



Oocyte polarity: a sign of oocyte quality?

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DISCLOSURE

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Presentation outline

1. Oocyte quality: impact, acquisition and approaches
2. Oocyte polarity: a feature of oocyte maturation
3. Oocyte polarity: a reflection of oocyte quality?
4. Future challenges

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Oocyte Quality: a major determinant of **Embryo Quality** and **Reproductive Success**

% Transfers that resulted in Live Births for ART Cycles using Fresh Embryos, from Own and Donor Eggs, by Woman's Age

Age (years)	Donor eggs (%)	Own eggs (%)
24	55	45
26	65	50
28	60	55
30	60	50
32	60	45
34	55	40
36	60	35
38	60	30
40	55	25
42	55	20
44	50	15
46	50	10
48	45	5

Oocyte Quality is an oocyte attribute of major relevance in the efficiency of the reproductive process, which strongly decreases with age!

2008 Assisted Reproductive Technology Report
Centers for Disease Control and Prevention, USA Department of Health
American Society for Reproductive Medicine
Society for Assisted Reproductive Technology
Dec 2010

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During **Oogenesis** the oocyte acquires **critical functionalities** for initial **Development**

Growth

Maturation

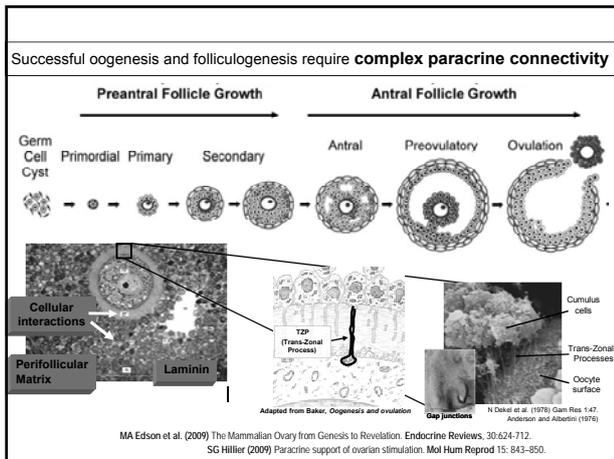
◆ **Oocyte growth phase:**

- Oocyte diameter increases
- High transcriptional and translational activity
- Accumulation of RNA / proteins
- Building of new structures (zona pellucida, cortical granules)

◆ **Oocyte maturation phase:**

- Nuclear / cytoplasmic events with resumption of meiosis and arrest at M_{II} shortly before ovulation
- Organelle redistribution
- Cell polarity and asymmetric division

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Oocyte Quality can be viewed as the acquisition of a series of competences during oogenesis

- 1. Meiotic**
 - a. competence to reach the metaphase II arrest
 - b. competence to allow correct meiotic chromosome segregation
- 2. Activation**

competence to fuse with sperm, finish meiosis, block polyspermy, and form pronuclei
- 3. Developmental**

competence to trigger and support embryonic development

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Oocyte Quality is a very complex attribute !

- decreases strongly with age !
- ... and ...
- depends on the female clinical situation !
- varies within the same egg cohort !
- Can be affected by culture conditions !

However, we infer Oocyte Quality a posteriori

Can we learn to predict Oocyte Quality ?

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1. Learn from the oocyte / follicle that produced a viable embryo

Ongoing implantation

Individual culture - follow-up : find markers

<p>METABOLIC MARKERS : Production / Consumption (H. Leese)</p> <p>SECRETOME (M. Katz-Jaffe)</p> <p>CHROMOSOMAL ANALYSIS PFS / PGD</p>	<p>CUMULUS CELL FUNCTION Gene Expression Metabolic Function</p> <p>OOCYTE (Invasive) Fluorescence Microscopy</p> <p>OOCYTE (Non-Invasive) PoleScope</p>	<p>BIOMARKERS Serum / Follicle Fluid</p> <p>ULTRASOUND Folic. Vol. Increase</p> <p>DOPPLER Peri-follicular Vascular Function</p>
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Live healthy birth

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2. Study and manipulate oogenesis in experimental models

- early stages -

Oogonia cluster breakdown and Primordial follicle formation

Preantral to antral follicle transition

Rodrigues et al. (2008) Oogenesis: prospects and challenges for the future. *J Cell Physiol* 216:355-365.

