

Methods to assess the status of the sperm DNA:

The Comet and TUNEL Assays

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Learning Objectives

- Background and nature of Comet and TUNEL assays
 - What is measured?
 - Differences in results due to method variants
 - Differences due to different DNA organization
- Notes on the Halosperm® assay
- Alternative interpretation of assay results

Origin of the Comet assay

- Undamaged DNA is believed to remain in organized structure with nuclear proteins while damaged DNA cannot maintain the high degree of organization
- Single-cell gel electrophoresis – originally after exposure to chemical substance
 - Detection of genotoxic potential of chemical compounds
 - Human biomonitoring – effects on entire human organisms
 - Developed for various somatic cell types and tissues
 - Requires relatively few cells, rapidly performed, relatively eas

Origin of the Comet assay

- Principle:
 - Cells embedded in an agarose matrix
 - Cell lysis – salt, detergent, pH
 - Electrophoresis, pH
 - DNA staining
 - Microscopy
 - Image analysis

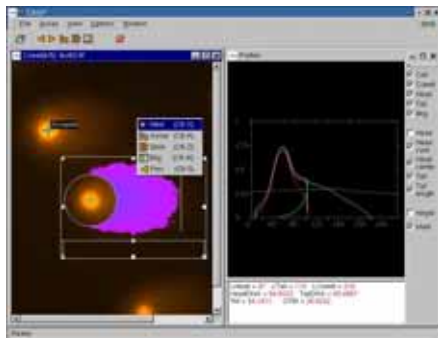
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Origin of the Comet assay

- Alkaline Comet – high pH
 - Single- and double strand DNA breaks
 - Alkali labile sites
 - Incomplete repair sites
 - Crosslinks
- Neutral Comet – pH around 7
 - single- and double strand breaks
- Standardization
 - International recommendations, somatic cells:
 - International Workshop on Genotoxicity Testing (IWGR)
 - Studies on germ cells:
 - only limited data from studies done under controlled conditions and GLP

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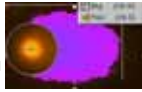
Calculation of results CASP – Comet Assay Software Project www.casplab.com



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Calculation of results CASP – Comet Assay Software Project
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- **HeadArea** Area of the comet head
- **TailArea** Area of the comet tail
- **HeadDNA** Sum of intensities of pixels in the head
- **TailDNA** Sum of intensities of pixels in the tail
- **HeadDNA%** Percent of intensity of pixels in the comet head
- **TailDNA%** Percent of intensity of pixels in the comet tail
- **HeadRadius** Radius of the comet head
- **TailLength** Length of the comet tail
- **CometLength** Length of the entire comet from head area to end of tail
- **HeadMeanX** Center of gravity of intensity in the head (x coordinate)
- **TailMeanX** Center of gravity of intensity in the tail (x coordinate)
- **TailMoment** TailDNA% x TailLength
- **OliveTailMoment** TailDNA% x (TailMeanX-HeadMeanX)



Alterations of the Comet assay for sperm investigations

- Additional steps to allow access to the sperm DNA
 - Many different approaches, various background levels
 - In general much higher "background" levels in spermatozoa
- Sperm Chromatin Decondensation
 - Proteinase K after lysis, various temperatures, duration 3-18 h
 - RNase and Proteinase K
 - DMSO
 - DTT – zinc-binding and disulfide-bridge cleaving agent
 - Combined with Lithium diiodosalicylate (LIS)
 - Shortest time for decondensation
 - Reduced baseline DNA breaks

Alterations of the Comet assay for sperm investigations

- No standard developed for sperm Comet electrophoresis
 - pH
 - Time
 - Voltage and amperage
- Alkaline conditions increase DNA migration (background) in somatic cells
 - NOT due to pre-existing DNA breaks
 - Due to alkali-sensitive sites
 - In published sperm studies 5-97% of DNA migrates!

Alterations of the Comet assay for sperm investigations

- Sperm Comet assay often run under neutral conditions
 - Supposedly lower background
 - Alkaline conditions appear to result in higher variability between studies
 - NOT specific for DNA double strand breaks
 - Mainly single-strand breaks
 - Lower sensitivity

Studies of genotoxicity in vitro exposure

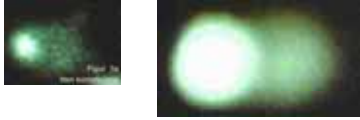
- DNA of ejaculated sperm is resistant to X-ray or H₂O₂ exposure
 - Difficult to induce DNA breaks with external agents WHEN THE NUCLEAR ORGANIZATION IS INTACT!
- Exposure to chemical compounds
 - Agents able to bind to and interact with DNA
 - Presence during decondensation?
- Still valid to investigate potential DNA damaging compounds *in vitro*

In vivo exposure – human biomonitoring

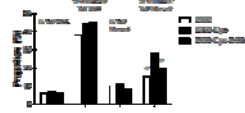
- Few results from studies on humans – much variation
- Age, tobacco smoking, environmental exposure
- Confounding factors
 - Abstinence time (storage before ejaculation)
 - Post-ejaculatory storage conditions

Chromatin stabilization and Comet assay

- Exposure of sperm to Cysteine results in huge increase in DNA loss



- If Cysteine is washed away before Comet assay, Tail Moment is not increased



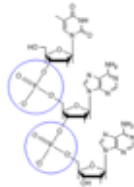
- Tu et al, 2009, Poster presentation at American Society of Andrology, Philadelphia, PA, USA, April 4-7.

