

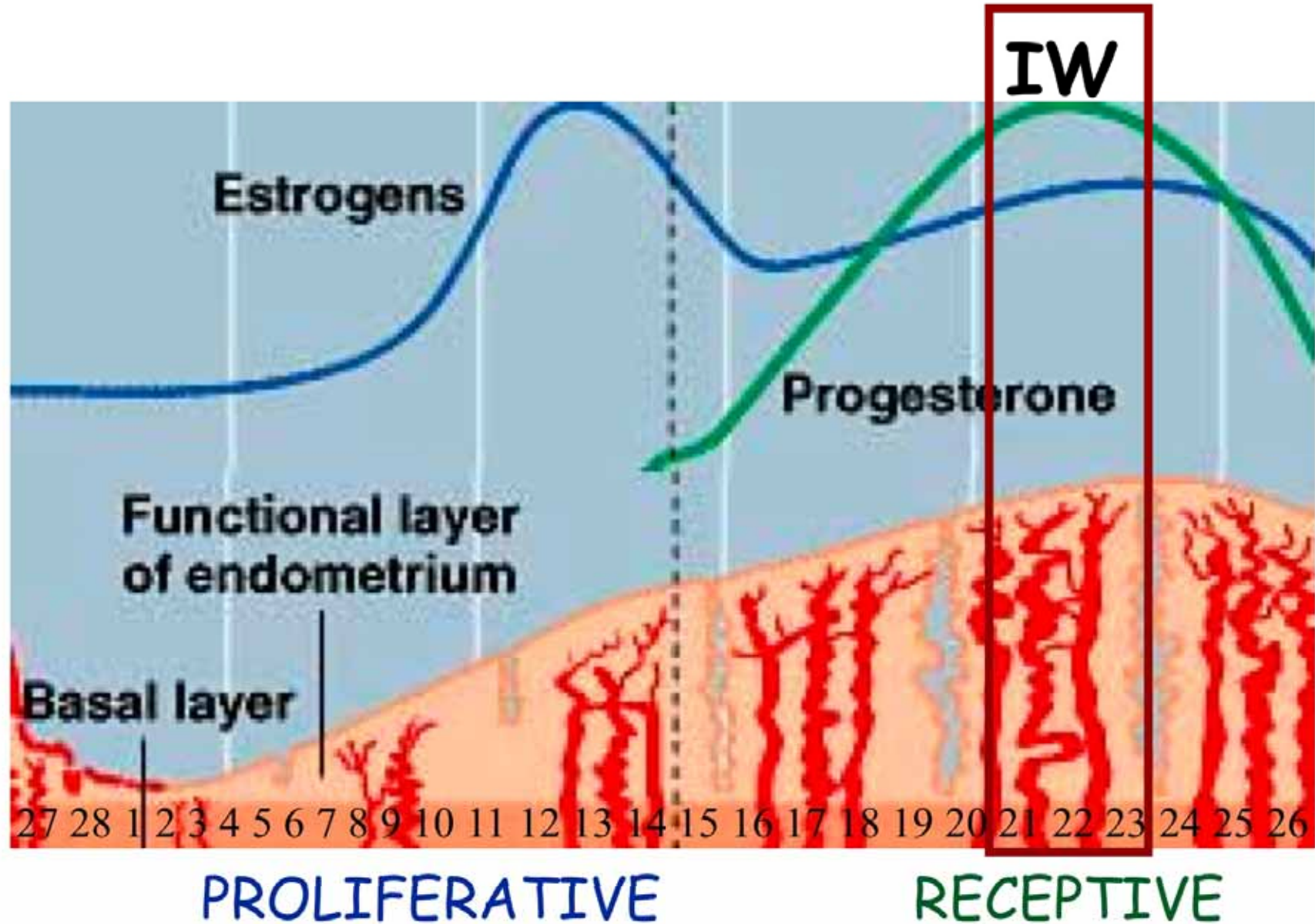
Genes targeted by the estrogen and progesterone receptors in human endometrium

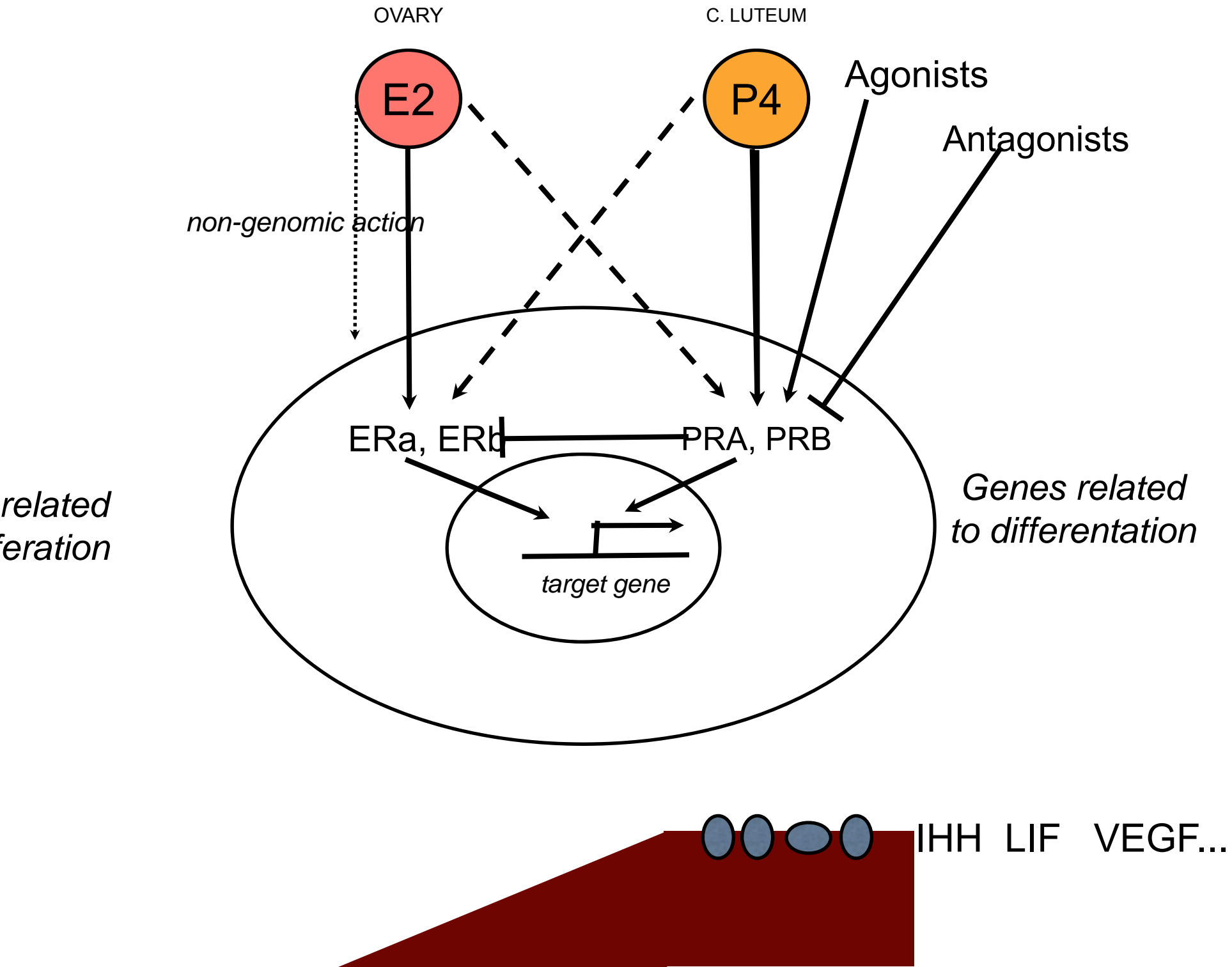
Karin Tamm

Centre for Biology of Integrated Systems
Tallinn University of Technology



Human Endometrium





E2, P4 10^{-8} M 45'

