *in vivo* production from mESC.
- Transplantation of ESC aggregates to testicular capsule.
- ESC derived cells can participate in spermatogenesis and produce functional germ cells *in vitro*

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Dexeus CMR[B]

**In Vitro-Differentiated Embryonic Stem Cells Give Rise to Male Gametes that Can Generate Offspring Mice** Short Article

Nayernia et al, *Developmental Cell* 11, 125-132, July 2006

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Dexeus CMR[B]

- Establishment of two cell lines (Spermatogonial stem cells- SSCs) with patterns of differentiation towards male germ cells .
- Derived from mESC (directed gene expression for premeiotic and haploid male germ cells)
- Motility of cells
- Formation of tail-like structures

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Dexeus CMR[B]

- Positive for PGCs, premeiotic (Oct 4, Fragilis, Stella,...), meiotic (Scp3, Acr) and postmeiotic male germ cells markers
- Formation of sperm structures.
- Acrosome like structures
- Condensation of the nucleus

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Dexeus CMR[B]

- Disruption of methylation imprints?
- ICSI of in vitro-generated cells (haploid cells) into oocytes of wildtype females.
- Polar body extrusion, PN formation and normal features of embryo development.
- Transfer into the oviducts of pseudopregnant females
- 65 embryos transferred
- 12 animals born.
- Larger or smaller offspring
- Premature death (5 days to 5 months after birth).
- Abnormal methylation patterns and phenotypic abnormalities

⇒ interference with the essential reprogramming events in gametogenesis

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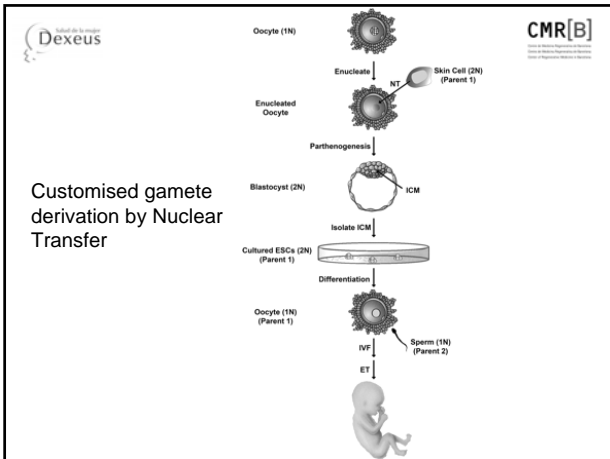
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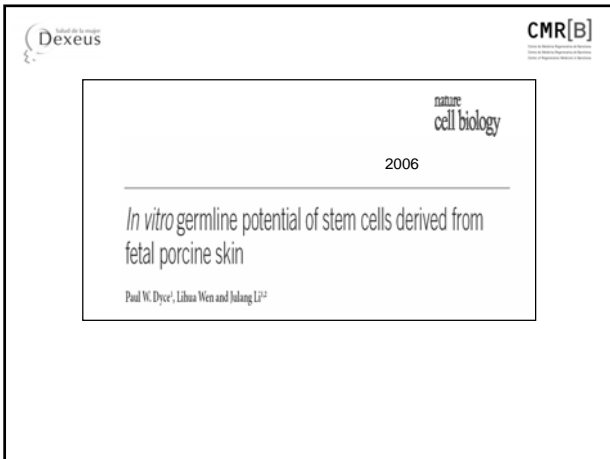
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**Stem cells isolated from the skin of porcine fetuses have the intrinsic ability to differentiate into oocyte-like cells.**

**When differentiation was induced, a subpopulation of these cells expressed markers (Oct4, GDF9b, DAZL and Vasa), consistent with germ-cell formation.**

**On further differentiation, these cells formed follicle-like aggregates that secreted oestradiol and progesterone: responded to gonadotropin stimulation.**

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Dexeus CMR[B]

- Some of these aggregates extruded large oocyte-like cells that expressed oocyte markers, such as ZP, and meiosis markers.(SCP3).
- Some of these oocyte-like cells spontaneously developed into parthenogenetic embryo-like structures.