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2. Study and manipulate oogenesis in experimental models

- late stages -

- ovarian follicle development -

primordial → preantral → antral

Time

- weeks, months -

↑

oocyte maturation

- hours -

Ovulation, Oocyte pick-up

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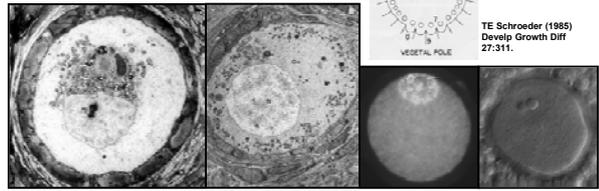
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**Immature oocytes exhibit polarity
Eccentric GV specifies animal pole**



TE Schroeder (1985)
Develop Growth Diff
27:311.



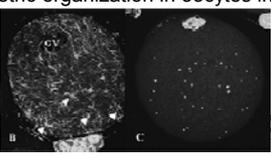
Human primary oocyte in primordial follicle **Rabbit** primary oocyte in primordial follicle **Hamster** and **Mouse** full grown primary oocytes from antral follicles

“All animal eggs have a polar structure” (Raven, 1961)

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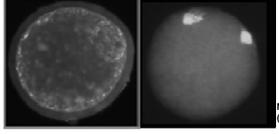
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Asymmetric organization in oocytes intensifies with maturation



mouse

DF Albertini, SL Barrett (2004) The developmental origins of mammalian oocyte polarity. *Sem Cell Dev Biol* 15:599.



hamster

Mafalda Rato, CE Plancha (2005) Personal results

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The second meiotic spindle seems cortically located from the beginning

Wang et al. (2011) Mechanism of the chromosome-induced polar body extrusion in mouse eggs. Cell Division 6:17.

Oocyte polarity ensures asymmetrical division of cytoplasmic content between the gamete and the polar bodies in oocyte maturation and activation

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- Oocyte IVM was observed about 50 years before clinical IVF has started.

SPONTANEOUS MATURATION (rabbit)

Oocytes spontaneously resume meiosis after removal from the follicular environment

Pincus and Enzmann (1935) Comparative behaviour of mammalian eggs *in vitro* and *in vivo*. J Exp Med 62:655.

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IVM oocytes exhibit less fertilization and developmental competence than *In Vivo* matured ones (IVO)

Schroeder and Eppig 1988; Eppig and O'Brien 1998; Cha and Chian 1998; Mermillod *et al.* 1999; Moor *et al.* 2001; Trounson *et al.* 2001

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Strategies to find morphological evidence to explain functional differences between IVM and IVO oocytes

γ-tubulin / pericentrin restricted to pointed MII spindle poles in IVO oocytes

Area/Volume and Pole Width of MII meiotic spindle increase in IVM oocytes

Ibanez *et al.* (2005) Genetic strain variations in the metaphase-II phenotype of mouse oocytes matured *in vivo* or *in vitro*. *Reproduction* 130:845-55.

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Area (Volume) of MII meiotic spindle is higher in non-gonadotropin supplemented IVM

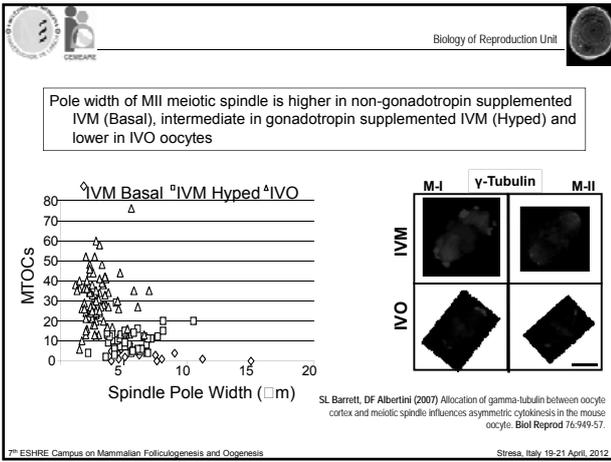
Spindle Area
IVM > IVO, IVO-N

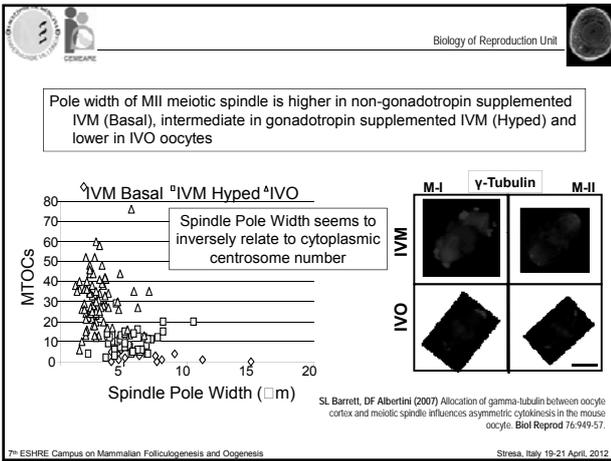
Quantification of Spindle Size and Shape