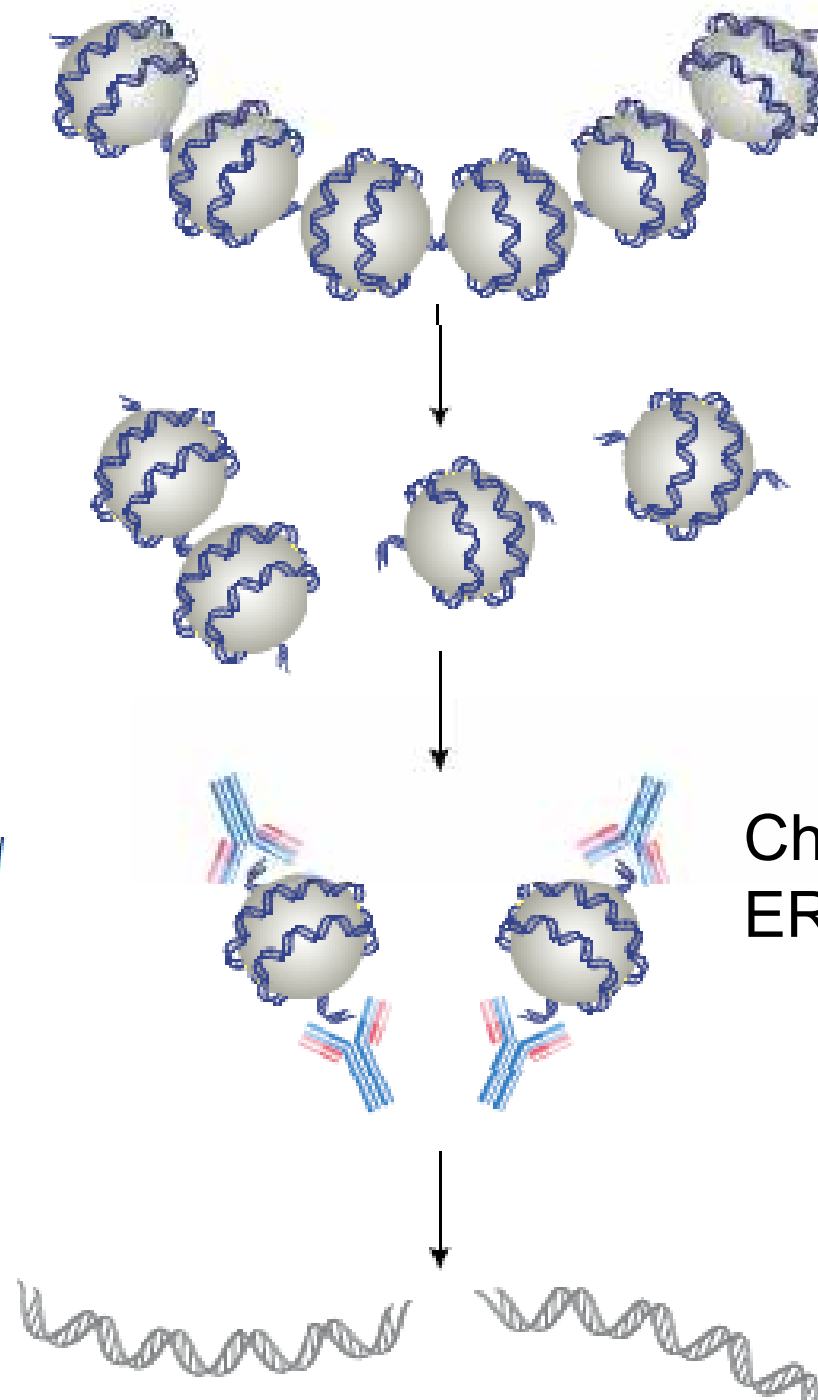
Method Overview

Cells are fixed with formaldehyde to cross-link histone and non-histone proteins to DNA.

Chromatin is digested with Micrococcal Nuclease into 150-900 bp DNA/protein fragments.

Antibodies specific to histone or non-histone proteins are added and the complex co-precipitates and is captured by Protein G agarose or Protein G magnetic beads.

Cross-links are reversed, and DNA is purified and ready for analysis.



ChIP using Ab-s against
ER α , ER β , PRAB,

ChIP enriched

Pre-
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nes

A. HEC1A cell line

B. RL95-2 cell line

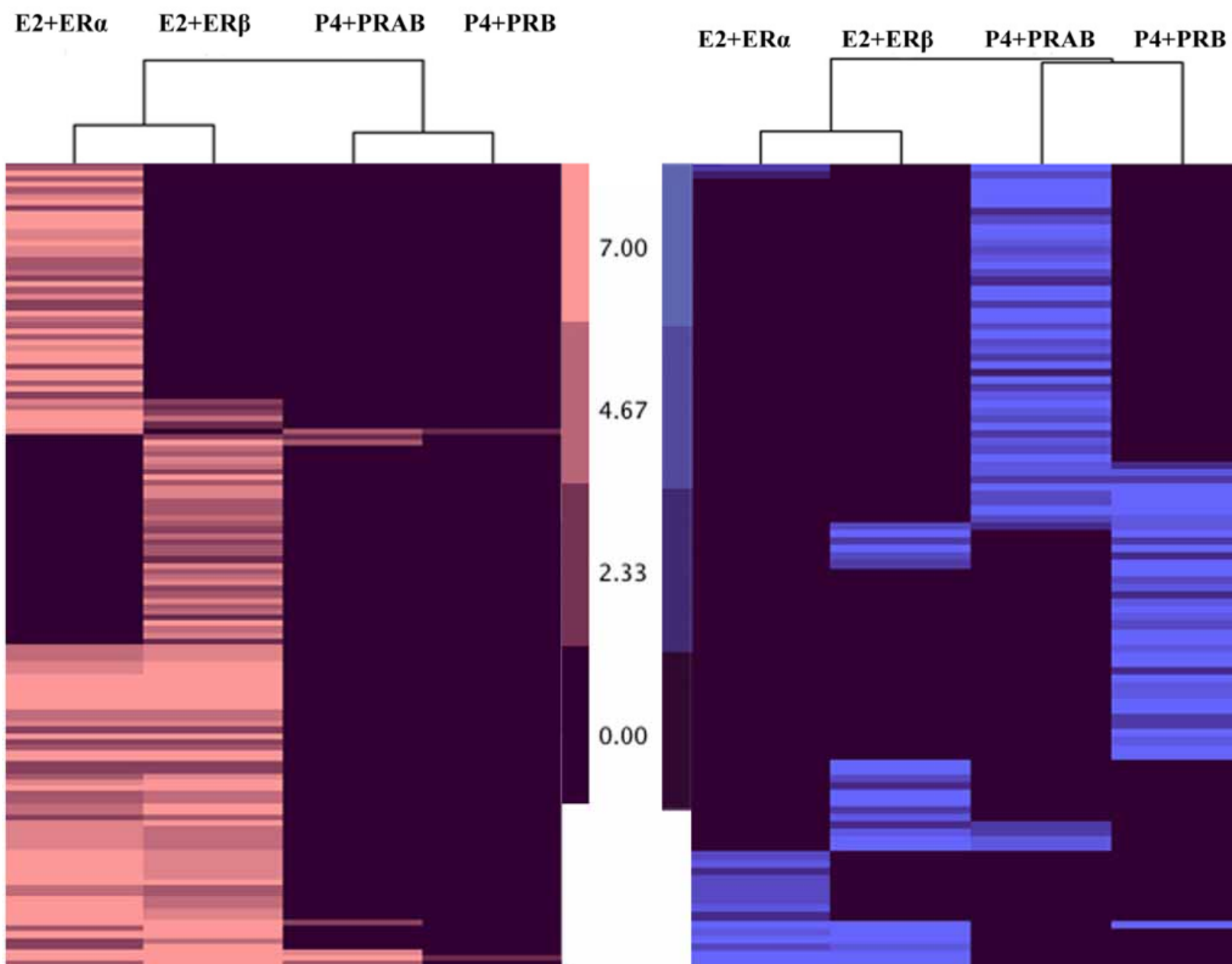
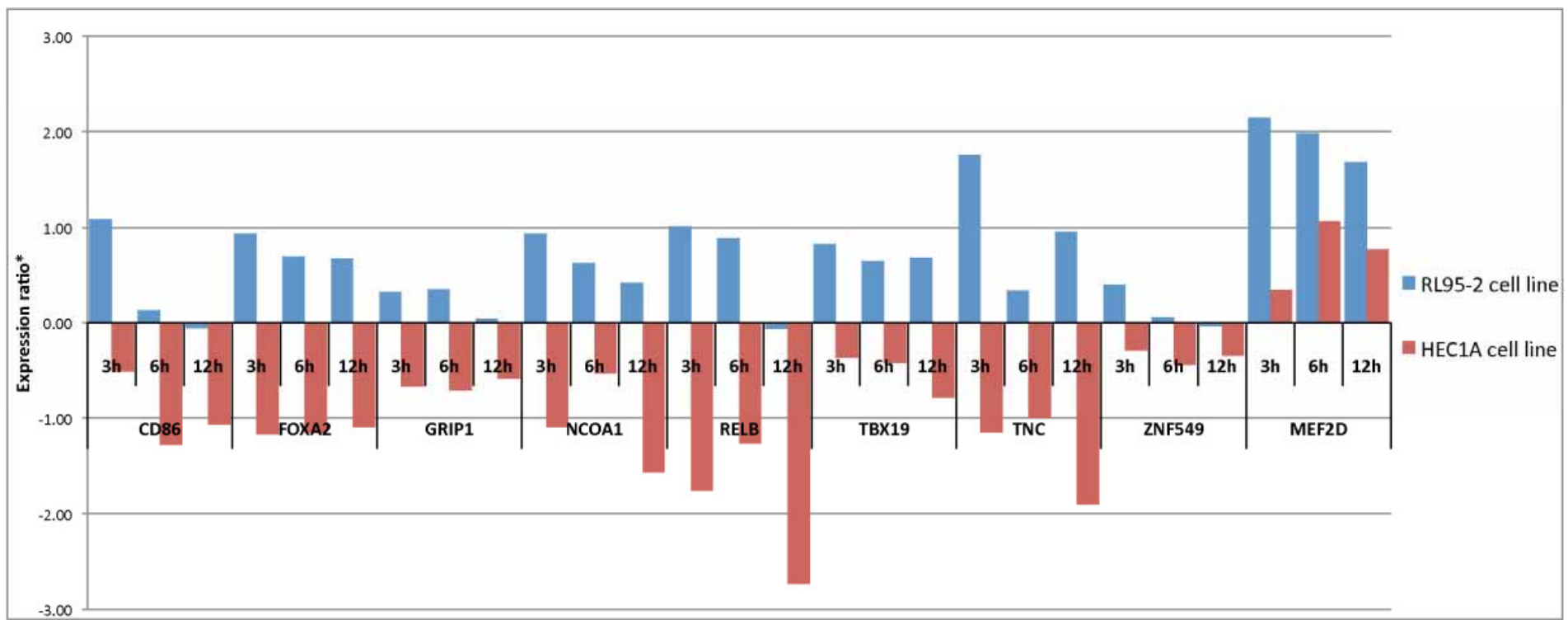
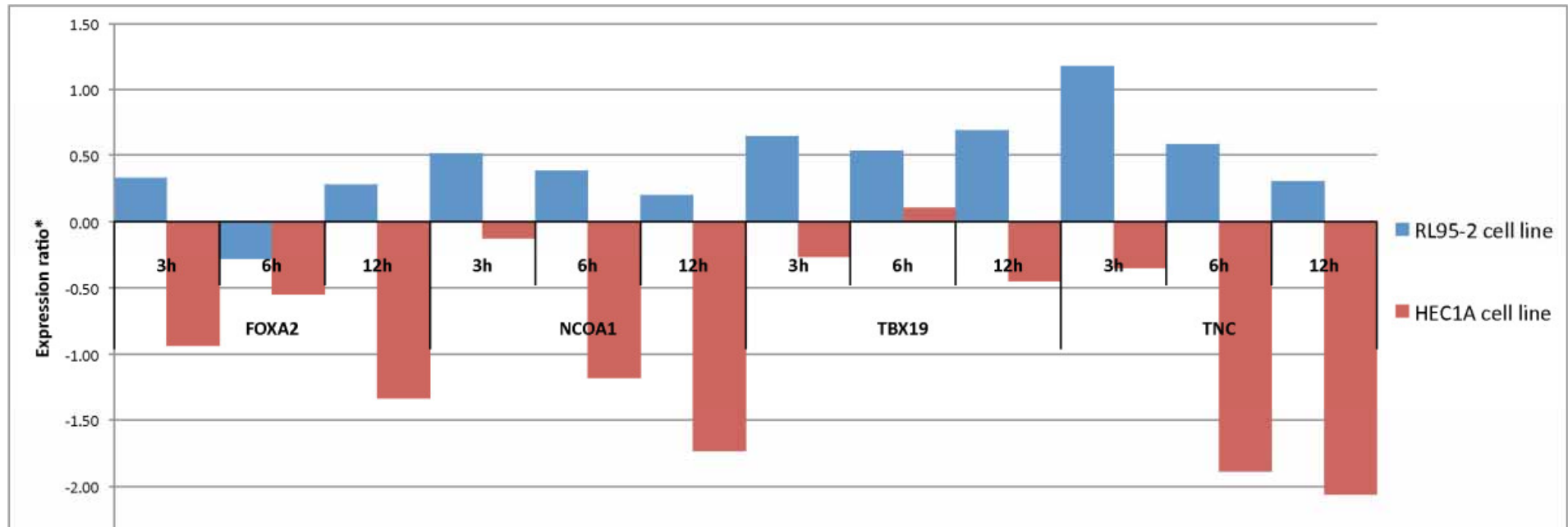