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Dexeus CMR[B]

Laboratory Investigation (2006) 86, 654–663  
© 2006 Elsevier Inc. All rights reserved. DOI:10.1038/labinvest.2006.100  
www.elsevier.com/locate/jlabinv

### Derivation of male germ cells from bone marrow stem cells

Karin Nayernia<sup>1</sup>, Jan Ho Lee<sup>1</sup>, Nadja Drusenheimer<sup>2</sup>, Jessica Nolte<sup>3</sup>, Gerald Wulf<sup>4</sup>, Ralf Dressel<sup>5</sup>, Jörg Gromoll<sup>1</sup> and Wolfgang Engel<sup>1</sup>

<sup>1</sup>Institute of Human Genetics, University of Göttingen, Göttingen, Germany; <sup>2</sup>Department of Haematology and Oncology; <sup>3</sup>Department of Cellular and Molecular Immunology, University of Göttingen, Göttingen, Germany and <sup>4</sup>Institute of Reproductive Medicine, University of Münster, Münster, Germany

Recent studies have demonstrated that somatic stem cells have a more flexible potential than expected, whether put into tissue or cultured under different conditions. Bone marrow (BM)-derived stem cells can transdifferentiate into multilineage cells, such as muscle of mesoderm, lung and liver of endoderm, and brain and skin of ectoderm origin. Here we show that **BM stem cells are able to transdifferentiate into male germ cells.** For derivation of male germ cells from adult BM stem (BMS) cells, we used the Stra8-enhanced green fluorescence protein (EGFP) transgenic mouse line expressing EGFP specifically in male germ cells. BMS cell-derived germ cells expressed the known molecular markers of perinatal germ cells, such as fragilis, stella, Rnf17, Hth and Oct4, as well as molecular markers of spermatogonial stem cells and spermatogonia including Rbm, c-Kit, Tex19, Stra8, Pih12, Dazl, Hsp90 $\alpha$ ,  $\beta$ 1- and  $\beta$ 2-integrins. Our ability to derive male germ cells from BMS cells reveals novel aspects of germ cell development and opens the possibilities for use of these cells in reproductive medicine.

Laboratory Investigation (2006) 86, 654–663. doi:10.1038/labinvest.3700429; published online 1 May 2006

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Dexeus CMR[B]

## Conclusions

- Functional gamete-like cells can be obtained from embryonic and fetal stem cells.
- BM stem cells give rise to male gametes
- Customised gametes may be obtained from ESC using NT techniques.
- Derivation of gametes from SC brings new insights into germ cell development , epigenetic reprogramming and germline gene modification

\* Stem cell derived gametes can become a valuable resource for research. The use of such gametes in ART remains a "distant prospect"

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# Assisted Zona Hatching

## What is the evidence?

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### Embryo Assisted hatching

- Assisted hatching (AH) involves the artificial thinning or breaching of the ZP.
- Has been proposed as one technique to improve implantation and pregnancy rates following *in vitro* fertilization (IVF).
- An increased implantation rate following mechanical opening of the ZP (partial zona dissection-PZD) was first reported in 1990 (Cohen *et al* 1990)
- The assisted hatching procedure can be performed on day 3 or day 5 after fertilization.
  - drilling with acidified Tyrode's solution
  - PZD with a glass microneedle
  - Laser photoablation
  - Use of a piezomicromanipulator

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### Embryo Assisted hatching

TABLE 1  
Studies reporting ongoing pregnancies/live births.

