



Organised by the ESHRE Special Interest Group "Stem Cells"
in collaboration with the Spanish Stem Cell Bank (Centre de Medicina
Regenerativa de Barcelona, Centro de Investigación Príncipe Felipe -
Valencia and Centro de Investigaciones Biomédicas - Granada)

Basic course on "Update on pluripotent
stem cells (hESC and iPS)
Hands on course on "Derivation and culture
of pluripotent stem cells"

ESHRE Campus 2010
Valencia Spain, 8-12 November 2010

Cell reprogramming (induced pluripotent stem cells, iPS)

Angel Raya
ICREA Research Professor
Institute for Bioengineering of Catalonia
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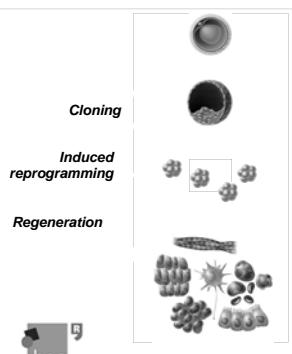
November 8, 2010



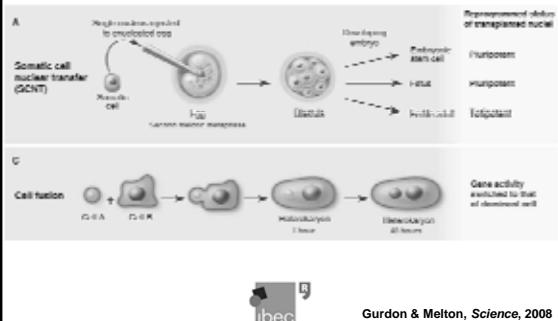
The progressive loss of potency

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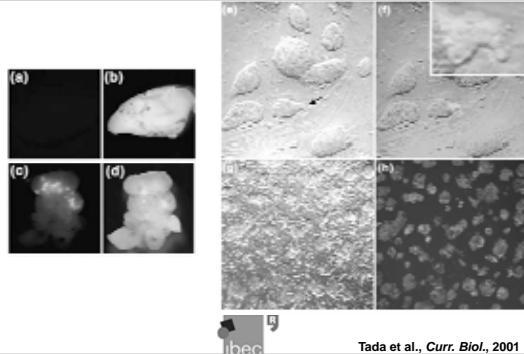
POTENCY