Fig. 4. Clustering of genes (ER α , ER β , PRAB, PRB) and conditions in HEC1A and RL95-2



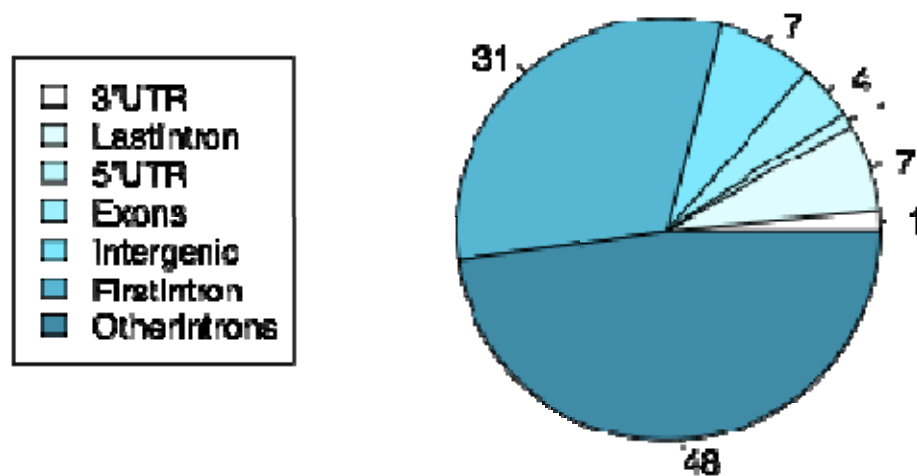
mediated time-dependant gene expression compared to non-treated samples.



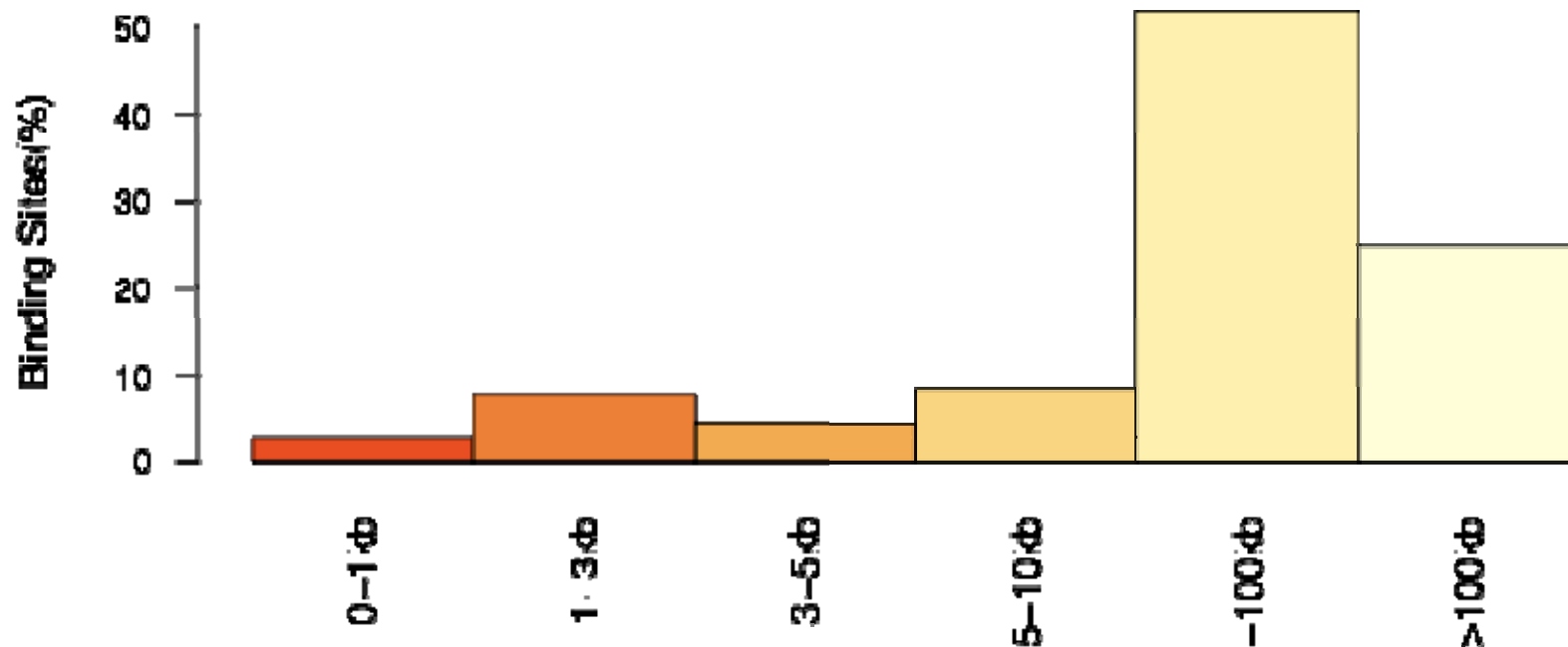
Identifying ER and PR targets and action on entire human genome

- ChIP-Seq: identifying ER and PR targets in whole genome using *Illumina Genome Analyzer IIe*
 - *in vitro*: Ishikawa cell line
 - *in vivo*: Endometrial biopsy samples
- RNA-Seq: describing the entire transcriptome of human endometrium.
 - *in vitro*
 - *in vivo*