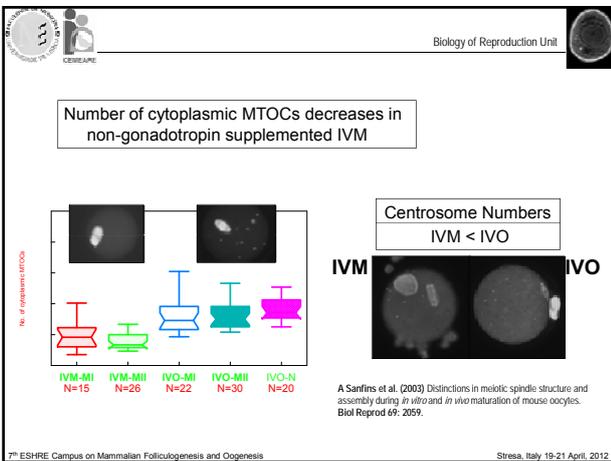
$$\text{Area of spindle} = \left(\frac{h/2(a+c)}{2} \right) + \left(\frac{h/2(b+c)}{2} \right)$$

A Sanfins *et al.* (2003) Distinctions in meiotic spindle structure and assembly during *in vitro* and *in vivo* maturation of mouse oocytes. *Biol Reprod* 69: 2059.

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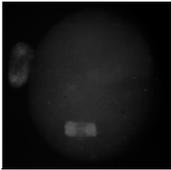




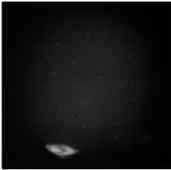
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Spindle anchoring at the cortex is partially lost during non-gonadotropin supplemented IVM

IVM



IVO



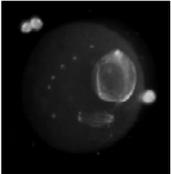
CE Plancha et al (2005) Cell polarity during folliculogenesis and oogenesis. RBM online 10:478-484.

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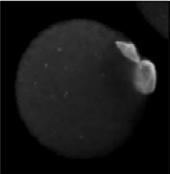
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Retention of MTOCs and oocyte volume is partially lost during non-gonadotropin supplemented IVM

IVM



IVO



Polar Bodies formed *in vitro* contain MTOCs

SL Barrett, DF Albertini (2007) Allocation of gamma-tubulin between oocyte cortex and meiotic spindle influences asymmetric cytokinesis in the mouse oocyte. Biol Reprod 76:949-57.

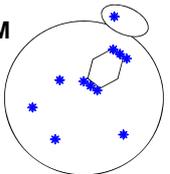
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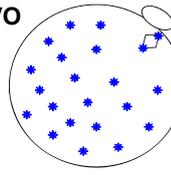
Morphological signs of oocyte polarity and asymmetric division partially lost in mouse non-gonadotropin supplemented IVM

Working Model

IVM



IVO



A Santoro et al (2003) Distinctions in meiotic spindle structure and assembly during *in vitro* and *in vivo* maturation of mouse oocytes. Biol Reprod 69: 2059.

CE Plancha et al (2005) Cell polarity during folliculogenesis and oogenesis. RBM online 10:478-484.

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Morphological signs of oocyte polarity and asymmetric division partially lost in mouse non-gonadotropin supplemented IVM

Working Model

IVM IVO

Are these morphological differences correlated with the described decrease in quality in IVM oocytes ?

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Fertilization rate: presence of 2-PN 6h post-IVF

Mouse

Group	Fertilization Rate (%)
IVO (n=227)	81%
IVM (n=246)	48%
IVM-Rosco (n=270)	41%

Day 1 - PN Check

CE Plancha et al. (2005) Developmental competence of differently matured mouse oocytes. Poster at XXI ESHRE Annual Meeting, Copenhagen, Denmark

Evidence of fertilization competence loss !

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Blastomere numbers at the blastocyst stage

Mouse

Group	Median Blastomere Number
IVO-N	~85
IVO	~85
IVM	~65
IVM-Rosco	~75

ICM TE

ICM ? TE ?

Intermediate

Evidence of developmental competence loss!