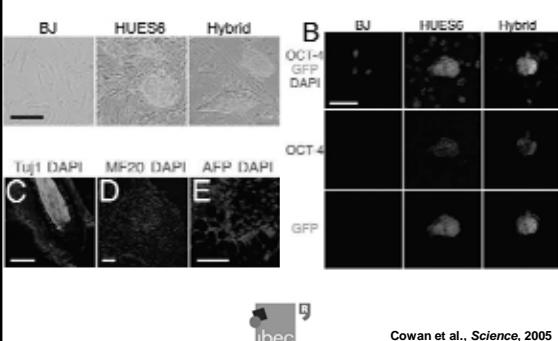
Reprogramming to pluripotency



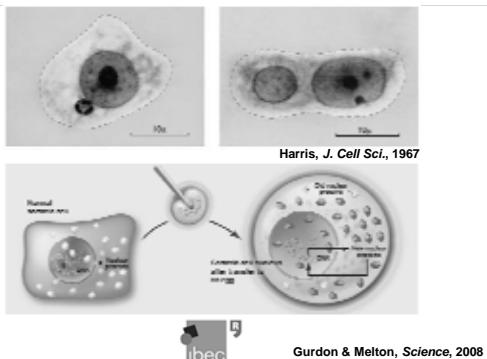
Reprogramming through cell fusion



Reprogramming through cell fusion



Reprogramming through cell fusion



Induced reprogramming of mouse cells

Oct3/4 Ecat1 β -catenin

Sox2 Ecat8 Klf4

Nanog Dppa2 c-Myc

Dppa3 Stat3

Dppa4 E-Ras

Dppa5

Dnmt3l

Fbx15

Gdf3

Sox15

Fth1l7

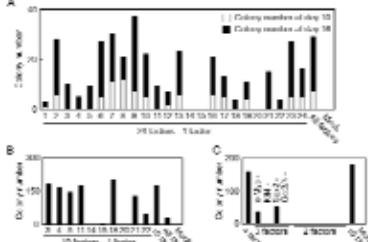
Sall4

Rex1

Utf1

Grb2

Tcl1



Takahashi & Yamanaka, *Cell*, 2006

Induced reprogramming of mouse cells

Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors

Kousaku Okita¹ and Shinya Yamanaka^{1,2*}

¹Cell Type Control Research Center, RIKEN, Saitama 351-0198, Japan; ²Graduate School of Medicine, Kyoto University, Kyoto 606-8501, Japan

ARTICLES

Generation of germline-competent induced pluripotent stem cells

Kousaku Okita¹, Tomoko Ichisaka^{1,2} & Shinya Yamanaka^{1,2*}



Induced reprogramming of human cells

Induction of Pluripotent Stem Cells from Adult Human Fibroblasts by Defined Factors

Cell

Kazutoshi Takahashi,¹ Jun Takagi,² Miki Ohnuki,¹ Megumi Haga,^{1,3} Tomoko Okuda,^{1,3} Kunihiro Tomoda,¹

and Atsushi Yamada^{1,3}

Scienceexpress

Report

Induced Pluripotent Stem Cell Lines Derived from Human Somatic Cells

Jiayoung Yu,^{1,2} Masayo A. Yabuuchi,² Kuni Saito, Ota,^{1,2} Ayako Andou, Shiro Taniguchi,^{1,2} Shinobu T. Ito,^{1,2} Keiko Ito,^{1,2} Shunji Imai,³ Bell Non,³ Yoshimasa A. Andou,^{1,2} Yukio Kubota,¹ Kozo Stewart,¹ Iggy J. Shinkura,^{1,2} James A. Thomson,^{1,2,*} &

Vol 45(1) January 2006 doi:10.1126/science.112634

science

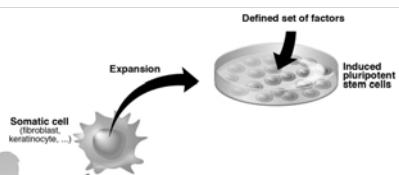
Reprogramming of human somatic cells to pluripotency with defined factors

In-Hyun Park¹, Rui Zhao¹, Jason A. West¹, Akiko Yabuuchi¹, Hongguang Huo¹, Tan A. Ince¹, Paul H. Leroi¹,