% Peaks overlapping gene features

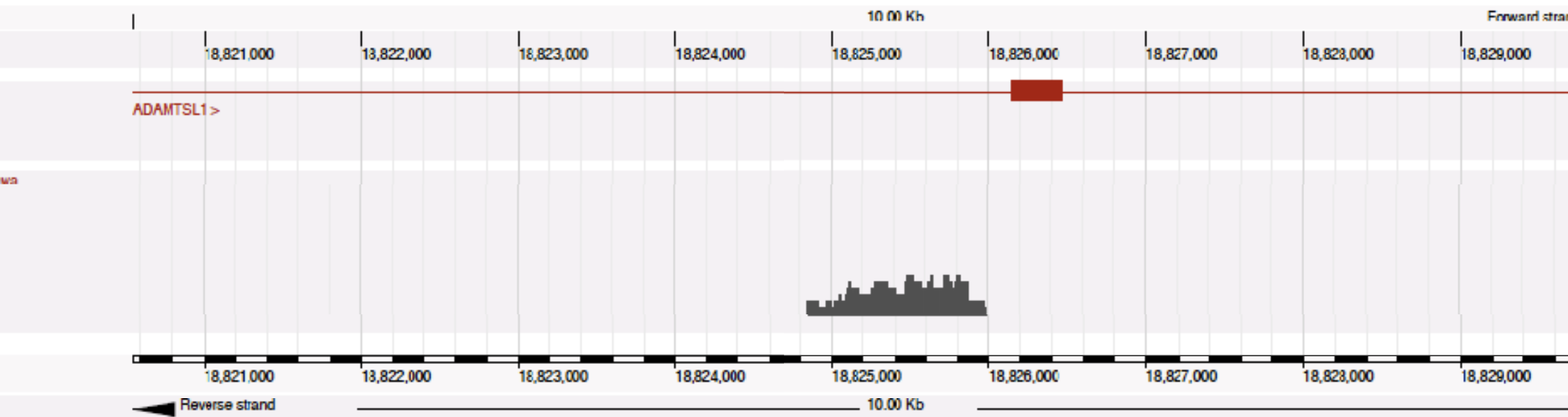


Distance to nearest downstream gene



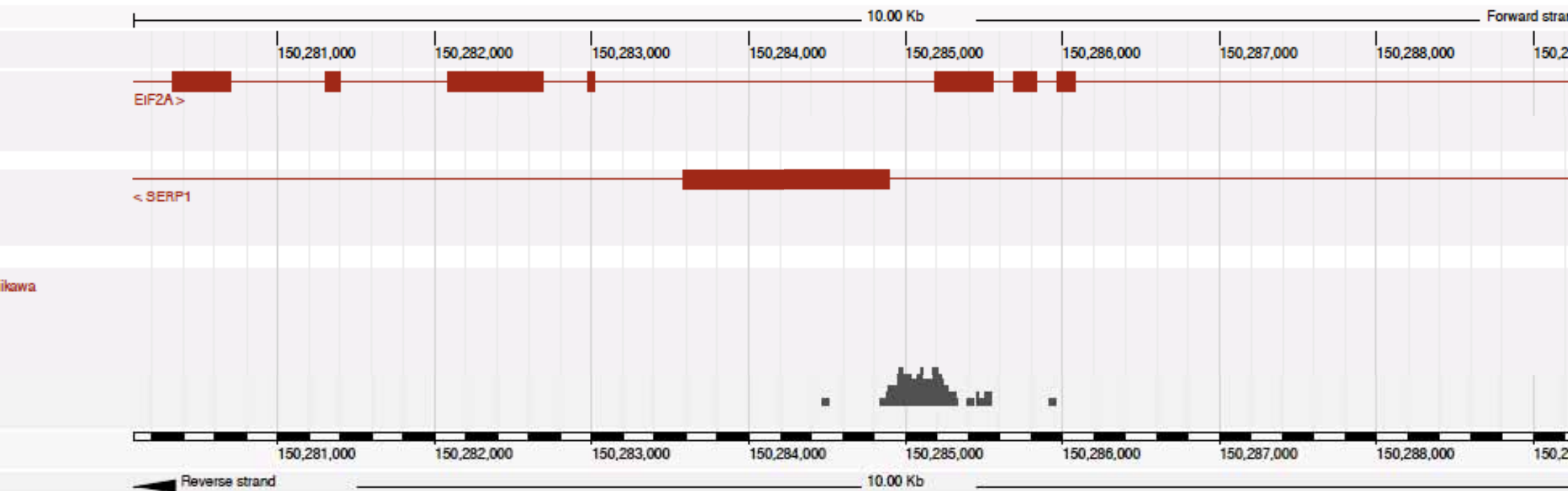
ER target in *ADAMTSL1* after E2 treatment

encodes a secreted protein and member of the ADAMTS (a disintegrin and metalloproteinase with thrombospondin motif) family. ADAMTS have important functions in the extracellular matrix. Alternative splicing results in multiple transcript variants encoding distinct p