Origin of the TUNEL assay

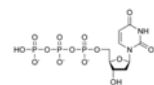
- Terminal deoxynucleotidyl transferase dUTP nick end labeling
- Assessment of programmed cell death – apoptosis
- Non random DNA breaks by endogenous endonucleases creating short, regular DNA sections
 - Fast renewing tissues – gut epithelium

Origin of the TUNEL assay

- Markers of “open” DNA endings – “nicks”
 - Single strand breaks of phosphodiester bonds
- Proteins stripped off cell nuclei with proteinase K, 15 min, room temperature
 - Inactivation of endogenous peroxidase with H₂O₂
 - Addition of Terminal deoxynucleotidyl transferase (TdT) that labels DNA endings with dUTP



phosphodiester bonds
source: Wikipedia



dUTP - Chemical structure of Deoxyuridine triphosphate; source: Wikipedia

Adjustment of the TUNEL assay for human spermatozoa

- Cell lysis and nuclear decondensation
 - NaCl, EDTA, Tris, detergent
 - RNase
 - Proteinkinase K
- Fixation with paraformaldehyde
- Permeabilization with detergent

- Assessment by fluorescence microscopy or flow cytometry

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TUNEL assay for human spermatozoa

- Expose to nucleotides (dUTP) and TdT enzymes
 - Positive control: DNase I treatment
 - Negative control: absence of TdT enzymes

- More stain bound to DNA endings:
 - Positive controls (DNase I)
 - Exposure to H₂O₂

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TUNEL assay and chromatin stability

- Using In Situ Cell Detection Kit (Fluorescein)
 - Positive controls only stained 15-53% of sperm
 - High percentage TUNEL-positive spermatozoa related to
 - Low seminal zinc concentration
 - Long abstinence time
 - Long time between ejaculation and start of TUNEL preparation

% TUNEL (N=9), coefficient of correlation (r)			
	r _s	r _t	P _{ns}
Zinc conc. (nM)	-0.73	-0.53	0.008
Abstinence time (days)	0.48	0.63	0.01
Analysis delay (hrs)	0.95	0.98	0.0001
% TUNEL (Pos. control)	0.31	0.09	ns (0.42)
Sperm conc. (10 ⁶ /mL)	-0.17	-0.08	ns (0.87)

From Björk et al, 2009, Poster presentation at the American Society of Andrology, Philadelphia, PA, USA, April 4-7.

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Some notes on the Halosperm® assay

Principle

- Gel embedded spermatozoa
- Denaturation with 0.08 N HCl
- Deproteinization with detergent, DTT and washed with EDTA
- Dehydrated with increasing ethanol concentration



Interpretation

- Spermatozoa **with** DNA-halo: low DNA fragmentation
- Spermatozoa **without** DNA-halo: fragmented DNA outside the core of the nucleus washed away

Fernandez et al (2003) The sperm chromatin dispersion test: a simple method for the determination of sperm DNA fragmentation. J Androl, 24, 59-66.

Clinical usefulness – basic aspects

- A test that correlates well with for instance ART outcome is not necessarily clinically useful
- A clinically useful test must distinguish between at least two groups of patients
 - Overlap between groups is a bad "sign" for the test
- The frequency of the "disorder" in the investigated population is also an essential factor
 - A rare state of health can be concealed by numerous false positive cases

Clinical usefulness – basic aspects

- Good Laboratory Practice (GLP) – *diagnosis* and *prognosis* for infertile couples
 - validation and evaluation of methods using statistical measures like Odds Ratio and Relative Risk, not correlations
 - "What is the probability that this patient has a certain disorder?"
- Reproductive Toxicology
 - Evaluation of effects of chemical compounds
 - Comparison of groups with different exposure
 - Comparison of spermatozoa with different exposures
 - Difference between controls and exposed individuals or cells
 - "Where is the baseline level?"

Summary

- Results from Comet, TUNEL and Halosperm® assays
 - Depends on access to the DNA
 - Level and type of sperm chromatin stabilization likely to play a crucial role for the outcome
 - Use in reproductive medicine is different from reproductive toxicology
 - Absolute level of DNA damage cf. Relative effect of exposure

References

- <http://www.casplab.com/>
- <http://www.cometassay.com/>
- Burlinson et al (2007) *Mutat Res*, **627**, 31-35.
- Fernandez et al (2003) *J Andrology*, **24**, 59-66.
- From Björk et al (2009). *J Androl*, **Suppl March/April**, p40-41.
- Gavrieli et al(1992) *J Cell Biol*, **119**, 493-501.
- Gold et al (1993) *J Histochem Cytochem*, **41**, 1023-1030.
- Lewis et al(2008) *Mutagenesis*, **23**, 163-170.
- Schlegel & Paduch (2005) *Fertility and sterility*, **84**, 854-859.
- Shen & Ong (2000) *Free Radic Biol Med*, **28**, 529-536.
- Speit et al (2009) *Mutat Res*, **681**, 3-12.
- Tu et al (2009) *J Androl*, **Suppl March/April**, p42.