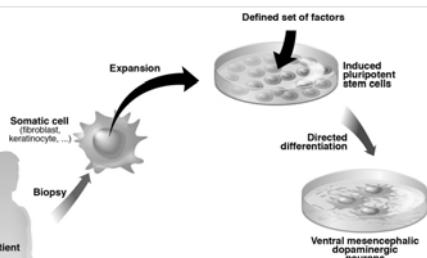
M. William Lentsch¹ & George Q. Daley¹



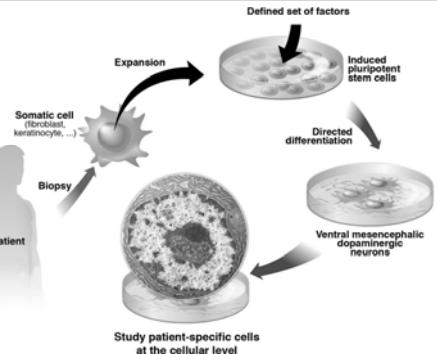
Modeling human disease with iPS cells



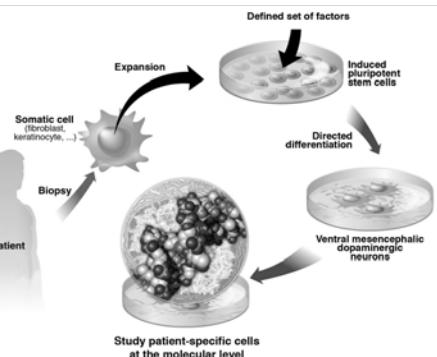
Modeling human disease with iPS cells



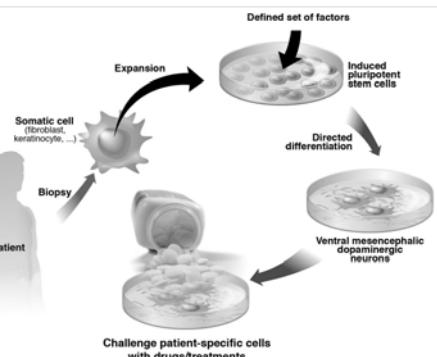
Modeling human disease with iPS cells



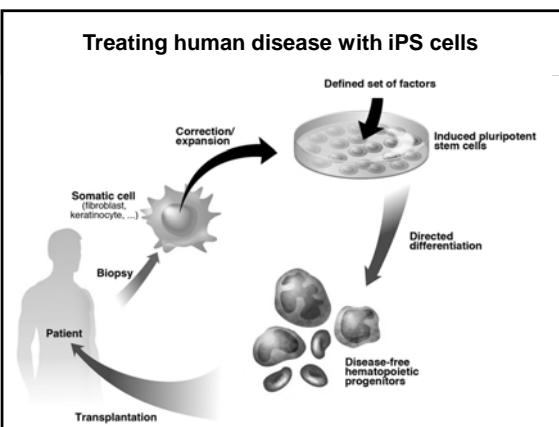
Modeling human disease with iPS cells



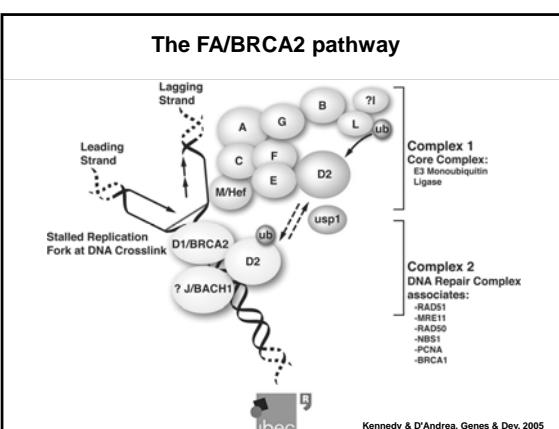
Modeling human disease with iPS cells



Treating human disease with iPS cells



The FA/BRCA2 pathway



Generation of FA patient-specific iPS cells

Juan Bueren
CIEMAT

	Promoters	Transgene	ID	Source	Passages
RV	viral LTR	FANCA	FA5	Skin	High
		FANCA-EGFP			
		FANCA-Neo			
LV	internal promoters: SFFV, CMV, PGK, VAV	for FANCA	FA90	Skin	High
		FANCA			
		FANCA-Wpre			
RV	viral LTR	FANCD2	FA430	Skin	Mid
		FANCD2-EGFP			
		FANCD2-Neo			
LV	internal promoters: VAV	FANCD2-Wpre	FA431	Skin	Low

Raya et al., Nature 2009

Generation of FA patient-specific iPS cells

Patient ID	FA group	Somatic cell	Attempts	iPS-like colonies	Lines generated	Lines characterized	Markers	In vitro	Terat.
FAS	A	Fibr.	5	0	0	NA	NA	NA	NA
FAS	A	cFibr.	5	0	0	NA	NA	NA	NA
FA90	A	Fibr.	3	0	0	NA	NA	NA	NA
FA90	A	cFibr.	3	~37	10	5	5	3	cFA90-44-X
FA153	A	Fibr.	5	0	0	NA	NA	NA	NA
FA153	A	cFibr.	5	0	0	NA	NA	NA	NA
FA404	A	Fibr.	3	0	0	NA	NA	NA	NA
FA404	A	cFibr.	3	~30	2	2	2	2	cFA404-FIPS4FX
FA404	A	Kerat.	3	0	0	NA	NA	NA	NA
FA404	A	cKerat	3	~30	3	3	3	3	cFA404-KIPS4FX
FA430	D2	Fibr.	6	0	0	NA	NA	NA	NA
FA430	D2	cFibr.	6	0	0	NA	NA	NA	NA
FA431	D2	Fibr.	3	~10	2*	2*	NA	NA	
FA431	D2	cFibr.	3	~10	2	2	2	2	NT
									cFA431-44-X