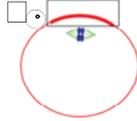
CE Plancha et al. (2005) Developmental competence of differently matured mouse oocytes. Poster at XXI ESHRE Annual Meeting, Copenhagen, Denmark

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Oocyte polarity

1. Is an important feature of oocyte maturation
2. May reflect oocyte quality, since partial loss of morphological signs of polarity correlate with a decrease in fertilization and developmental competences



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To assess if manipulating polarity in mouse oocyte maturation will reflect upon the fertilization and developmental competence levels

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This means that even a very small oocyte, in the limit of being a very large first polar body, is able to be fertilized and initiate embryo development.



Fertilization of each cell



2nd polar body extrusion from each cell

J Otsuki *et al.* (2012) Symmetrical division of mouse oocytes during meiotic maturation can lead to the development of twin embryos that amalgamate to form a chimeric hermaphrodite. *Hum Repr* 27(2):380-387.

- Asymmetry of the first meiotic division does not seem essential for embryo development.
- Asymmetry of the second meiotic division, however, was maintained.

Remaining questions :

- was there a decrease in fertilization and developmental rates?
- had zygotes been separated to develop independently, what would these rates be?
- what would happen in case we prevent asymmetry of second meiotic division?

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To assess if manipulating polarity in mouse oocyte maturation will reflect upon the fertilization and developmental competence levels

To probe other models besides the mouse

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In humans detectable signs of oocyte polarity loss do not seem to correlate with decrease in fertilization and developmental competence

L De Santis *et al.* (2005) Polar body morphology and spindle imaging as predictors of oocyte quality. *RBM online* 11:36-42.

Existing non-invasive approaches may not be enough to accurately discriminate different categories of oocytes

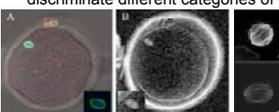


Table 2

Comparison of Polyscope and confocal microscopy measurements of the spindle longitudinal axis in categories A and B, i.e., showing a clear bipolar configuration. Confocal microscopy measurements between groups A and B were also compared separately in fresh and frozen-thawed oocytes. Orange numbers are in parentheses.

	Fresh		Frozen-thawed	
	Polyscope	Confocal	Polyscope	Confocal
A	10.0 ± 2.1 (38) ^a	10.0 ± 2.8 (38) ^a	8.0 ± 1.0 (20) ^a	10.0 ± 1.8 (20) ^a
B	10.7 ± 2.2 (20) ^a	10.3 ± 2.8 (20) ^a	9.1 ± 2.2 (20) ^a	14.0 ± 3.3 (20) ^a

^ap < .001.

Reprints: <http://www.reprints.org/journal/2005/11/36-42>

© Colicchia *et al.* (2010) Comparative analysis of the metaphase II spindle of human oocytes through polarized light and high performance confocal microscopy. *Fertil Steril* 93:2056–2064.

Pre- and post-ovulatory ageing associate with a loss of oocyte polarity

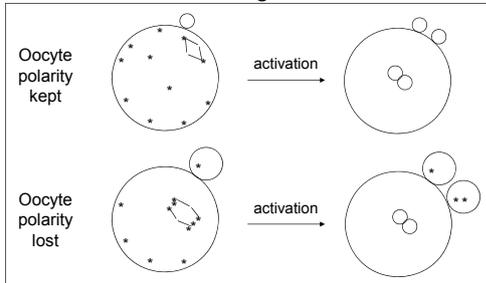
S Brunet MH Verthac (2011) Positioning to get out of meiosis: the asymmetry of division. *Hum Repr Update* 17:68–75.

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Open questions :

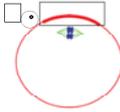
- what are the consequences of preventing asymmetry in the second meiotic division?
- should we focus our attention on the second polar body, besides the spindle?
- will more powerful non-invasive morphological approaches allow us to better associate oocyte polarity and quality?
- can we identify specific human situations or treatments where assessing oocyte polarity will reveal more important than in others?

New working model



Take-home messages

1. Oocyte quality is a relevant property acquired during oogenesis that has remained elusive to predictive studies
2. Oocyte polarity is an important feature of oocyte maturation
3. Oocyte polarity may reflect oocyte quality, since partial loss of morphological signs of polarity correlate with a decrease in fertilization and developmental competences
4. Future studies are needed to assess the value of oocyte polarity as a sign of oocyte quality



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- | | |
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- | | | |
|--------------------|-------------|------------|
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Oocyte polarity: a sign of oocyte quality?

Probably yes, but it is still not clear how ...

Thank You !