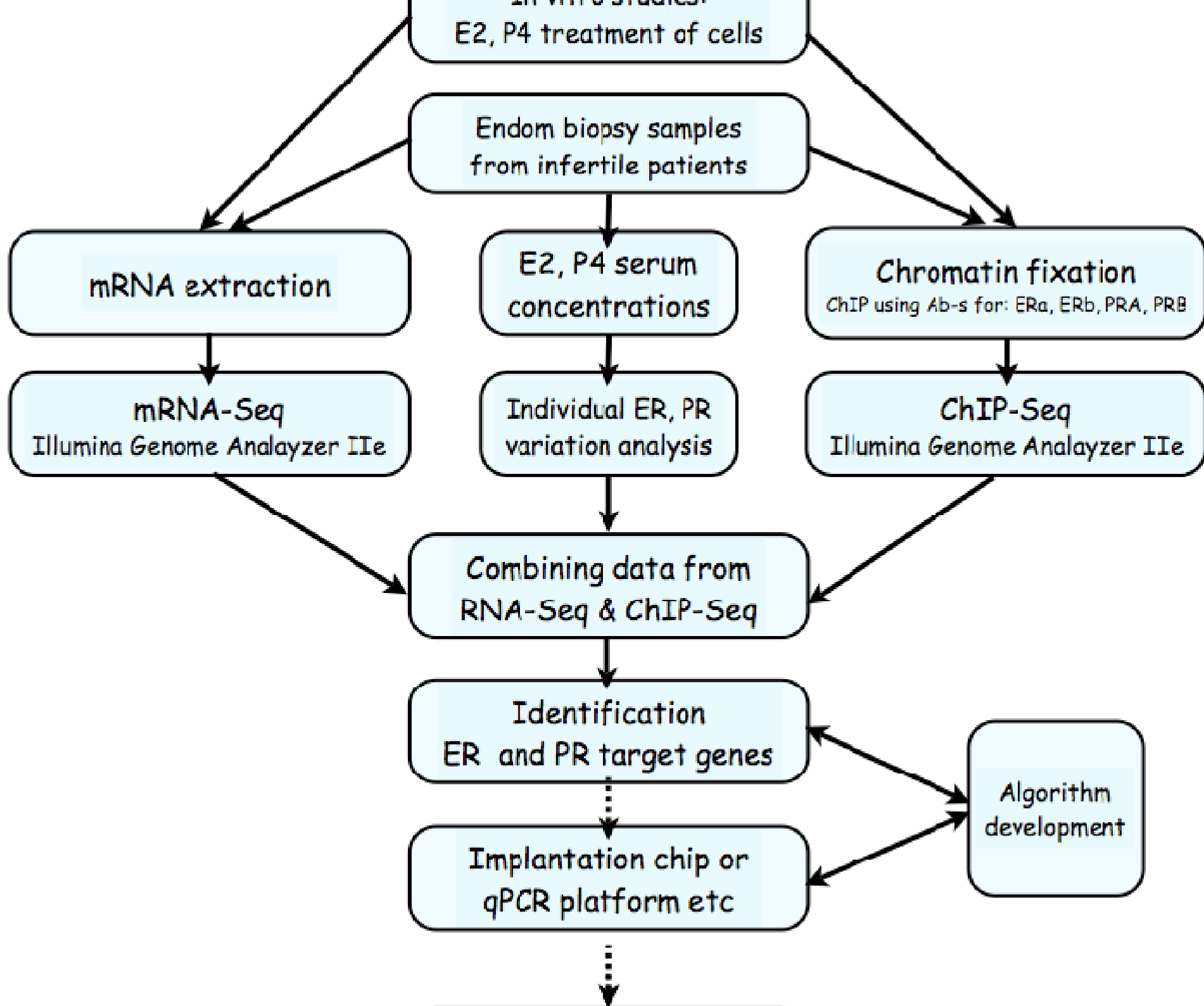


ER target in *EIF2A* without E2 treatment

aryotic Translation Initiation Factor 2



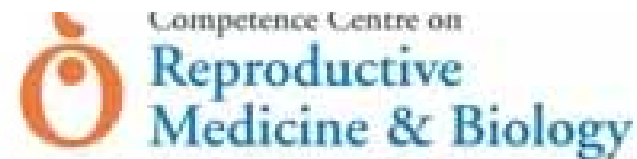
PeakAnalyz



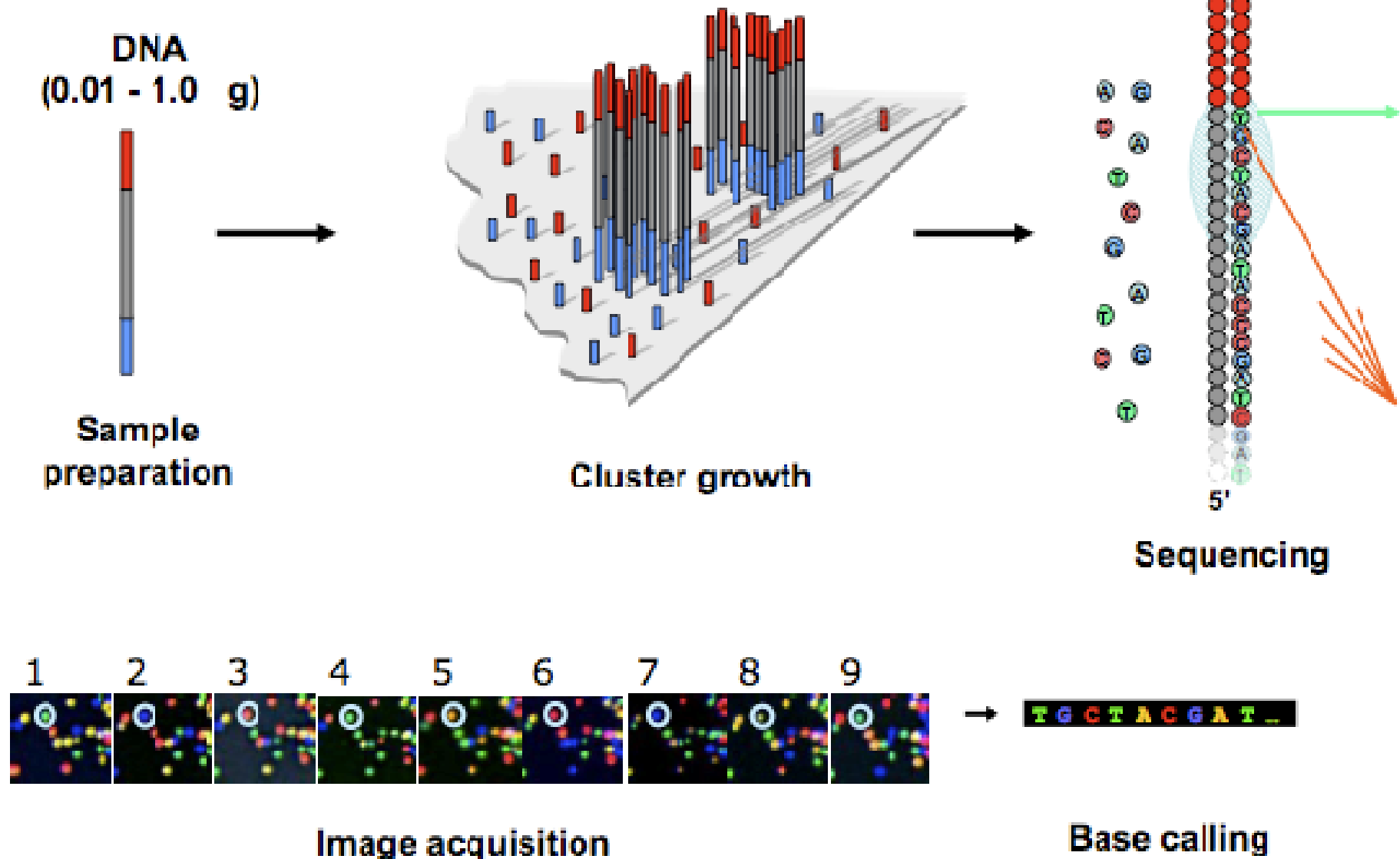
Many thanks

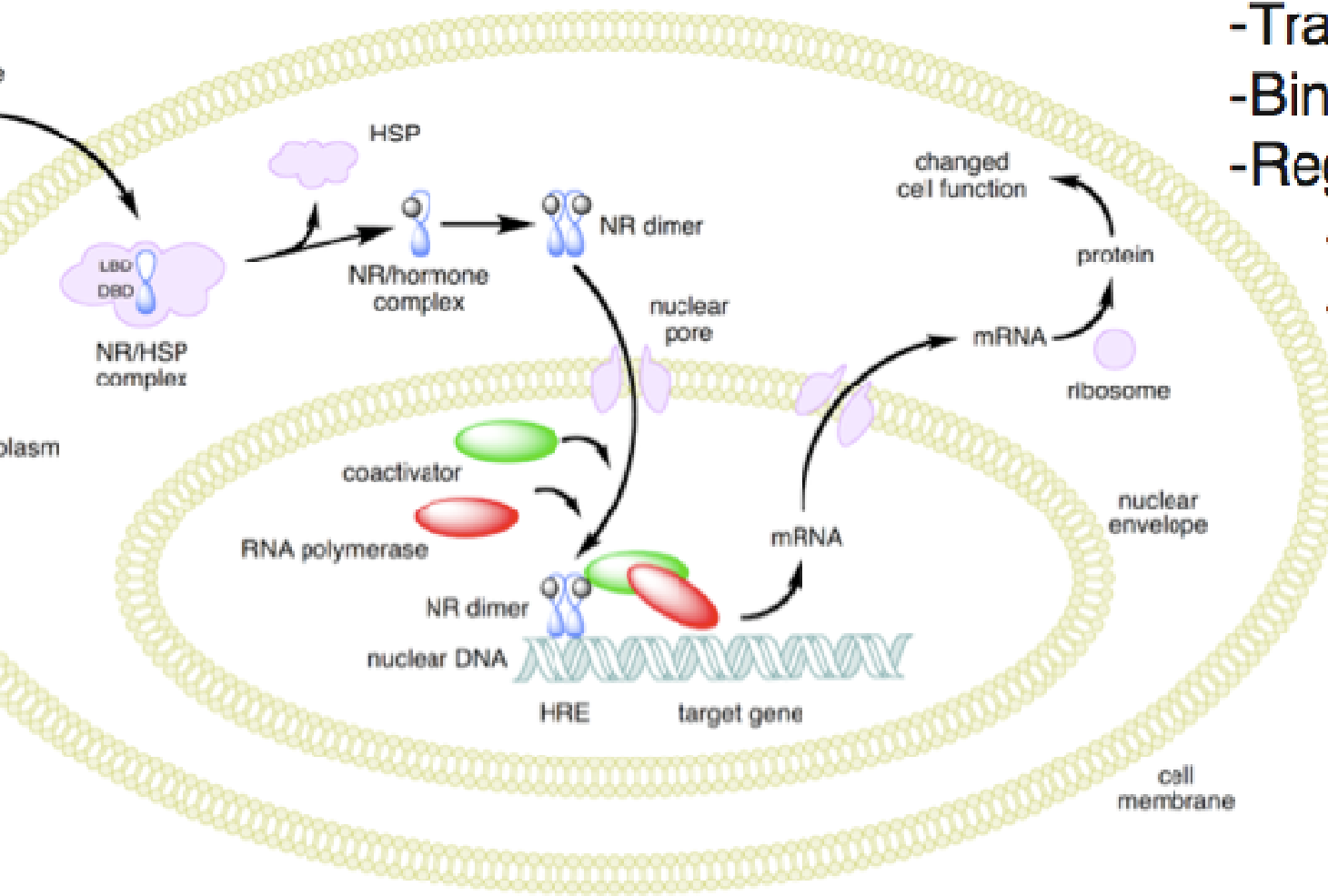
- Miia Rõõm
- Jaak Simm
- Kairi Tammoja

- Madis Metsis
- Andres Salumets
- Sten Linnarson



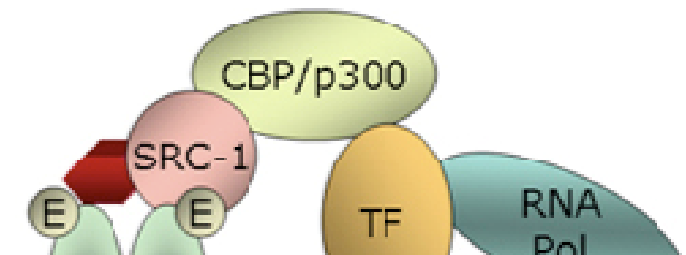
Solexa Sequencing

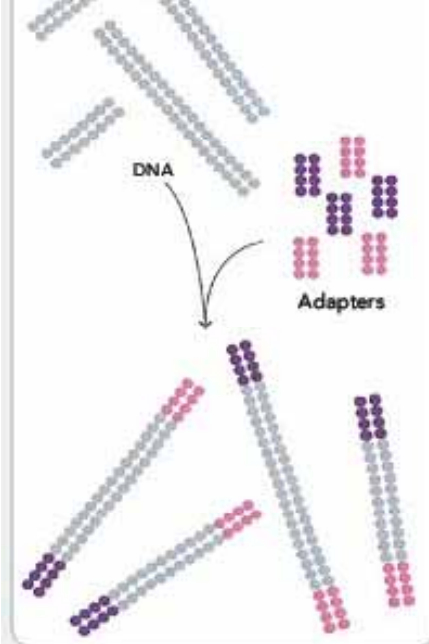




- Hormone binding to the NR
- Translocation to the nucleus
- Binding to the HRE
- Regulation transcription of target gene
- mRNA translated into protein
- Change in cell function