Raya et al., Nature 2009

Criteria for defining *bona fide* iPS cells

Self-renewal

> 20 passages

Karyotypic stability

Pluripotency

Expression of pluripotency-associated markers

In vitro differentiation

Teratoma formation

Molecular

DNA fingerprinting

Integration of reprogramming transgenes

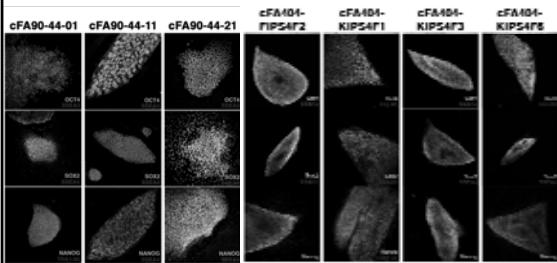
Silencing of reprogramming transgenes

Reprogramming of gene expression profile

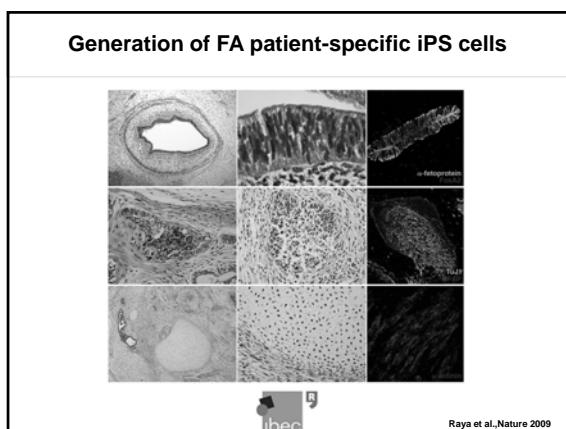
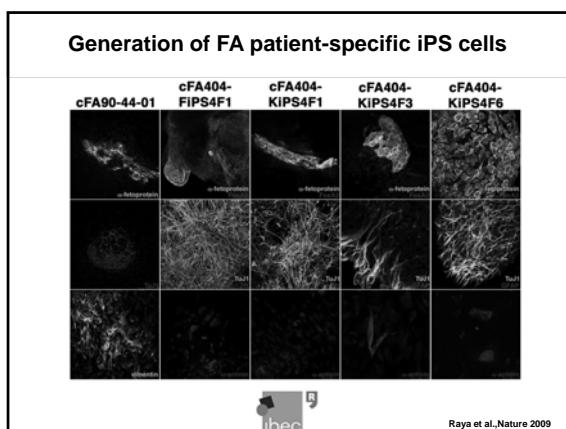
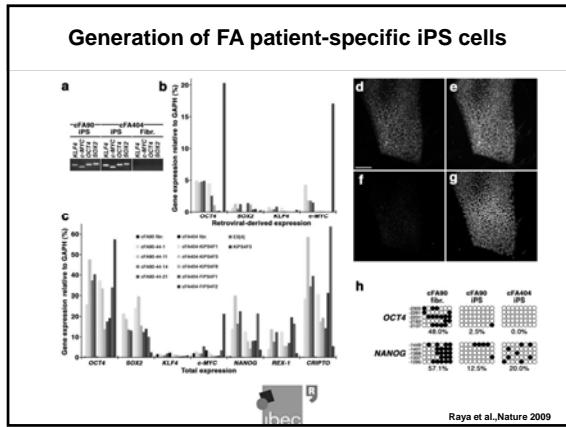
Reprogramming of DNA methylation profile