Study included	No. of patients		No. of ET		Mean age (y)		Implantation rate (%)				Clinical pregnancy rate (%)		Ongoing deliveries (%)	
	Ctrl	AH	Ctrl	AH	Ctrl	AH	Ctrl	AH	Ctrl	AH	Ctrl	AH	Ctrl	AH
Cohen <i>et al</i> 1990 (2)	89	89	229	229	36.7	36.5	21*	29*	47	54	20/88 (23)	34/99 (34)	Delivered	ET
Haddad <i>et al</i> 1999 (3)	60	60	192	192	35.8	35.9	17.1	17.8	36.1	42.1	21/90 (23)	23/90 (26.7)	Delivered	ET
Hart <i>et al</i> 1999 (3)	7	13	4.8 ± 0.7	4.9 ± 0.7	30 ± 6.8	30 ± 6.8	10.7*	9.9*	42	33	9/7 (13)	2/10 (20)	Delivered	PR
Lambert <i>et al</i> 1999 (3)	50	42	212	180	35.5	35.3	11.3*	11.1*	41.7	36.0	17/49 (34.7)	13/41 (31.7)	Ongoing pregnancy	PR
Mikrou <i>et al</i> 2000 (10)	25	27	75	80	33.2	32.1	NA	NA	40	48	9/25 (36)	10/27 (37)	Delivered	cycle
Mikrou <i>et al</i> 2000 (10)	41	39	135	117	36.3	37.3	NA	NA	7.25	225	3/4 (7.5)	6/39 (28)	Delivered	cycle

Ctrl = control, AH = assisted hatching, PR = patient.  
 \* Data recorded per embryo replaced.  
 † Mean ± SD.  
 ‡ Number of gestational sacs per number of embryos transferred.  
 § Significantly different, P < 0.05, Fisher's exact test.  
 NA = Not Assisted Hatching, NA = Not Assisted.

The role of assisted hatching in *in vitro* fertilization: a review of the literature. A Committee opinion  
 The Practice Committee of the Society for Assisted Reproductive Technology and the Practice Committee of the American Society for Reproductive Medicine  
 Fertil Steril Vol 85, Suppl 4, Nov 2006

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Conclusions

- The available published evidence does not support the routine or universal application of assisted hatching in all IVF cycles at this time.
- Assisted hatching may be clinically useful in patients with a poor prognosis, including those with 2 failed IVF cycles and poor embryo quality and advanced age women (38 years of age)
- Higher clinical pregnancy and implantation rates have been observed after assisted hatching. However, delivery rates have not significantly improved, possibly because the small sample sizes in studies reporting delivery rates have lacked sufficient power to detect a difference.

Individual ART programs should evaluate their own patient populations to determine which subgroup may benefit from AH

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Preimplantation genetic screening (PGS)  
Is it useful for advanced age patients?

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Preimplantation genetic screening for abnormal number of chromosomes (aneuploidies) in in vitro fertilisation or intracytoplasmic sperm injection (Review)

Twisk M, Mastenbroek S, van Wely M, Heineman MJ, Van der Veen F, Repping S

This record should be cited as:  
Twisk M, Mastenbroek S, van Wely M, Heineman MJ, Van der Veen F, Repping S. Preimplantation genetic screening for abnormal number of chromosomes (aneuploidies) in in vitro fertilisation or intracytoplasmic sperm injection. *Cochrane Database of Systematic Reviews* 2006, Issue 1. Art. No.: CD005291. DOI: 10.1002/14651858.CD005291.pub2.

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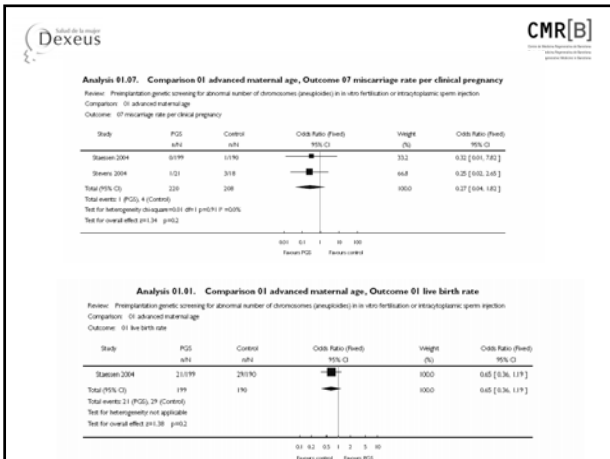
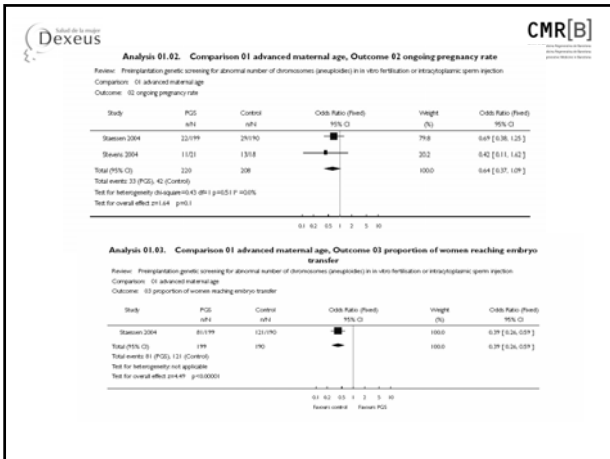
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*The* **NEW ENGLAND**  
**JOURNAL of MEDICINE**