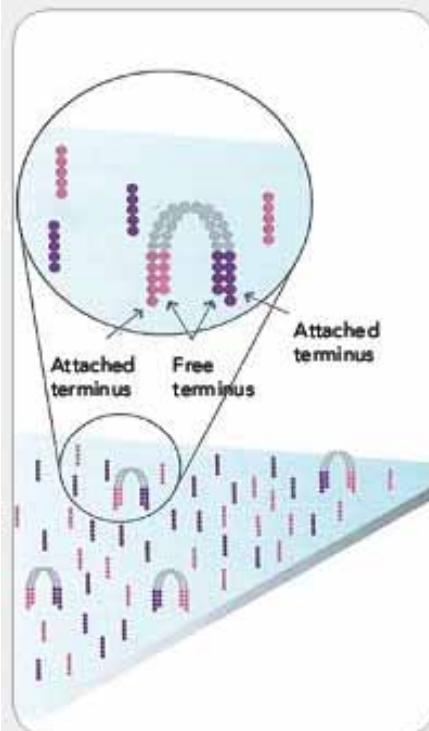
nuclear receptor DNA complex recruits



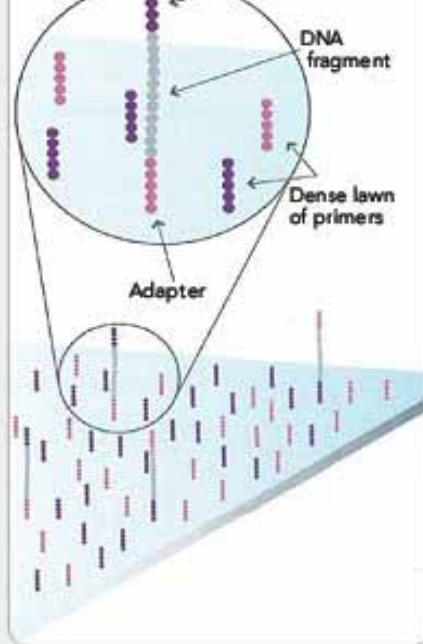


Randomly fragment genomic DNA and ligate adapters to both ends of the fragments.

4. FRAGMENTS BECOME DOUBLE STRANDED

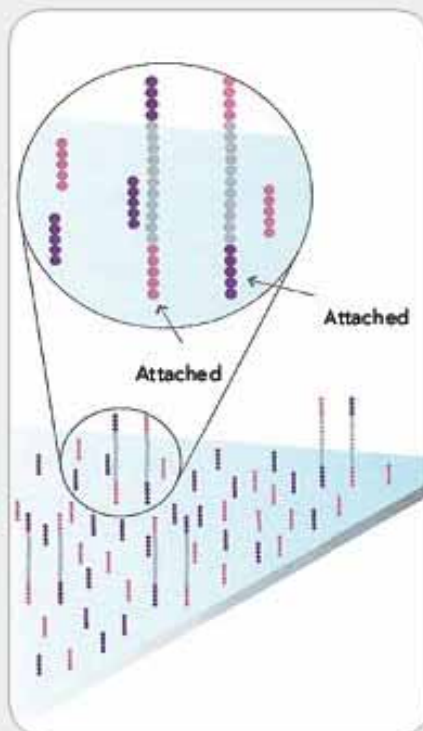


The enzyme incorporates nucleotides to

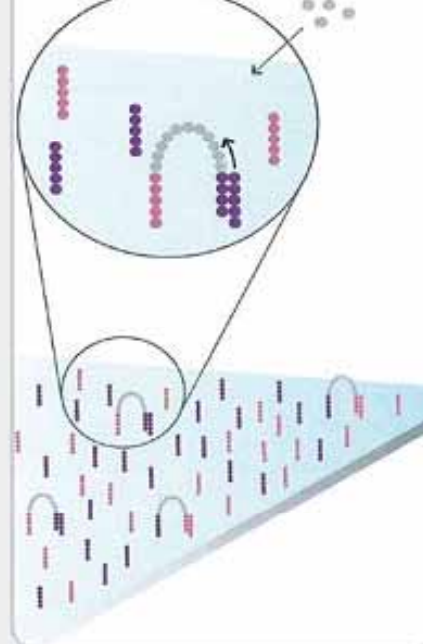


Bind single-stranded fragments randomly to the inside surface of the flow cell channels.

5. DENATURE THE DOUBLE-STRANDED MOLECULES

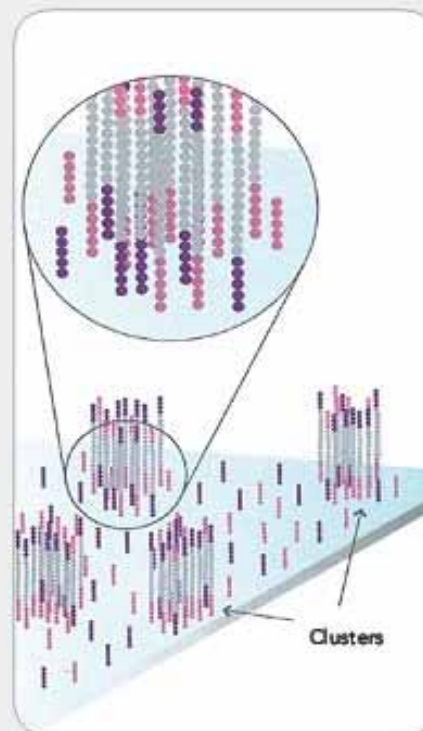


Denaturation leaves single-stranded



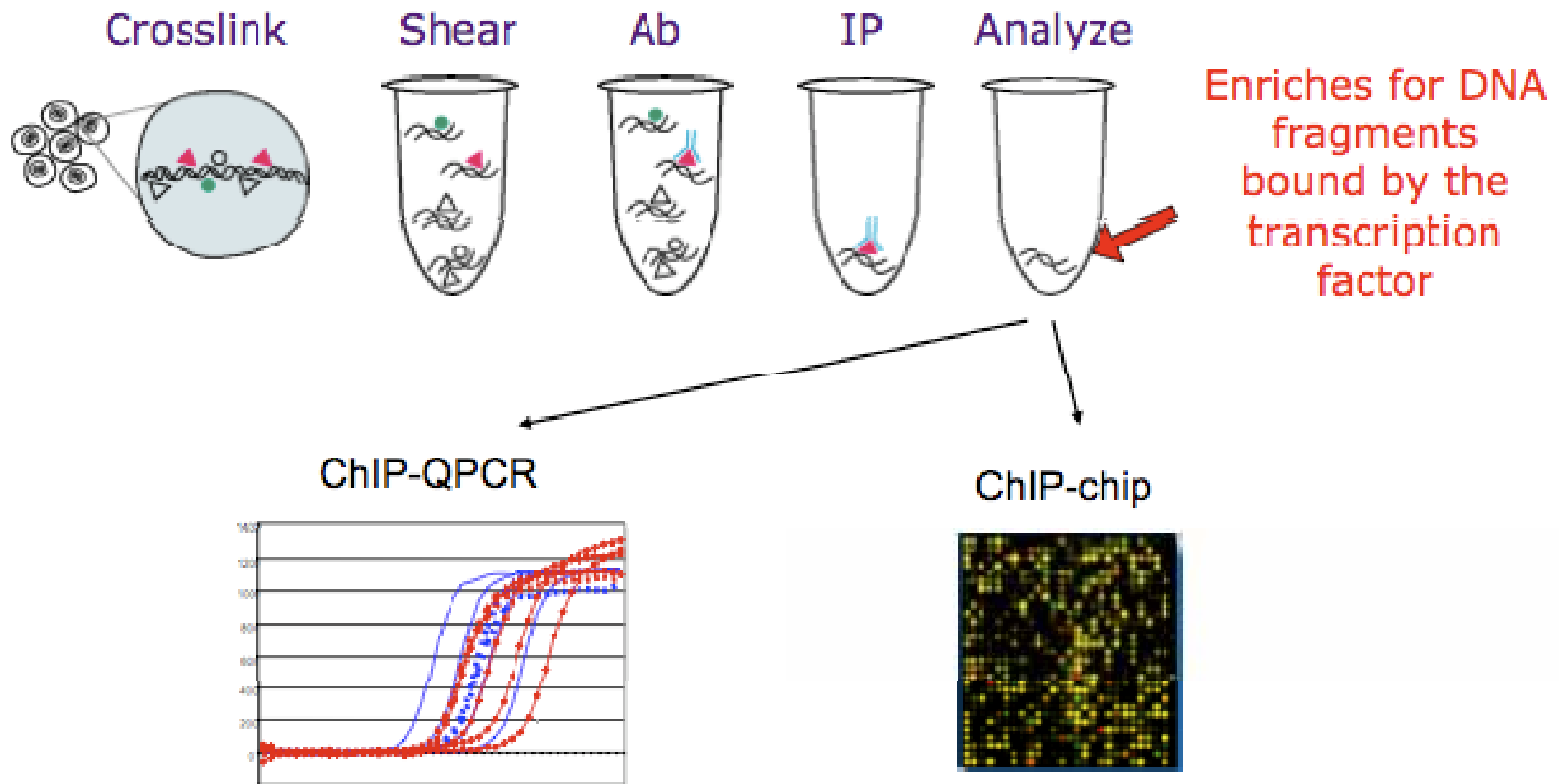
Add unlabeled nucleotides and enzyme to initiate solid-phase bridge amplification.