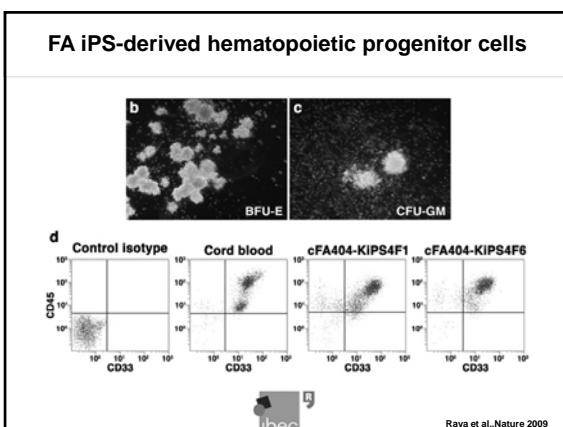
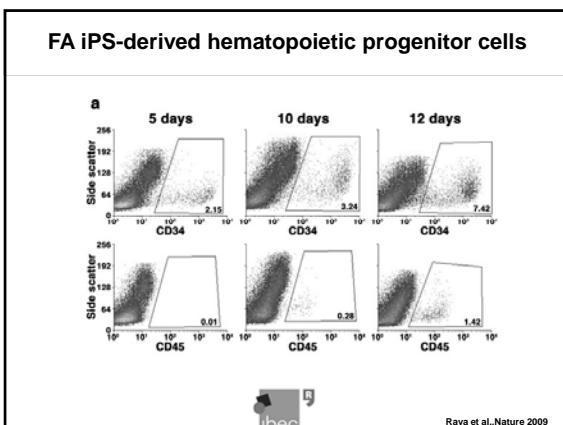
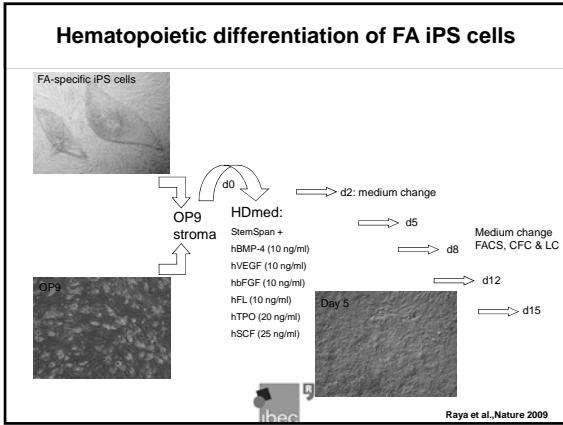