ESTABLISHED IN 1812      JULY 5, 2007      VOL. 357 NO. 1

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**In Vitro Fertilization with Preimplantation Genetic Screening**

Sebastiaan Mastenbroek, M.Sc., Moniek Twisk, M.D., Jannie van Echten-Arends, Ph.D.,  
 Birgit Sikkema-Raddatz, Ph.D., Johanna C. Korevaar, Ph.D., Harold E. Verhoeve, M.D., Naali E.A. Vogel, M.D.,  
 Eus G.J.M. Arts, Ph.D., Jan W.A. de Vries, Ph.D., Patrick M. Bossuyt, Ph.D., Charles H.C.M. Buys, Ph.D.,  
 Maas Jan Heijnen, M.D., Ph.D., Sjoerd Repping, Ph.D., and Fulco van der Veen, M.D., Ph.D.

**Dexeus** **CMR[B]**

**Table 2. Outcomes in Women Who Underwent Preimplantation Genetic Screening and in Controls.**

Outcome	Women Who Underwent Preimplantation Genetic Screening (N=206)	Controls (N=202)	Rate Ratio (95% CI) <sup>a</sup>	P Value
Women with an ongoing pregnancy — no. (%)	52 (25)	74 (37)	0.69 (0.51–0.93)	0.01
Women with ≥1 biochemical pregnancy — no. (%)	81 (39)	106 (52)	0.75 (0.60–0.93)	0.008
Total no. of biochemical pregnancies	94	118		
Women with ≥1 clinical pregnancy — no. (%)	61 (30)	88 (44)	0.68 (0.52–0.88)	0.003
Total no. of clinical pregnancies	67	92		
Women with ≥1 miscarriage — no. (%)	37 (18)	36 (18)	1.01 (0.67–1.53)	0.97
Total no. of miscarriages	43 <sup>†</sup>	44 <sup>‡</sup>		
Women with ≥1 live birth — no. (%)	49 (24)	71 (35)	0.68 (0.50–0.92)	0.01
Total no. of live births	59 <sup>§</sup>	85 <sup>¶</sup>		

<sup>a</sup> CI denotes confidence interval.  
<sup>†</sup> One miscarriage occurred at 18 weeks of gestation; all other miscarriages occurred before 12 weeks of gestation.  
<sup>‡</sup> All miscarriages occurred before 12 weeks of gestation.  
<sup>§</sup> There were 39 singleton and 10 twin births; one woman underwent elective termination of pregnancy, one pregnancy ended in an intrauterine death, and one premature delivery resulted in the postpartum death of a twin.  
<sup>¶</sup> There were 57 singleton and 14 twin births; two women underwent elective termination of pregnancy, and one pregnancy ended in an intrauterine death.

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- Dexeus** **CMR[B]**
- Multicentre trial of preimplantation genetic screening reported in the New England Journal of Medicine: an in-depth look at the findings.  
Cohen J, Grifo JA.
  - Substandard application of preimplantation genetic screening may interfere with its clinical success.  
Munné S, Gianaroli L, Tur-Kaspa I, Magli C, Sandalinas M, Grifo J, Cram D, Kahraman S, Verlinsky Y, Simpson JL.
  - IVF with preimplantation genetic screening, a promising new treatment with unexpectedly negative health outcomes: the Hippocratic role of Data Monitoring Committees.  
Ankum WM, Reitsma JB, Offringa M; Hippocratic role of Data Monitoring Committees.

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