6. COMPLETE AMPLIFICATION

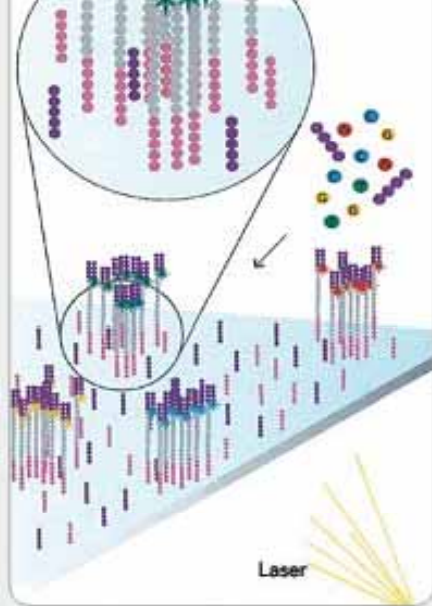


Several million dense clusters of double-

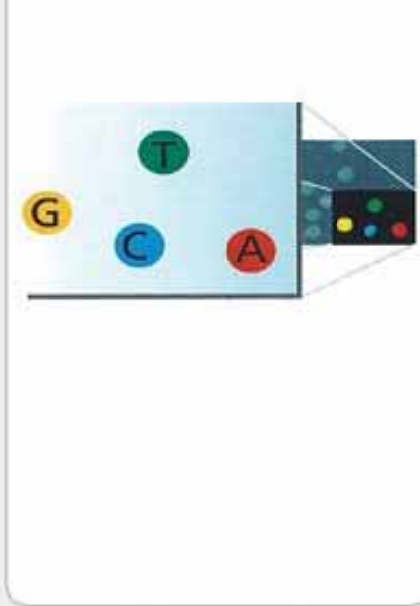
Transcription factor binding site assays (ChIP)



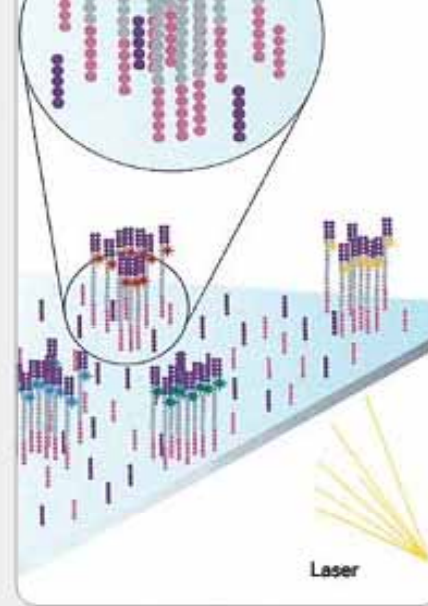
Resolution is about ~500bp



First chemistry cycle: to initiate the first sequencing cycle, add all four labeled reversible terminators, primers and DNA polymerase enzyme to the flow cell.

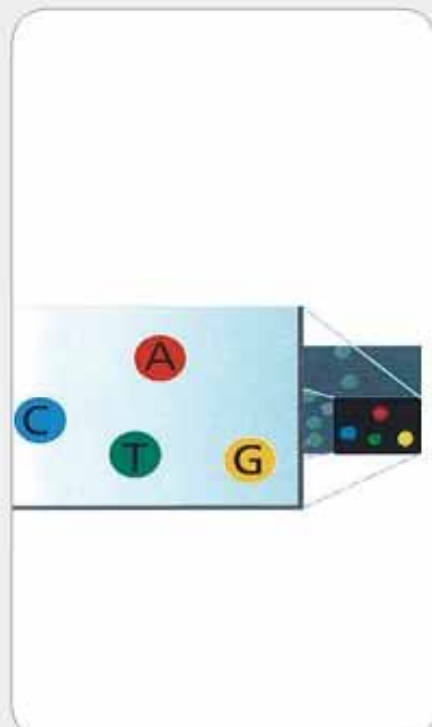


After laser excitation, capture the image of emitted fluorescence from each cluster on the flow cell. Record the identity of the first base for each cluster.

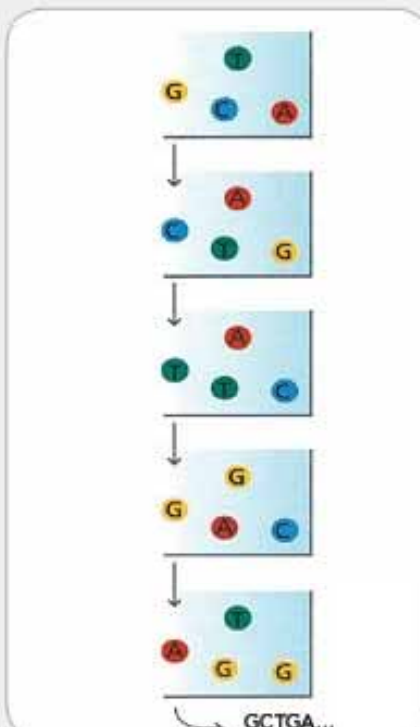


Second chemistry cycle: to initiate the next sequencing cycle, add all four labeled reversible terminators and enzyme to the flow cell.