Generation of FA patient-specific iPS cells



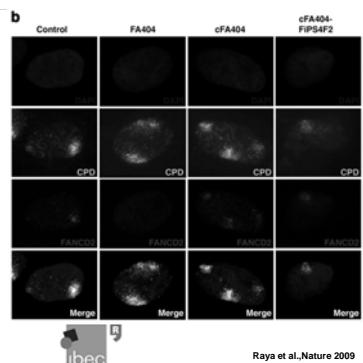
Raya et al., Nature 2009



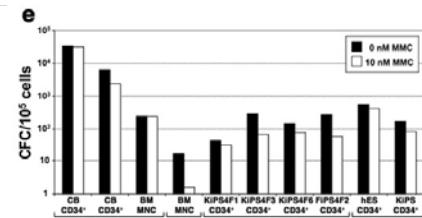


FA iPS-derived cells are disease free

Jordi Surrallés
UAB



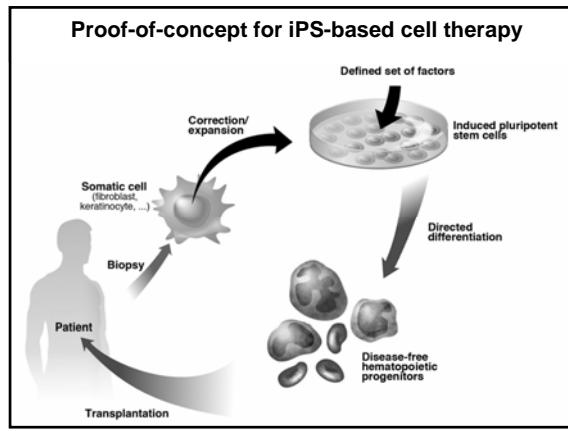
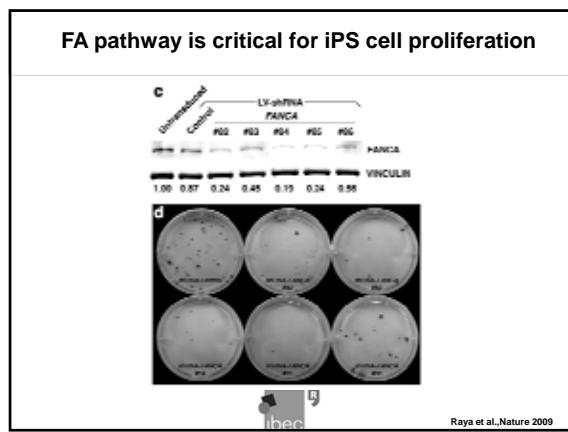
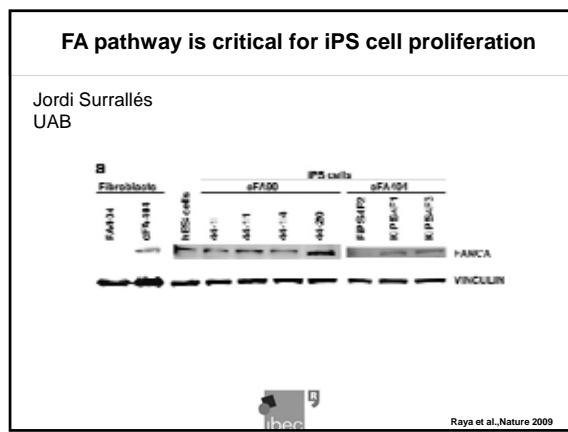
Disease-free hematopoietic progenitors



Generation of FA patient-specific iPS cells

Patient ID	FA group	Somatic cell	Attempts	iPS-like colonies	Lines generated	Markers	In vitro	Terat.
FA5	A	Fibr.	5	0	0	NA	NA	NA
FA5	A	cFibr.	5	0	0	NA	NA	NA
FA90	A	Fibr.	3	0	0	NA	NA	NA
FA90	A	cFibr.	3	~37	10	5	5	3
					cFA90-44-X			
FA153	A	Fibr.	5	0	0	NA	NA	NA
FA153	A	cFibr.	5	0	0	NA	NA	NA
FA404	A	Fibr.	3	0	0	NA	NA	NA
FA404	A	cFibr.	3	~30	2	2	2	2
					cFA404-FIP54FX			
FA404	A	Kerat.	3	0	0	NA	NA	NA
FA404	A	cKerat	3	~30	3	3	3	3
					cFA404-KIP54FX			
FA430	D2	Fibr.	6	0	0	NA	NA	NA
FA430	D2	cFibr.	6	0	0	NA	NA	NA
FA431	D2	Fibr.	3	~10	2*	2*	NA	NA
FA431	D2	cFibr.	3	~10	2	2	2	NT
					cFA431-44-X			

Raya et al., Nature 2009



Shortcomings of iPS cell therapy

Derivation of human iPS cells
Retroviral integrations
Clinical-grade lines

Control of cell proliferation
Selective killing
Understanding self-renewal

Specific differentiation protocols
Directed differentiation protocols
Clinical-grade protocols



Acknowledgements

Angel Raya
Ignasi Rodríguez-Pizà
Rita Vassena
María José Barrero
Antonella Consiglio
Eduard Sleep
Federico González
Gustavo Tiscornia
Elena Garreta
Trond Aasen
Anna Veiga
Juan Carlos Izpisúa Belmonte
CMRB



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Inder Verma
Salk Institute