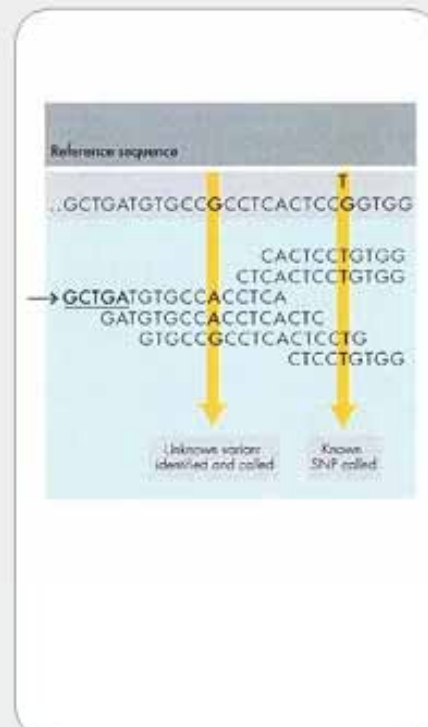
10. IMAGE SECOND CHEMISTRY CYCLE



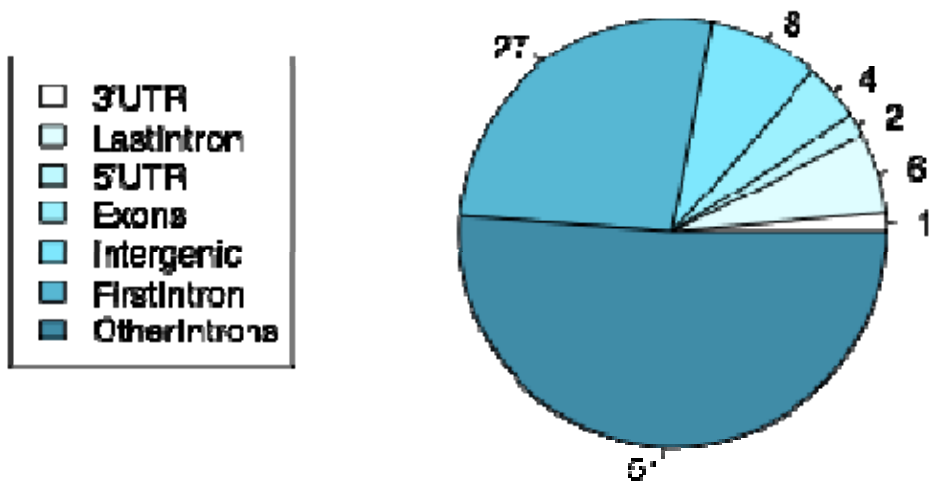
11. SEQUENCE READS OVER MULTIPLE CHEMISTRY CYCLES



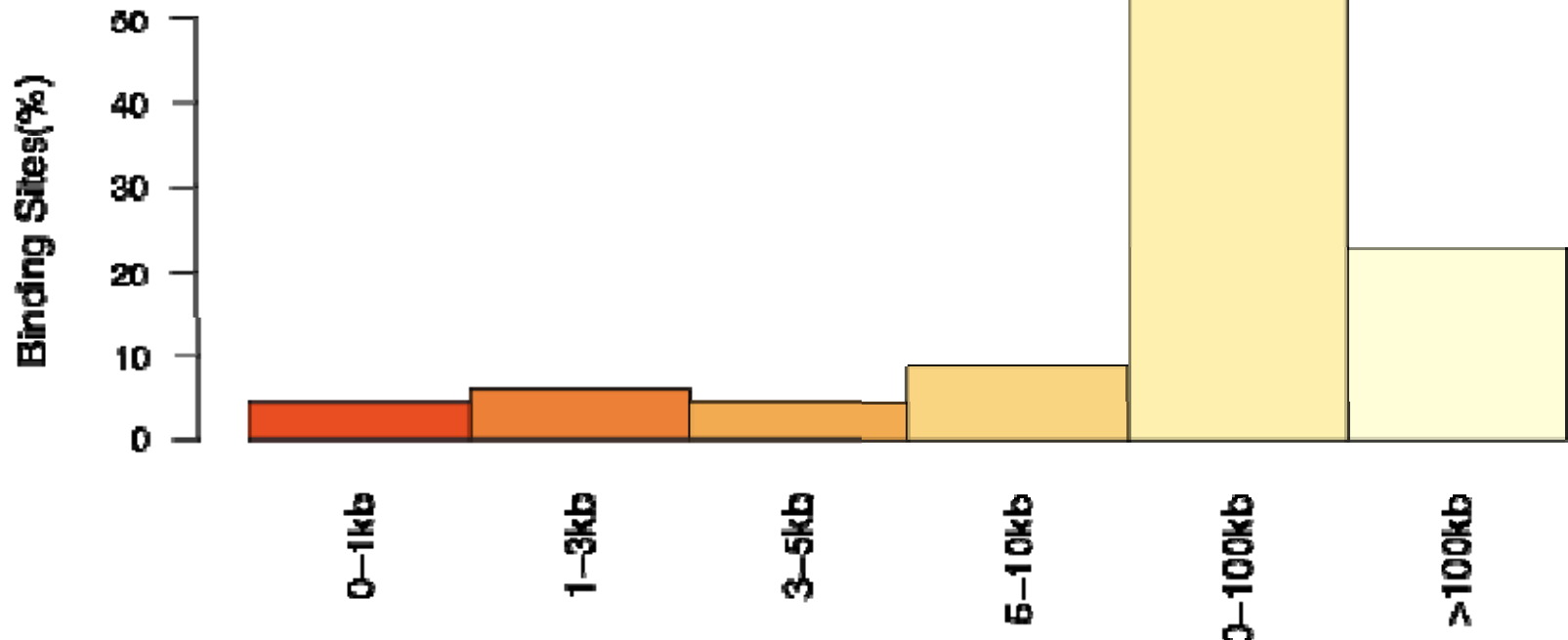
12. ALIGN DATA



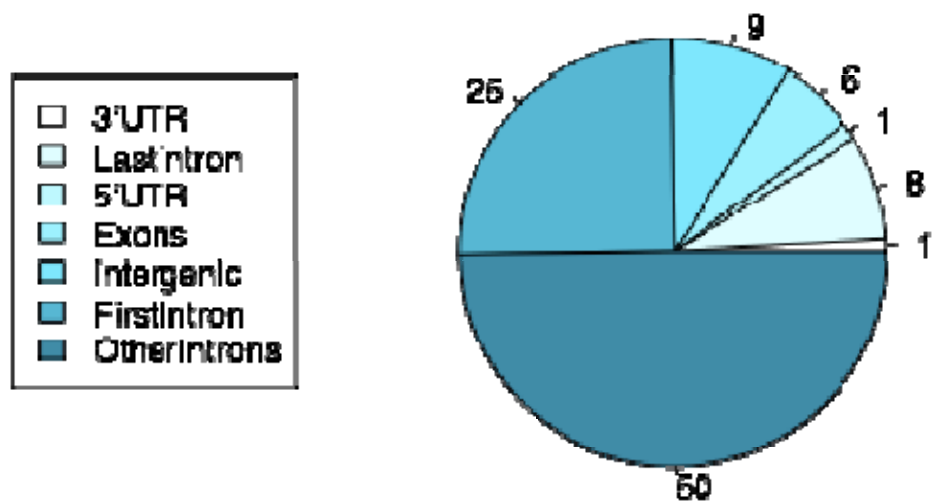
% Peaks overlapping gene features



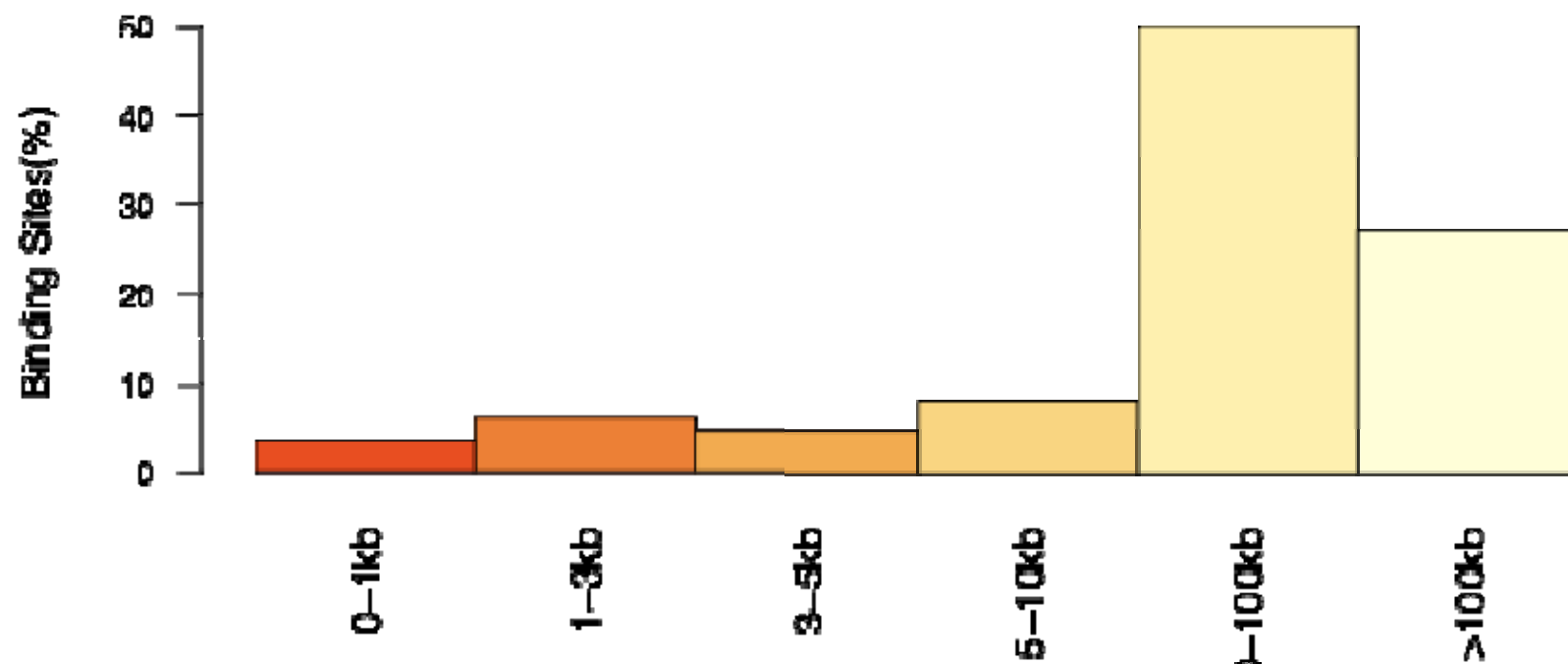
Distance to nearest downstream gene



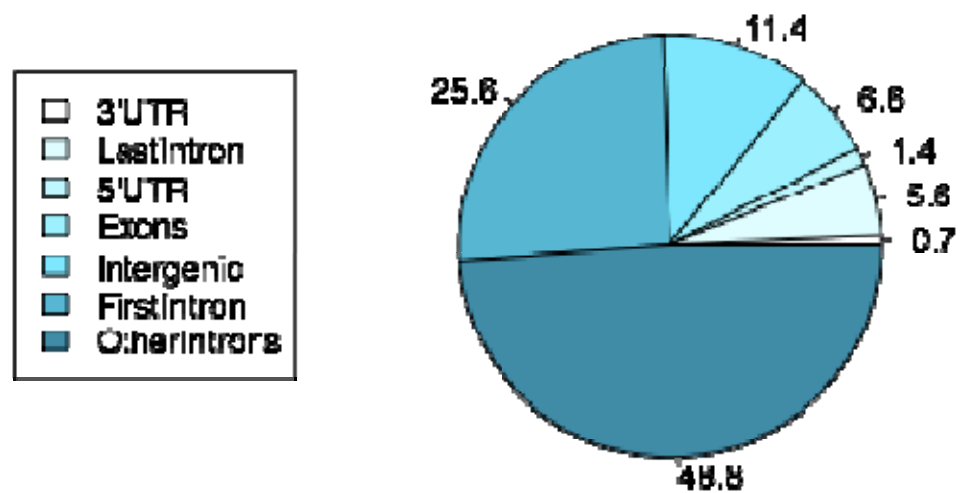
% Peaks overlapping gene features



Distance to nearest downstream gene



% Peaks overlapping gene features



Distance to nearest downstream gene

